

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization  
International Bureau



(43) International Publication Date  
20 September 2001 (20.09.2001)

PCT

(10) International Publication Number  
WO 01/68870 A2

(51) International Patent Classification<sup>7</sup>: C12N 15/54, 9/10, 5/10, C07K 16/40, C12Q 1/68, G01N 33/53, A01H 5/00, C12N 15/82

(74) Agent: MOORE, Mark, D.; Williams, Morgan & Amer-  
son, P.C., Suite 250, 7676 Hillmont, Houston, TX 77040  
(US).

(21) International Application Number: PCT/US01/08352

(81) Designated States (national): AE, AG, AL, AM, AT, AU,  
AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ,  
DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR,  
HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR,  
LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ,  
NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM,  
TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.

(22) International Filing Date: 15 March 2001 (15.03.2001)

(84) Designated States (regional): ARIPO patent (GH, GM,  
KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian  
patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European  
patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE,  
IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF,  
CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

(25) Filing Language: English

Published:

(26) Publication Language: English

— without international search report and to be republished  
upon receipt of that report

(30) Priority Data:  
09/525,885 15 March 2000 (15.03.2000) US

For two-letter codes and other abbreviations, refer to the "Guid-  
ance Notes on Codes and Abbreviations" appearing at the begin-  
ning of each regular issue of the PCT Gazette.

(71) Applicants: UNIVERSITY OF FLORIDA [US/US];  
P.O. Box 115500, Gainesville, FL 32611-5500 (US).  
CARNEGIE MELLON UNIVERSITY [US/US]; Suite  
302, 4615 Forbes Avenue, Pittsburgh, PA 15213 (US).

(72) Inventors: HANSON, Andrew, D.; 507 NW 39th Road  
#163, Gainesville, FL 32607 (US). NUCCIO, Michael, L.;  
P.O. Box 1915, Melrose, FL 32666 (US). HENRY, Susan,  
A.; 4400 Fifth Avenue, Pittsburgh, PA 15213 (US).



WO 01/68870 A2

(54) Title: S-ADENOSYL-L-METHIONINE:PHOSPHOETHANOLAMINE N-METHYLTRANSFERASE COMPOSITIONS  
AND METHODS FOR MODULATING LIPID BIOSYNTHESIS IN PLANTS

(57) Abstract: Disclosed are compositions and methods for increasing the nutritional value of plants and plant parts. In illustrative embodiments PEAMT and  $\Delta$  PEAMT polynucleotide and polypeptide compositions are disclosed as well as their use in modulating the levels of lipid compounds, and particularly, regulating the level of phosphatidylcholine, and its precursors in plants and seeds derived therefrom. Also disclosed are methods for modulating the level of glycine betaine and choline-*O*-sulfate in cells, and increasing the tolerance of transformed plants to osmotic and cryogenic stress.

**DESCRIPTION****S-ADENOSYL-L-METHIONINE:PHOSPHOETHANOLAMINE N-METHYLTRANSFERASE  
COMPOSITIONS AND METHODS FOR MODULATING LIPID BIOSYNTHESIS IN PLANTS**

5

**1. BACKGROUND OF THE INVENTION**

The present application claims priority to U. S. Patent Application Serial No. 09/525,885, filed March 15, 2000; the entire specification, claims and figures of which is specifically incorporated herein by reference in its entirety without disclaimer. The United 10 States government has certain rights in the present invention pursuant to Grant number 98-35100-6149 from the United States Department of Agriculture, and Grant number GM19629 from the National Institutes of Health.

**1.1 FIELD OF THE INVENTION**

15 The present invention relates to the genetic manipulation of plants, particularly to methods and compositions for altering the lipid content in plants, plant tissues, and seeds derived therefrom. Disclosed are compositions encoding S-adenosyl-L-methionine:phosphoethanolamine N-methyltransferase (PEAMT) peptides and polypeptides, as well as polynucleotides encoding these peptides and proteins, antibodies and antigen binding 20 fragments immunospecific for such compositions, and methods employing PEAMT compositions in the preparation of transgenic plants, and in altering lipid biosynthesis in plants and plant cells.

**1.2 DESCRIPTION OF RELATED ART**

25 Flowering plants are unusual in how they synthesize choline (Cho) moieties. In leaves and other vegetative tissues, the first and committing step is *N*-methylation of phosphoethanolamine (P-EA) to give phosphomonomethylethanolamine (P-MME), and the subsequent *N*-methylations occur at the phosphobase level, the phosphatidyl base level, or both, depending on the species (Datko and Mudd, 1988a; 1988b; Rhodes and Hanson, 1993) 30 (FIG. 1A). For example, in spinach and sugar beet leaves, essentially all flux through the last

two methylations is at the phosphobase level (Hanson and Rhodes, 1983; Summers and Weretilnyk, 1993), whereas in soybean cells it is at the Ptd-base level (Datko and Mudd, 1988a). In contrast, Cho synthesis in the bacterium *Rhodobacter sphaeroides*, in fungi, and in mammalian liver proceeds solely via the sequential methylation of 5 phosphatidylethanolamine (Ptd-EA) (Vance *et al.*, 1997; Kanipes and Henry, 1997; Arondel *et al.*, 1993). Nerve tissues have a phosphobase methylation route as well as a phosphatidylbase route, but the first methylation appears not to be restricted to the phosphobase level (Mukherjee *et al.*, 1995; Andriamanpanry *et al.*, 1991).

The initial methylation of P-EA in plants is catalyzed by 10 *S*-adenosyl-L-methionine:phosphoethanolamine *N*-methyltransferase (PEAMT), which has been detected in all species tested (Datko and Mudd, 1988b; Summers and Weretilnyk, 1993; Nuccio *et al.*, 1998). PEAMT was recently purified 5,400-fold from spinach leaves, giving a preparation containing several polypeptides (Smith *et al.*, 1999). This preparation catalyzed methylation of P-MME and phosphodimethylethanolamine (P-DME) as well as P-EA, and 15 these activities co-purified in a constant ratio through the three last steps in the procedure (Smith *et al.*, 1999). These data suggest that PEAMT could be trifunctional, but do not rule out a duo or trio of similar *N*-methyltransferases that act on different phosphobases. Pathways involving one, two or three *N*-methyltransferases all have precedents. In *R. sphaeroides* and liver a single Ptd-EA *N*-methyltransferase mediates all three methylations 20 (Vance *et al.*, 1997; Arondel *et al.*, 1993) whereas in *Saccharomyces cerevisiae* and *Schizosaccharomyces pombe* there are two enzymes, one mediating the first methylation of Ptd-EA and another mediating primarily the last two (Kanipes and Henry, 1997). The phosphobase pathway in nerve tissues has three separate *N*-methyltransferases (Mukherjee *et al.*, 1995).

25 Certain plants (*e.g.*, spinach, sugar beet) use large amounts of Cho to produce the osmoprotectant glycine betaine (GlyBet) via the pathway Cho → betaine aldehyde → GlyBet (Rhodes and Hanson, 1993). Because GlyBet accumulation contributes to resistance to salinity and drought stress, there has been much interest in engineering GlyBet synthesis in plants that do not naturally produce it (Nuccio *et al.*, 1999). However, when enzymes for 30 Cho oxidation to GlyBet are expressed in such plants (*e.g.*, tobacco, canola), they accumulate

little GlyBet, apparently in part because their endogenous Cho supply is inadequate (Nuccio *et al.*, 1998; Huang *et al.*, 1999). This has focused attention on the pathway of Cho biosynthesis and its regulation (Nuccio *et al.*, 1999).

Biochemical and physiological evidence shows that the PEAMT-mediated step is a 5 control point in the biosynthesis of Cho moieties, and that at least two mechanisms are involved. One is feedback inhibition: PEAMT activity in crude *Lemna* extracts (Mudd and Datko, 1989a; 1989b) and purified spinach preparations (Smith *et al.*, 1999) is inhibited by P-Cho, and <sup>14</sup>C tracer data for sugar beet leaf tissue indicate that this occurs *in vivo* (Hanson and Rhodes, 1983). Another mechanism may be regulation of PEAMT gene expression. 10 The *de novo* synthesis of Cho in *Lemna*, soybean and carrot cells is suppressed by exogenous Cho, and this suppression is accompanied by a decrease in extractable PEAMT activity (Mudd and Datko, 1989a; 1989b). Conversely, salinization in spinach, which increases the consumption of Cho in GlyBet synthesis, causes an increase in PEAMT activity (Weretilnyk *et al.*, 1995). Direct evidence that flux through the PEAMT step limits the synthesis of Cho 15 moieties *in vivo* comes from tobacco engineered to convert Cho to GlyBet; supplying MME or DME increases the flux to Cho and GlyBet but supplying EA does not (Nuccio *et al.*, 1998).

### 1.3 DEFICIENCIES IN THE PRIOR ART

20 A goal of plant breeding has been to modulate the lipid content in plant seeds, and to alter the levels of various plant oils. Despite the importance of PEAMT as the committing step in the biogenesis of Cho moieties, it has not been cloned or unambiguously characterized with respect to the reaction(s) it catalyzes, nor have the genes for any other plant *N*-methyltransferase participating in Cho biogenesis been cloned.

25

## 2. SUMMARY OF THE INVENTION

The present invention overcomes these and other limitations inherent in the prior art by providing compositions and methods for increasing the nutritional value of plants. Methods are also provided particularly for modulating the levels of cellular intermediates, 30 such as phosphoethanolamine, phosphomonomethylethanolamine, phosphodimethyl-

ethanolamine, phosphocholine, phosphatidyl choline, choline, glycine betaine, and choline-*O*-sulfate in plants, plant tissues, and seeds. Methods are also disclosed for altering the lipid content in transformed plants cells expressing one or more of these polynucleotide constructs, in a transgenic plant, and in the seed, tissues, and cells derived from such a 5 transgenic plant.

Disclosed are novel plant *S*-adenosyl-L-methionine:phosphoethanolamine *N*-methyltransferase (PEAMT) polypeptides, antibodies and antigen binding fragments immunospecific therefor, and polynucleotides that encode these enzymes, and peptides and epitopic or antigenic fragments thereof. Also disclosed are novel amino acid sequences 10 comprising a truncated *S*-adenosyl-L-methionine:phosphoethanolamine *N*-methyltransferase (ΔPEAMT) polypeptide, and polynucleotides encoding these truncated PEAMT variants. Also disclosed are methods of identifying PEAMT- and ΔPEAMT-specific polypeptide, antibody, and polynucleotide compositions, methods for preparing PEAMT and ΔPEAMT recombinant host cells, vectors, virus, and expression constructs, methods for the 15 recombinant production of PEAMT and ΔPEAMT polypeptides, and methods for making transgenic plants that express one or more exogenously provided PEAMT- or ΔPEAMT-specific transgenes. These compositions find particular use in modulating the levels of phospholipids in plant cells, and for altering the levels of compounds including phosphocholine, phosphatidyl choline, and their precursors in plant cells, tissues, fruits and 20 for modulating the oil content of plant seeds.

The invention also provides isolated polynucleotides that: (a) encode at least a first polypeptide having PEAMT or ΔPEAMT activity that comprises an at least 27 amino acid contiguous sequence from SEQ ID NO:2 or SEQ ID NO:4; (b) encode at least a first polypeptide having PEAMT or ΔPEAMT activity that is at least about 75% or greater identical 25 to either the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4; (c) comprise at least a first nucleic acid segment that comprises an at least 26 contiguous nucleotide sequence from SEQ ID NO:1 or SEQ ID NO:3; (d) hybridize to the sequence of SEQ ID NO:1 or SEQ ID NO:3; or (d) hybridize to a nucleic acid sequence of from about position 254 to about position 30 1735 of SEQ ID NO:1 or to the nucleic acid sequence of SEQ ID NO:3, or that hybridize to one or more complements thereof, under moderate to moderately stringent hybridization conditions.

## 2.1 PEAMT PEPTIDES, POLYPEPTIDES AND COMPOSITIONS THEREOF

The present invention provides an isolated peptide or polypeptide that comprises, consists essentially of, or consists of, at least a first sequence region that preferably shares at least about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, or about 80% or higher sequence identity with the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4, which encode PEAMT and  $\Delta$ PEAMT polypeptides from spinach, respectively. Likewise, the present invention provides an isolated peptide or polypeptide that comprises, consists essentially of, or consists of, at least a first sequence region that preferably shares at least about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, or about 90%, or higher sequence identity with the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4. The invention also provides at least a first isolated peptide or polypeptide that comprises, consists essentially of, or consists of, at least a first sequence region that preferably shares at least about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about 99%, or higher sequence identity with the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4. In an illustrative embodiment, preferred PEAMT and  $\Delta$ PEAMT polypeptides include those polypeptides that comprise, consist essentially of, or consist of the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.

Such polypeptides may be used in a variety of embodiments, methods, and uses, and particularly in those methods and uses as described herein. For example, the polypeptides of the invention may be used to catalyze one or more of the biochemical reactions disclosed herein, and/or may also be used in the identification of PEAMT- or  $\Delta$ PEAMT-specific antibodies or antigen binding fragments thereof, or may in fact be used in a method for producing an immune response in an animal, and also in the preparation of antibodies and antigen binding fragments that specifically bind to one or more PEAMT or  $\Delta$ PEAMT peptides or polypeptides, using one or more immunological methods known to those of skill in the art, including those methods as described hereinbelow. The PEAMT- or  $\Delta$ PEAMT-specific antibodies and antigen binding fragments so produced also represent important embodiments of the present invention. Such antibodies may be suitably packaged in an immunodetection kit,

along with an immunodetection reagent, and instructions for using the antibody in methods such as ELISAs and other immunoaffinity methodologies to detect the presence of PEAMT or  $\Delta$ PEAMT peptides or polypeptides in a target sample, such as in a plant cell, a plant part, or a plant extract.

5 Highly preferred polypeptides, peptides and proteins of the invention include those peptides and polypeptides that comprise all, substantially all, or an antigenic fragment from, at least a first plant PEAMT or  $\Delta$ PEAMT polypeptide. Highly preferred PEAMT and  $\Delta$ PEAMT peptides or polypeptides are those that comprise, consist essentially of, or consist of, at least a first sequence region that comprises an at least about 27, an at least about 28, an 10 at least about 29, an at least about 30, an at least about 31, or an at least about 32 or more contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4, and particularly those peptides and polypeptides comprising such a sequence and that possesses PEAMT or  $\Delta$ PEAMT activity when expressed in a suitable plant host cell cultured under the appropriate conditions for such activity. Likewise, PEAMT or  $\Delta$ PEAMT peptides or polypeptides that 15 comprise, consist essentially of, or consist of, at least a first sequence region that comprises an at least about 33, 34, 35, 36, 37, 38, 39, or 40 or more contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4, are also highly preferred in the practice of the present invention, as are those that comprise, consist essentially of, or consist of, at least a first sequence region that comprises an at least about 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50 or 20 more contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4.

As such, exemplary highly-preferred PEAMT or  $\Delta$ PEAMT peptides and polypeptides are those that comprise, consist essentially of, or consist of, at least a first sequence region that comprises an at least about 51, 52, 53, 54, 55, 56, 57, 58, 59 or 60 or more contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4, and that possess PEAMT or 25  $\Delta$ PEAMT activity when expressed in a suitable plant host cell cultured under the appropriate conditions for such enzymatic activity. Likewise, exemplary highly-preferred PEAMT or  $\Delta$ PEAMT peptides and polypeptides are those that comprise, consist essentially of, or consist of, at least a first sequence region that comprises an at least about 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, or 80 or more contiguous amino acid sequence 30 from SEQ ID NO:2 or SEQ ID NO:4, and that possess PEAMT or  $\Delta$ PEAMT activity when

expressed in a suitable plant host cell are also contemplated to be particularly useful in the methods disclosed herein.

Even more highly preferred PEAMT or  $\Delta$ PEAMT peptides and polypeptides are those that comprise, consist essentially of, or consist of, at least a first sequence region that comprises an at least about 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, or 100 or more contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4, and that possess PEAMT or  $\Delta$ PEAMT activity when expressed in a suitable plant host cell are also contemplated to be particularly useful in the methods disclosed herein. Exemplary preferred PEAMT or  $\Delta$ PEAMT polypeptides and proteins are those that comprise, consist essentially of, or consist of, at least a first sequence region that comprises an at least about 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 205, 210, 215, 220, 225, 230, 235, 240, 245, 250, 255, 260, 265, 270, 275, 280, or even 285 or more contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4 (and even up to and including the 289 amino acid full-length sequence of SEQ ID NO:4). Highly preferred PEAMT polypeptides also include those proteins and polypeptides that comprise, consist essentially of, or consist of, at least a first sequence region that comprises an at least about 290, 295, 300, 310, 320, 330, 340, 350, 360, 370, 380, 390, 400, 410, 420, 430, 440, 450, 460, 470, 480, 490, or even up to an including the 494-amino acid full-length sequence of SEQ ID NO:2. The plant-derived PEAMT and  $\Delta$ PEAMT polypeptides and proteins of the invention also exemplified by proteins such as those isolated from spinach, that comprise, consist essentially of, or consist of, at least a first sequence region that comprises the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.

## 2.2 PEAMT POLYNUCLEOTIDES AND POLYNUCLEOTIDE COMPOSITIONS

In a another embodiment, the invention concerns polynucleotides that comprise at least a first nucleic acid segment that: (a) comprises, consists essentially of, or consists of a sequence that encodes one or more PEAMT or  $\Delta$ PEAMT polypeptides as described herein, or (b) comprises, consists essentially of, or consists of, a sequence of at least about 27 or more contiguous nucleotides from SEQ ID NO:1 or SEQ ID NO:3. As such, polynucleotides that comprise at least a first nucleic acid segment that encodes one or more PEAMT or

ΔPEAMT polypeptides, and that comprises, consists essentially of, or consists of a nucleic acid sequence of from about 30 or 40 to about 10,000 nucleotides are contemplated to be particularly preferred in the methods of the present invention. Similarly, polynucleotides that comprise at least a first nucleic acid segment that encodes one or more PEAMT or ΔPEAMT 5 polypeptides and that comprises, consists essentially of, or consists of, a sequence region of from about 50 or 60 to about 8,000 or 9,000 nucleotides, or those polynucleotides that comprise at least a first nucleic acid segment that encodes one or more PEAMT or ΔPEAMT polypeptides and that comprises, consists essentially of, or consists of, a sequence region of from about 70 or 80 to about 6,000 or 7,000 nucleotides, or those polynucleotides that 10 comprise at least a first nucleic acid segment that encodes one or more PEAMT or ΔPEAMT polypeptides and that comprises, consists essentially of, or consists of, a sequence region of from about 90 or 100 to about 4,000 or 5,000 nucleotides in length, are particularly preferred in the practice of the present invention.

For polynucleotides encoding PEAMT or ΔPEAMT polypeptides, such 15 polynucleotides preferably comprise at least a first nucleic acid segment that comprises, consists essentially of, or consists of, a sequence from at least about 27 to about 2235 contiguous nucleotides from SEQ ID NO:1, or from at least about 27 to about 1120 contiguous nucleotides from SEQ ID NO:3.

Highly preferred polynucleotides are those that comprise at least a first nucleic acid 20 segment that comprises, consists essentially of, or consists of, a sequence that encodes a polypeptide that comprises an at least about 30 contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4 are contemplated to be particularly preferred in the methods of the present invention.

Naturally, all intermediate contiguous sequences are contemplated to fall within the 25 scope of the present invention. For example, polynucleotides that comprise at least about 27, at least about 28, at least about 29, at least about 30, at least about 31, at least about 32, at least about 33, at least about 34, at least about 35, at least about 36, at least about 37, at least about 38, at least about 39, at least about 40, at least about 41, at least about 42, at least about 43, at least about 44, at least about 45, at least about 46, at least about 47, at least about 48, at 30 least about 49, at least about 50, at least about 51, at least about 52, at least about 53, at least

about 54, at least about 55, at least about 56, at least about 57, at least about 58, at least about 59, at least about 60, at least about 61, at least about 62, at least about 63, at least about 64, at least about 65, at least about 66, at least about 67, at least about 68, at least about 69, at least about 70, at least about 71, at least about 72, at least about 73, at least about 74, at least about 5 75, at least about 76, at least about 77, at least about 78, at least about 79, at least about 80, at least about 85, at least about 90, at least about 95, or at least about 100 or more contiguous nucleotides from SEQ ID NO:1 or SEQ ID NO:3 are contemplated to be particularly preferred in the methods of the present invention, and are contemplated to be particularly preferred polynucleotide compositions.

10 Likewise, PEAMT- and  $\Delta$ PEAMT-encoding polynucleotides that comprise at least about 110, at least about 120, at least about 130, at least about 140, at least about 150, at least about 160, at least about 170, at least about 180, at least about 190, or at least about 200 or more contiguous nucleotides from SEQ ID NO:1 or SEQ ID NO:3 are contemplated to be particularly preferred polynucleotide compositions. PEAMT-specific polynucleotides that 15 comprise at least about 250, at least about 300, at least about 350, at least about 400, at least about 450, at least about 500, at least about 550, at least about 600, or more contiguous nucleotides from SEQ ID NO:1 or SEQ ID NO:3 are contemplated to be particularly preferred polynucleotide compositions.

When it is desirable to employ PEAMT- or  $\Delta$ PEAMT-encoding polynucleotides that 20 are significantly more homologous to the polynucleotide sequences disclosed herein, polynucleotide compositions may be selected that encode PEAMT- or  $\Delta$ PEAMT-derived peptides that comprise at least about 650, at least about 700, at least about 750, at least about 800, at least about 850, at least about 900, at least about 950, at least about 1000, at least about 1050, at least about 1100 or more contiguous nucleotides from SEQ ID NO:1 or SEQ 25 ID NO:3, and even those up to and including the full-length DNA sequence disclosed in SEQ ID NO:3 are contemplated to be particularly preferred polynucleotide compositions. Likewise, those sequences that comprise at least about 1200, about 1300, about 1400, about 1500, about 1600, about 1700, about 1800, about 1900, about 2000, about 2100, about 2200, and even those up to and including the full-length DNA sequence of SEQ ID NO:1 are also 30 contemplated to be particularly preferred polynucleotide compositions.

Likewise, the PEAMT and  $\Delta$ PEAMT polynucleotide compositions of the present invention also encompass at least a first nucleic acid segment that comprises, consists essentially of, or consists of a sequence that encodes a polypeptide having PEAMT or  $\Delta$ PEAMT activity, and particularly those that comprise a nucleic acid sequence that encodes 5 an at least about 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 or more contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4.

The invention further provides isolated nucleic acid segments that: (a) encodes a polypeptide having  $\Delta$ PEAMT activity and that comprises an at least 27 contiguous amino acid sequence from SEQ ID NO:4; (b) encodes a polypeptide having  $\Delta$ PEAMT activity and at 10 least about 75% sequence identity with the amino acid sequence of SEQ ID NO:4; (c) comprises an at least 26 contiguous nucleotide sequence from SEQ ID NO:3; or (d) hybridizes to the sequence of SEQ ID NO:3, or that hybridizes to the complement thereof, under stringent hybridization conditions.

Exemplary PEAMT and  $\Delta$ PEAMT nucleic acid segments include those that comprise, 15 consist essentially of, or consist of, at least a first sequence region that encodes a polypeptide having an at least 28 or 29 contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4, although longer contiguous sequences such as at least 30, at least 31, at least 33, at least 35, or at least 37 or more contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4 are also contemplated to be particularly preferred. In illustrative embodiments, 20 the isolated PEAMT or  $\Delta$ PEAMT nucleic acid segment comprises at least a first sequence region that encodes a polypeptide comprising the sequence of SEQ ID NO:4.

The isolated polynucleotides of the invention preferably comprise at least a first sequence region that encodes at least a first peptide or polypeptide that has at least about 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79% or 80% or greater sequence identity with 25 the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4. More preferably, the polynucleotides of the invention preferably comprise at least a first sequence region that encodes at least a first peptide or polypeptide that has at least about 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, or 90% or greater sequence identity with the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4. More preferably still, the polynucleotides of the invention 30 preferably comprise at least a first sequence region that encodes at least a first peptide or

polypeptide that has at least about 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% or greater sequence identity with the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.

Preferred polynucleotides of the present invention typically will comprise at least a first sequence region that comprises an at least 31 contiguous nucleotide sequence from between about position 254 to about position 1735 of SEQ ID NO:1, or an at least 31 contiguous nucleotide sequence from SEQ ID NO:3, although those that comprise at least a first sequence region that comprises longer contiguous nucleotide sequences from between about position 254 to about position 1735 of SEQ ID NO:1, or longer contiguous nucleotide sequences from SEQ ID NO:1, such as those comprising at least about 32, about 33, about 34, about 35, about 36, about 37, about 38, about 39, about 40, about 41, about 42, about 43, about 44, about 45, about 46, about 47, about 48, about 49, or about 50, or more contiguous nucleotide sequence from between about position 254 to about position 1735 of SEQ ID NO:1, or from SEQ ID NO:3, are also highly preferred. Likewise, more preferred polynucleotides are those that comprise at least a first sequence region that comprises an at least about 51 contiguous nucleotide sequence from between about position 254 to about position 1735 of SEQ ID NO:1, or from SEQ ID NO:3, although longer contiguous nucleotide sequences from between about position 254 to about position 1735 of SEQ ID NO:1, or from SEQ ID NO:, such as those comprising at least about 52, about 53, about 54, about 55, about 56, about 57, about 58, about 59, about 60, about 61, about 62, about 63, about 64, about 65, about 66, about 67, about 68, about 69, or about 70, or more contiguous nucleotide sequence between about position 254 to about position 1735 of SEQ ID NO:1, or from SEQ ID NO:3, are even more highly preferred in particular embodiments of the invention.

In fact, the PEAMT-and ΔPEAMT-encoding polynucleotides of the present invention typically will comprise at least a first sequence region that comprises, consists essentially of, or 25 consists of, an at least about 71 contiguous nucleotide sequence from between about position 254 to about position 1735 of SEQ ID NO:1, or from SEQ ID NO:3, or may comprise, consist essentially of, or consist of, at least about 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, or even about 100 or more contiguous nucleotides from between about position 254 to about position 1735 of SEQ ID NO:1, or from 30 SEQ ID NO:3, and in fact, may comprise, consist essentially of, or consist of, at least about

105, 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, or 200 or more contiguous nucleotides between about position 254 to about position 1735 of SEQ ID NO:1, or from SEQ ID NO:3, and in fact, may comprise, consist essentially of, or consist of, at least about 205, 210, 215, 220, 225, 230, 235, 240, 245, 250, 255, 260, 265, 270, 5 275, 280, 285, 290, 295, or 300, 325, 350, 375, 400, 425, 450, 475, or 500 or more contiguous nucleotides from between about position 254 to about position 1735 of SEQ ID NO:1, or from SEQ ID NO:3. Indeed in many embodiments of the invention, the PEAMT-and  $\Delta$ PEAMT-encoding polynucleotides of the present invention will comprise at least a first sequence region that comprises, consists essentially of, or consists of, an at least about 500, 525, 550, 575, 600, 10 625, 650, 675, 700, 725, 750, 775, 800, 825, 850, 875, 900, 925, 950, 975 or 1000 or more contiguous nucleotides from between about position 254 to about position 1735 of SEQ ID NO:1, or from SEQ ID NO:3, and even up to an including all, or substantially all of the entire nucleotide sequence disclosed in SEQ ID NO:1 or SEQ ID NO:3.

Preferred polynucleotide compositions of the present invention typically will comprise, 15 consist essentially of or, will consist of a sequence region that hybridizes to the sequence of from position 254 to 1735 of SEQ ID NO:1 or to the sequence of SEQ ID NO:3, under stringent hybridization conditions. Such stringent hybridizations are well known to those of skill in the art, as are the methods for obtaining and identifying polynucleotides that hybridize to a selected target sequence. For example, as described hereinbelow, stringent hybridization 20 conditions comprising a salt concentration of from about 0.02 M to about 0.15 M, and a temperature of from about 50°C to about 70°C are particularly preferred.

A further embodiment of the present invention concerns an isolated polynucleotide that comprises: (a) a sequence region that comprises, consists essentially of, or consists of, at least 26 contiguous nucleotides that have the same sequence as, or are complementary to, at least 26 25 contiguous nucleotides of SEQ ID NO:1 or SEQ ID NO:3; or (b) a sequence region of from 26 to about 10000 nucleotides in length that hybridizes to the nucleic acid segment of SEQ ID NO:1 or SEQ ID NO:3; or the complement thereof, under hybridization conditions comprising a salt concentration of from about 0.02 M to about 0.15 M, and a temperature of from about 50°C to about 70°C. Such polynucleotides may range in size from on the order of about 100 to 30 about 11,000 nucleotides in length, with intermediate ranges such as from about 1000 to

about 9,000 nucleotides in length, or from about 2000 to about 7,000 nucleotides in length, or from about 4000 to about 6000 nucleotides in length being particular preferred.

Preferred polynucleotide compositions will typically comprise an RNA, a PNA, or a DNA segment, as described hereinbelow. Such compositions may be comprised within a recombinant vector such as a plasmid, cosmid, phage, phagemid, baculovirus, bacterial artificial chromosome, or yeast artificial chromosome vector. Likewise, the disclosed polynucleotides may be comprised within a recombinant virus or virion. It may be operably linked to a promoter, and particularly to a heterologous promoter such as a plant-expressible constitutive, inducible, or tissue-specific promoter. Exemplary plant-expressible promoters include those listed herein in Table 1, Table 2, and Table 3, and include such well known promoters as corn sucrose synthetase 1, corn alcohol dehydrogenase 1, corn light harvesting complex, corn heat shock protein, pea small subunit RuBP carboxylase, Ti plasmid mannopine synthase, Ti plasmid nopaline synthase, petunia chalcone isomerase, bean glycine rich protein 1, Potato patatin, lectin, CaMV 35S, ALS, ubiquitin, globulin 1, cruciferin, napin,  $\beta$ -conglycinin, phaseolin,  $\gamma$  zein, or the S-E9 small subunit RuBP carboxylase promoter.

Such polynucleotides may be used, for example, in preparing a recombinant vector, a transgenic plant, or a recombinant polypeptide composition. Such polynucleotide compositions may also be used as a probe for screening a plant nucleic acid library to identify a gene encoding a polypeptide having PEAMT or  $\Delta$ PEAMT activity. Alternatively, their sequence information may be used in the preparation of a target sequence probe to employ a computer-based algorithm to search a computerized database of sequences such as genomic, or expressed sequence tags, cDNAs, and the like to identify a gene encoding a polypeptide having PEAMT or  $\Delta$ PEAMT activity.

The recombinant vectors of the present invention may also be used in producing a transformed plant cell or plant tissue, a pluripotent plant cell, or a transgenic plant that expresses a polypeptide having PEAMT or  $\Delta$ PEAMT activity.

### 2.3 PEAMT VECTORS, AND HOST CELL

In a related embodiment, the invention provides a host cell comprising at least a first recombinant vector that comprises at least a first heterologous expression unit that comprises at least a first heterologous promoter operably linked to at least a first sequence region that 5 encodes a PEAMT or  $\Delta$ PEAMT polypeptide. Such a host cell may be a bacterial cell such as an *Escherichia*, *Salmonella* or *Agrobacterium* cell, or alternatively, may be an eukaryotic cell, such as a plant cell. Alternatively, the polynucleotide may be comprised within a virus, virion, or viral vector. Exemplary recombinant vectors described in Example 1 have been designated pREP3-PEAMT and pREP3- $\Delta$ PEAMT, which encode PEAMT and  $\Delta$ PEAMT polypeptides, 10 respectively.

### 2.4 METHODS FOR DETECTING PEAMT COMPOSITIONS

The invention also provides hereinbelow methods for detecting a PEAMT- or a  $\Delta$ PEAMT-encoding polynucleotide in a sample. Such a method typically involves the steps of: 15 (a) contacting a population of polynucleotides suspected of encoding a PEAMT or a  $\Delta$ PEAMT polypeptide with at least a first PEAMT/ $\Delta$ PEAMT polynucleotide, under conditions effective to allow hybridization of substantially complementary nucleic acids; and (b) detecting the hybridized complementary nucleic acids so formed. Such detection may readily be accomplished, for example, by labeling a selected PEAMT-specific nucleic acid segment with a 20 detectable label, and employing such a labeled segment as a probe in an assay such as a hybridization or Southern analysis.

A method is also provided for detecting a PEAMT or a  $\Delta$ PEAMT polypeptide in a biological sample. This method typically involves contacting a biological sample suspected of containing a PEAMT or a  $\Delta$ PEAMT polypeptide with a PEAMT- or  $\Delta$ PEAMT-specific 25 antibody or antigen binding fragment thereof, under conditions effective to allow the formation of immune complexes, and detecting the immune complexes so formed. Such detecting may be accomplished by labeling the primary antibody or antigen-binding fragment, or by utilizing a labeled secondary antibody that is immunospecific for the primary antibody, or the primary antigen binding fragment.

The invention also provides a method of identifying a nucleic acid segment that encodes a plant PEAMT or  $\Delta$ PEAMT polypeptide by using an *in vivo* complementation assay. The method generally involves expressing a plurality of plant polynucleotides suspected of encoding a plant PEAMT or  $\Delta$ PEAMT polypeptide in a population of *Schizosaccharomyces pombe* cho2<sup>-</sup> cells, and selecting at least a first cell from the population that is capable of growth on a minimal medium comprising ethanolamine under conditions effective for the growth of the cell, where growth of a cell is indicative of the cell's containing a PEAMT- or  $\Delta$ PEAMT- encoding polynucleotide. Such complementation methods are described in detail hereinbelow in Example 1.

10

## 2.5 PEAMT DETECTION KITS

The invention also provides nucleic acid detection kits that typically comprise in suitable container means, at least a first isolated PEAMT or a  $\Delta$ PEAMT-specific nucleic acid segment, and instructions for using the PEAMT- or  $\Delta$ PEAMT-specific nucleic acid segment to detect other PEAMT/ $\Delta$ PEAMT sequences or to use as probes or primers for related and DNA sequencing methodologies and the like. Such kits may also optionally comprise a detectable label, and/or one or more related or unrelated nucleic acid segments useful as positive or negative controls, respectively. Such kits may also comprise one or more restriction endonucleases.

20 Alternatively, the invention provides antibody or antigen binding fragment detection kits that typically comprise in suitable container means, at least a first isolated PEAMT or a  $\Delta$ PEAMT-specific peptide or polypeptide, and instructions for using the PEAMT- or  $\Delta$ PEAMT-specific peptide or polypeptide in one or more immunological assays to specifically detect the presence of an antibody or antigen binding fragment in a biological sample that is 25 immunospecific for the PEAMT/ $\Delta$ PEAMT peptide or protein. Such immunoassay kits may also optionally comprise a detectable label, and/or one or more related or unrelated polypeptides useful as positive or negative controls.

Likewise, the invention provides polypeptide detection kits that typically comprise in suitable container means, at least a first isolated PEAMT or a  $\Delta$ PEAMT-specific antibody or 30 antigen binding fragment, and instructions for using the PEAMT- or  $\Delta$ PEAMT-specific

antibody or antigen binding fragment as a probe in one or more immunological assays to specifically detect the presence of a PEAMT or  $\Delta$ PEAMT peptide or protein in a biological sample. Such immunoassay kits may also optionally comprise a detectable label, and/or one or more related or unrelated antibodies useful as positive or negative controls.

5

## 2.6 PEAMT TRANSGENIC PLANTS AND PRODUCTION METHODS THEREFOR

The invention further provides a transgenic plant that comprises: (a) a heterologous nucleic acid segment that comprises a PEAMT or a  $\Delta$ PEAMT polynucleotide; (b) a transformed host cell that expresses a PEAMT or a  $\Delta$ PEAMT polypeptide; (c) a recombinant 10 virus that expresses a PEAMT or a  $\Delta$ PEAMT polypeptide; or (d) a recombinant vector that encodes a PEAMT or a  $\Delta$ PEAMT polypeptide.

The transgenic plant preferably has stably incorporated into its genome a heterologous nucleic acid segment that comprises a PEAMT or a  $\Delta$ PEAMT polynucleotide, wherein the polynucleotide is operably linked to a promoter that expresses the polynucleotide in the cells 15 and tissues of the transgenic plant. Such transgenic plants are preferably monocotyledonous or dicotyledonous plants, such as grains, trees, legumes, fibers, vegetables, fruits, berries, nuts, citrus, grasses, cacti, succulents, flowers, or other ornamental plants.

Exemplary plants include, but are not limited to, corn, rice, millet, tobacco, alfalfa, soybean, bean, sorghum, pea, *Brassica*, safflower, potato, coconut, palm, pumpkin, squash, 20 poppy, sesame, peanut, cocoa, coffee, tomato, flax, sugar beets, canola, sunflower, cotton, kapok, wheat, oat, barley, walnut, pecan, almond, and rye.

The invention further discloses and claims progeny of any generation of such transgenic plant, as well as the seed of any generation of such transgenic plants, and seed of any 25 generation, offspring, or subsequent progeny of such transgenic plants. Particularly encompassed by the invention are seeds, nuts, legumes, and the like, that have an increased level of lipid, relative to untransformed plants of the same species that do not contain one or more exogenously provided PEAMT- or  $\Delta$ PEAMT-encoding transgenes. Such seeds are particularly preferred for animal foodstuffs, as well as those having increased protein and nutrition content suitable for human consumption.

30

## 2.7 METHODS FOR ALTERING PEAMT EXPRESSION IN TRANSGENIC PLANTS

The present invention also provides a method for modulating the amount or activity of a PEAMT or a  $\Delta$ PEAMT polypeptide in a plant, plant tissue, plant seed, or plant cell. This method typically involves expressing in such a plant, plant tissue, plant seed, or plant cell a 5 biologically effective amount of a PEAMT- or a  $\Delta$ PEAMT-specific polynucleotide. The term “biologically effective amount” will be understood by the skilled artisan to mean an amount of the polynucleotide composition that is effective to produce the desired phenotypic trait in the resulting transformed plant, plant tissue, plant seed, or plant cell, *e.g.*, an altered or increased 10 level or amount of PEAMT/ $\Delta$ PEAMT polypeptide or PEAMT/ $\Delta$ PEAMT activity in the plant, plant tissue, plant seed, or plant cell when compared to a similar untransformed or “wild-type” plant cell.

A method of altering, modulating, increasing, or decreasing the level of a lipid or phospholipid compound (such as those selected from the group consisting of phosphatidylcholine, phosphocholine, choline, choline-*O*-sulfate and glycine betaine) in a 15 plant, plant tissue, plant seed, or plant cell is also provided by the invention. This method generally comprises, expressing in a suitable plant cell a biologically effective amount of a PEAMT or a  $\Delta$ PEAMT polynucleotide. Such an effective amount will be recognized by the skilled artisan as an amount necessary to alter, modulate, decrease, increase, or improve the level or extent of one or more of the choline metabolites in the plant cells and plant tissues. 20 This method is particularly useful for increasing the level of or altering the lipid content of the cells, fruits, seeds, and tissues of the transformed plant, and for modulating the tolerance of the plant to osmotic or cryogenic stress.

This method provides means for modulating the biosynthesis of phosphatidylcholine, phosphocholine, choline, or metabolites thereof, such as, but not limited to, glycine betaine 25 or choline-*O*-sulfate in a plant, increasing the lipid level or altering the lipid compositions of plant cells, or improving the overall nutritional value of the plant for consumption by animals and/or humans.

Using the PEAMT compositions of the invention, it is possible to regulate the level of one or more cellular lipids in a transformed host cell expressing these compositions. This 30 method provides means for modulating the biosynthesis of the osmoprotecting and

5 cryoprotecting compounds of a cell such as glycine betaine and choline-*O*-sulfate. Such methods may further involve transforming the cell with an additional nucleic acid construct that encodes an enzyme involved in glycine betaine or choline-*O*-sulfate biosynthesis. Such enzymes may include for example, choline monooxygenase, choline oxidase, or choline dehydrogenase. The method may further comprise altering the level of the PEAMT precursors, phosphoethanolamine and/or ethanolamine to provide greater intracellular pools of the substrate for the PEAMT enzyme. The transformation of selected plants with one or more genetic constructs encoding choline monooxygenase, choline oxidase, or choline dehydrogenase may be required when the plant lacks such enzymes, as is the case with plants  
10 such as potatoes and rice. Also, it may be desirable to augment the level of one or more of these glycine betaine biosynthetic enzymes in plants that natively express one or more of such enzymes. This may be the case when transforming crops such as barley, corn, and sugar beets.

15 An exemplary method may further comprise the steps of growing the transformed plant under conditions effective for obtaining seeds from the plant, and collecting the seeds so produced by the transformed plant. This method is particularly desirable for the recovery of high-nutritional value seeds, grains, nuts, flours, and the like. The method is particularly desirable for the recovery of seeds having altered lipid content, or an altered distribution of the lipids native to the seed of the particular species from which the seed was obtained, when  
20 compared to an unmodified seed of the same species. The method may also further comprise the step of (e) transforming the plant cell with one or more additional polynucleotides that encode one or more phosphatidylcholine biosynthesis enzymes, such as one or more enzymes in the pathway for phosphocholine synthesis, to further increase in the cells and tissues of the plant the level of phosphatidylcholine, or to otherwise alter the lipid content of the cells.  
25 Increasing the amount of phosphatidylcholine or altering the lipid content of plant seed, may be achieved by further growing such a transgenic plant, under conditions effective to produce seed and then obtaining the seed produced from the plant.

## 2.8 COMPOSITIONS FOR GENE DETECTION, AMPLIFICATION, AND SEQUENCING

In related embodiments, the invention provides methods and compositions for detecting homologous PEAMT/ΔPEAMT-encoding polynucleotides and homologous PEAMT/ΔPEAMT polypeptides.

For detection and sequencing of polynucleotides, it is often desirable to isolate smaller polynucleotides for use as hybridization probes, synthesis or sequencing primers, and the like as described in detail herein. In such embodiments, shorter polynucleotide sequences are particularly desirable, including those that comprise a sequence of at least about 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, or 40 or more contiguous nucleotides from one or more of the DNA sequences disclosed in SEQ ID NO:1 and SEQ ID NO:3. These sequences find particular utility as probes for screening clone banks, colony blots, or as computer homology search strings for identifying homologous polynucleotide sequences *via* computer-based algorithm homology searches, or as primers in isolation and amplification methodologies such as, for example, PCR™, RT-PCR™, cloning studies, and such like. This is particularly important when it is desirable to screen a clone bank of sequences, such as for example, genomic or cDNA libraries or arrays, expressed sequence tags (ESTs) or genomic or chromosomal libraries.

For example, polynucleotides that comprise at least a first nucleic acid segment that comprises, consists essentially of, or consists of, at least about 27, at least about 28, at least about 29, at least about 30, at least about 31, at least about 32, at least about 33, at least about 34, at least about 35, at least about 36, at least about 37, at least about 38, at least about 39, at least about 40, at least about 41, at least about 42, at least about 43, at least about 44, at least about 45, at least about 46, at least about 47, at least about 48, at least about 49, at least about 50, at least about 51, at least about 52, at least about 53, at least about 54, at least about 55, at least about 56, at least about 57, at least about 58, at least about 59, or even at least about 60 or so nucleotides from one of the sequences disclosed in SEQ ID NO:1 or SEQ ID NO:3 are particularly suited for these embodiments.

## 2.9 IDENTIFICATION OF HOMOLOGOUS PEAMT POLYPEPTIDES AND POLYNUCLEOTIDES

In addition to the particular illustrative polypeptide and polynucleotide sequences disclosed herein, those having benefit of the present teaching are now also able to identify and characterize a wide variety of PEAMT and  $\Delta$ PEAMT homologs and/or isozymes, as well as to

5 identify, characterize, and sequence a variety of PEAMT/ $\Delta$ PEAMT-encoding polynucleotides from a variety of plant species. In fact, the inventors contemplate that any plant-derived PEAMT/ $\Delta$ PEAMT protein or peptide can be identified using the methods disclosed herein and may be obtained by using the immunological methods disclosed herein to obtain PEAMT/ $\Delta$ PEAMT proteins and peptides from a variety of disparate species. Alternatively, the  
10 inventors contemplate that those of skill in the art having the benefit of the teachings disclosed herein will be able to identify PEAMT/ $\Delta$ PEAMT-encoding polynucleotides either by comparison of one or more of the disclosed sequences to computer databases of plant EST sequences, and identification of highly homologous sequences, or alternatively, by traditional hybridization screening methods employing one or more labeled PEAMT- or  $\Delta$ PEAMT-  
15 specific polynucleotide sequences to screen a population of target nucleic acids, such as *e.g.*, a cDNA or other such genetic library, a colony or clone bank, or by screening individual isolates from particular plant species.

Because the inventors have successfully demonstrated the presence of PEAMT-specific polynucleotides and polypeptides in at least two plant genera (*i.e.* *Spinacia oleracea* and  
20 *Arabidopsis*), they contemplate the additional identification of related PEAMT and  $\Delta$ PEAMT polypeptides and the gene sequences that encode them. In particular, the inventors contemplate the identification of PEAMT variants, homologs, and related sequences using one or more of the methods disclosed herein to identify a family of PEAMT sequences. Likewise, one of skill in the art will even be able to utilize the teachings of the present disclosure to identify other  
25 PEAMT-like polypeptides and polynucleotides, including those from related and from distantly-related plant species and to use these sequences in the preparation of transgenic plants having modified lipid content or altered PEAMT-dependent biosyntheses. By hybridization, immunological, and computer-based homology algorithms, the inventors further contemplate the identification and characterization of PEAMT-specific compositions from species that are  
30 not yet even described or characterized as possessing PEAMT activity.

In addition to the particular full-length PEAMT polypeptides disclosed in SEQ ID NO:2 and the ΔPEAMT sequence shown in SEQ ID NO:4, the inventors also contemplate the preparation and use of substantially full-length sequences in certain embodiments. As such, polypeptides may be obtained that comprise from at least about 80% or so, and up to and 5 including those having at least about 99% of the full-length PEAMT primary amino acid sequence as disclosed herein, and yet still possess significant PEAMT enzymatic activity *in vitro* and *in vivo*. In fact, “truncated” polypeptides or “near-full-length” or “substantially full-length” polypeptides are well known in the plant molecular biological arts to often possess all, or almost all of the enzymatic activity that the full-length polypeptide possesses. In many 10 embodiments, these slightly shorter polypeptide sequences may be desirable for use in many of the disclosed methods. This is particularly true, when the creation of “chimeric” polypeptides is desired, as well as in the creation of hybrid polypeptides that have, for example, the addition of a particular amino acid sequence to “target” the localization of the polypeptide to a particular cellular location, or to a particular region of the plant in which the polypeptide is expressed. 15 For example, the preparation of a fusion protein that possesses both PEAMT activity, yet further comprises a sequence region that targets the peptide to a particular cellular region, such as the membrane, or to a particular organelle, *etc.* is often desirable. As such, truncated or fusion proteins that comprise only about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, 20 about 94%, about 95%, about 96%, about 97%, about 98%, or about 99% of the primary amino acid sequence as disclosed in SEQ ID NO:2 or SEQ ID NO:4 are particularly embodied by the present invention.

In the same that a PEAMT polypeptide need not include the entire sequences as disclosed in SEQ ID NO:2 and SEQ ID NO:4 to be useful in the practice of the present 25 methods, the primary amino acid sequence of a particular PEAMT polypeptide need not have complete sequence identity to either of the polypeptides disclosed in SEQ ID NO:2 and SEQ ID NO:4. In fact, the primary amino acid sequence of a particular PEAMT polypeptide need only comprise enough of the primary PEAMT sequence to substantially perform its enzymatic function *in vivo* or *in situ*. Owing to the nature of conservative amino acid replacement, indeed 30 several amino acids may be changed, altered, mutagenized, or even deleted in the primary

amino acid sequence of a particular PEAMT protein and yet still give rise to a functional PEAMT polypeptide which still possesses an enzymatic activity similar or identical to that of the native full-length PEAMTs disclosed in the following examples.

In fact, it is well known in the plant molecular biological arts that two polypeptides from different species may differ slightly, or even sometimes, substantially in their primary amino acid sequence, and yet, still possess the same biological activity. As such, homologous or “cognate” PEAMT polypeptides may be designed synthetically, site-specifically modified, or isolated from different biological sources, that possess similar PEAMT enzymatic activity, but yet share less than 100% identity at the primary amino acid level with one of the PEAMT sequences disclosed herein. In fact, such PEAMT homologous polypeptides may share approximately 60% or 65% sequence identity with one or more of the disclosed sequences herein. More homologous PEAMT sequences will include those polypeptides that are from about 70% to about 80% identical to either of the polypeptides of SEQ ID NO:2 or SEQ ID NO:4. Still more homologous PEAMT sequences will include those polypeptides that share from about 85% to about 95% sequence identity with one or more of the polypeptides disclosed in SEQ ID NO:2 or SEQ ID NO:4.

When highly homologous polypeptide are identified that possess PEAMT enzymatic activity, such as is often the case when polypeptides are obtained from closely-related species, cultivars, or hybrids, the PEAMT polypeptides identified may share about 96%, about 97%, about 98%, or even about 99% or more sequence identity with one or more of the sequences disclosed in SEQ ID NO:2 or SEQ ID NO:4. Naturally, all intermediate % identity values are contemplated to fall within the scope of the present disclosure. As such, polypeptides having about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, and those having about 98% primary amino acid sequence identity to the sequences disclosed in SEQ ID NO:2 or SEQ ID NO:4 are contemplated to be useful in the formulation of the methods and compositions of the present invention.

In preferred embodiments, the PEAMT polypeptides of the present invention comprise an at least about 7 or 8 contiguous amino acid sequence from SEQ ID NO:2 or SEQ

5 ID NO:4. Likewise, the PEAMT compositions of the present invention also encompass those polypeptides that have PEAMT activity, and that comprise an amino acid sequence of at least about 9 or 10 contiguous amino acids from SEQ ID NO:2 or SEQ ID NO:4. In more preferred embodiments, the PEAMT polypeptides of the present invention comprise an at least about 11 or 12 contiguous amino acid sequence from one of these full-length sequences. When it is desirable to identify PEAMT polypeptides that are still more homologous to the disclosed sequences, one may wish to utilize PEAMT polypeptides that comprise an at least about 13 or 14 or 15 or 16 contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4.

10 15 Highly preferred PEAMT polypeptides are those that comprise an at least about 17 or 18 or 19 or 20 contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4, and that have PEAMT enzymatic activity when expressed in a suitable plant host cell cultured under the appropriate conditions for PEAMT expression and enzymatic activity. Likewise, PEAMT polypeptides that comprise an at least about 21 or 22 or 23 or 24 contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4, and that have PEAMT enzymatic activity when expressed in a suitable plant host cell are also contemplated to be particularly useful in the methods disclosed herein.

20 25 In certain circumstances, it may be desirable to employ PEAMT polypeptides that are even more homologous to the sequences disclosed in SEQ ID NO:2 or SEQ ID NO:4. In those embodiments, the PEAMT polypeptides of the invention will preferably comprise a primary amino acid sequence that comprises an at least about 25, and at least about 30, an at least about 35, an at least about 40, an at least about 45, or an at least about 50 or so contiguous amino acid sequence selected from SEQ ID NO:2 or SEQ ID NO:4. Likewise, in other embodiments, it may be desirable to employ PEAMT polypeptides that are even more homologous to the sequences disclosed in SEQ ID NO:2 and SEQ ID NO:4. In those 30 embodiments, the PEAMT polypeptides of the invention will preferably comprise a primary amino acid sequence that comprises an at least about 55, and at least about 60, an at least about 65, an at least about 70, an at least about 75, or an at least about 80 or so contiguous amino acid sequence selected from SEQ ID NO:2 or SEQ ID NO:4. In fact, when more highly homologous PEAMT polypeptides are contemplated, those having an at least about

85, and at least about 90, an at least about 95, an at least about 100, an at least about 110, an at least about 120, an at least about 125, an at least about 130, an at least about 135, an at least about 140, an at least about 145, or an at least about 150, or so contiguous amino acid sequence selected from SEQ ID NO:2 or SEQ ID NO:4 will be particularly preferred.

5

## 2.10 RECOMBINANT VECTORS

One important embodiment of the invention is a recombinant vector that comprises a nucleic acid segment encoding one or more of the novel polypeptides disclosed herein. Such a vector may be transferred to and replicated in a prokaryotic or eukaryotic host, with 10 bacterial cells being particularly preferred as prokaryotic hosts, and plant cells being particularly preferred as eukaryotic hosts. In preferred embodiments, the recombinant vector comprises a nucleic acid segment encoding the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4. Highly preferred nucleic acid segments are those which comprise an at least 27 base pair contiguous sequence from SEQ ID NO:1 or SEQ ID NO:3.

15 Another important embodiment of the invention is a transformed host cell that expresses one or more of these recombinant vectors. The host cell may be either prokaryotic or eukaryotic, and particularly preferred host cells are those that express the nucleic acid segment(s) comprising the recombinant vector that encodes one or more PEAMT polypeptides. Bacterial cells are particularly preferred as prokaryotic hosts, and plant cells 20 are particularly preferred as eukaryotic hosts.

In accordance with the present invention, nucleic acid sequences include and are not limited to DNA, including and not limited to cDNA and genomic DNA, genes; RNA, including and not limited to mRNA and tRNA; PNAs (peptide nucleic acids), antisense sequences, nucleosides, and suitable nucleic acid sequences such as those set forth herein, as 25 well as variants in the nucleic acid sequences such as alterations, deletions, mutations, and homologs capable of expressing the PEAMT polypeptides of the present invention.

As such the present invention also concerns DNA segments, that are free from total genomic DNA and that encode the novel PEAMT proteins disclosed herein. DNA segments encoding these peptide species may prove to encode proteins, polypeptides, subunits, 30 functional domains, and the like of PEAMT-related or other non-related gene products. In

addition these DNA segments may be synthesized entirely *in vitro* using methods that are well known to those of skill in the art.

As used herein, the term "DNA segment" refers to a DNA molecule that has been isolated free of total genomic DNA of a particular species. Therefore, a DNA segment 5 encoding a PEAMT polypeptide or peptide refers to a DNA segment that contains PEAMT polypeptide coding sequences yet is isolated away from, or purified free from, total genomic DNA of the species from which the DNA segment is obtained. Included within the term "DNA segment", are DNA segments and smaller fragments of such segments, and also recombinant vectors, including, for example, plasmids, cosmids, phagemids, phage, viruses, 10 and the like.

Similarly, a DNA segment comprising an isolated or purified nucleic acid or gene sequence that encodes a PEAMT polypeptide refers to a DNA segment which may include in addition to peptide encoding sequences, certain other elements such as, regulatory sequences, isolated substantially away from other naturally occurring genes or protein-encoding 15 sequences. In this respect, the term "gene" is used for simplicity to refer to a functional protein-, polypeptide- or peptide-encoding unit. As will be understood by those in the art, this functional term includes both genomic sequences, operon sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides or peptides.

20 "Isolated substantially away from other coding sequences" means that the gene of interest, in this case, a gene encoding a PEAMT polypeptide, forms the significant part of the coding region of the DNA segment, and that the DNA segment does not contain large portions of naturally-occurring coding DNA, such as large chromosomal fragments or other functional genes or operon coding regions. Of course, this refers to the DNA segment as 25 originally isolated, and does not exclude genes, recombinant genes, synthetic linkers, or coding regions later added to the segment by the hand of man.

In particular embodiments, the invention concerns isolated DNA segments and recombinant vectors incorporating DNA sequences that encode a PEAMT peptide or polypeptide species that includes within its amino acid sequence an amino acid sequence 30 essentially as set forth in SEQ ID NO:2 or SEQ ID NO:4.

The term "a sequence essentially as set forth in SEQ ID NO:2 or SEQ ID NO:4" means that the sequence substantially corresponds to a portion of the sequence of SEQ ID NO:2 or SEQ ID NO:4, and has relatively few amino acids that are not identical to, or a biologically functional equivalent of, the amino acids of any of these sequences. The term 5 "biologically functional equivalent" is well understood in the art and is further defined in detail herein (e.g., see Illustrative Embodiments).

Accordingly, sequences that have between about 65% and about 75% or between about 75% and about 85%, or more preferably between about 86% and about 90%, or even more preferably between about 91% or 92% or 93% and about 97% or 98% or 99% amino 10 acid sequence identity or functional equivalence to the amino acids of SEQ ID NO:2 or SEQ ID NO:4, will be sequences that are "essentially as set forth in SEQ ID NO:2 or SEQ ID NO:4."

It will also be understood that amino acid and nucleic acid sequences may include additional residues, such as additional N- or C-terminal amino acids or 5' or 3' sequences, 15 and yet still be essentially as set forth in one of the sequences disclosed herein, so long as the sequence meets the criteria set forth above, including the maintenance of biological protein activity where protein expression is concerned. The addition of terminal sequences particularly applies to nucleic acid sequences that may, for example, include various non-coding sequences flanking either of the 5' or 3' portions of the coding region or may include 20 various internal sequences, *i.e.* introns, which are known to occur within genes.

The nucleic acid segments of the present invention, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is 25 therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol.

For example, nucleic acid fragments may be prepared that include a short contiguous stretch encoding the peptide sequence disclosed in SEQ ID NO:2 or SEQ ID NO:4, or that 30 are identical to or complementary to DNA sequences which encode the peptide disclosed in

SEQ ID NO:2 or SEQ ID NO:4, and particularly the DNA segments disclosed in SEQ ID NO:1 or SEQ ID NO:3.

Highly preferred nucleic acid segments of the present invention comprise one or more PEAMT-encoding genes of the invention, or a portion of one or more PEAMT-encoding genes of the invention. For certain application, relatively small contiguous nucleic acid sequences are preferable, such as those which are about 14 or 15 or 16 or 17 or 18 or 19, or 5 20, or 21, or 22, or 23, or 24, or 25, or 26, or 27, or 28, or 29, or 30-50, 51-80, 81-100 or so nucleotides in length. Alternatively, in some embodiments, and particularly those involving preparation of recombinant vectors, transformation of suitable host cells, and preparation of 10 transgenic plant cell, longer nucleic acid segments are preferred, particularly those that include the entire coding region of one or more PEAMT-encoding genes. As such, the preferred segments may include those that are up to about 20,000 or so nucleotides in length, or alternatively, shorter sequences such as those about 19,000, about 18,000, about 17,000, about 16,000, about 15,000, about 14,000, about 13,000, about 12,000, 11,000, about 10,000, 15 about 9,000, about 8,000, about 7,000, about 6,000, about 5,000, about 4,500, about 4,000, about 3,500, about 3,000, about 2,500, about 2,000, about 1,500, about 1,000, about 500, or about 200 or so base pairs in length. Of course, these numbers are not intended to be exclusionary of all possible intermediate lengths in the range of from about 20,000 to about 15 nucleotides, as all of these intermediate lengths are also contemplated to be useful, and 20 fall within the scope of the present invention.

It will be readily understood that "intermediate lengths", in these contexts, means any length between the quoted ranges, such as 14, 15, 16, 17, 18, 19, 20, *etc.*; 21, 22, 23, 24, 25, 26, 27, 28, 29, *etc.*; 30, 31, 32, 33, 34, 35, 36..... *etc.*; 40, 41, 42, 43, 44..... *etc.*, 50, 51, 52, 53..... *etc.*; 60, 61, 62, 63.... *etc.*, 70, 80, 90, 100, 110, 120, 130..... *etc.*; 200, 210, 220, 230, 25 240, 250..... *etc.*; including all integers in the entire range from about 14 to about 10,000, including those integers in the ranges 200-500; 500-1,000; 1,000-2,000; 2,000-3,000; 3,000-5,000 and the like.

In a preferred embodiment, the nucleic acid segments comprise a sequence of from about 1800 to about 18,000 base pair in length, and comprise one or more genes that encode

a PEAMT polypeptide as disclosed in SEQ ID NO:2 or SEQ ID NO:4, and particularly those polynucleotide sequences disclosed in SEQ ID NO:1 or SEQ ID NO:3.

It will also be understood that this invention is not limited to the particular nucleic acid sequences which encode peptides of the present invention, or which encode the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4, including the DNA sequences that are particularly disclosed in SEQ ID NO:1 and SEQ ID NO:3. Recombinant vectors and isolated DNA segments may therefore variously include the peptide-coding regions themselves, coding regions bearing selected alterations or modifications in the basic coding region, or they may encode larger polypeptides that nevertheless include these peptide-coding regions or may encode biologically functional equivalent proteins or peptides that have variant amino acids sequences.

The DNA segments of the present invention encompass biologically functional, equivalent peptides. Such sequences may arise as a consequence of codon redundancy and functional equivalency that are known to occur naturally within nucleic acid sequences and the proteins thus encoded. Alternatively, functionally-equivalent proteins or peptides may be created *via* the application of recombinant DNA technology, in which changes in the protein structure may be engineered, based on considerations of the properties of the amino acids being exchanged. Changes designed by man may be introduced through the application of site-directed mutagenesis techniques, *e.g.*, to introduce improvements to the antigenicity of the protein or to test mutants in order to examine activity at the molecular level

If desired, one may also prepare fusion proteins and peptides, *e.g.*, where the peptide-coding regions are aligned within the same expression unit with other proteins or peptides having desired functions, such as for purification or immunodetection purposes (*e.g.*, proteins that may be purified by affinity chromatography and enzyme label coding regions, respectively).

Recombinant vectors form further aspects of the present invention. Particularly useful vectors are contemplated to be those vectors in which the coding portion of the DNA segment, whether encoding a full-length protein or smaller peptide, is positioned under the control of a promoter. The promoter may be in the form of the promoter that is naturally associated with a gene encoding peptides of the present invention, as may be obtained by

isolating the 5' non-coding sequences located upstream of the coding segment or exon, for example, using recombinant cloning and/or PCR™ technology, in connection with the compositions disclosed herein.

5    **2.11 METHODS FOR PREPARING TRANSFORMED HOST CELLS AND TRANSGENIC PLANTS**

As described above, the invention encompasses a transgenic plant having incorporated into its genome a transgene that encodes a PEAMT polypeptide that comprises at least a first contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4. A further aspect of the invention is a transgenic plant having incorporated into its genome a 10 transgene, that comprises an at least 27 base pair contiguous nucleic acid sequence from SEQ ID NO:1 or SEQ ID NO:3. Also disclosed and claimed are progeny of such a transgenic plant, as well as its seed, progeny from such seeds, and seeds arising from the second and subsequent generation plants derived from such a transgenic plant.

The present invention also provides methods for producing a transgenic plant that 15 expresses a nucleic acid segment encoding the novel PEAMT polypeptides or proteins of the present invention. The process of producing transgenic plants is well known in the art. In general, the method comprises transforming a suitable host cell with one or more DNA segments that contain one or more promoters operatively linked to a coding region that encodes one or more of the disclosed PEAMT proteins. Such a coding region is generally 20 operatively linked to a transcription-terminating region, whereby the promoter is capable of driving the transcription of the coding region in the cell, and hence providing the cell the ability to produce the recombinant protein *in vivo*. Alternatively, in instances where it is desirable to control, regulate, or decrease the amount of a particular recombinant PEAMT expressed in a particular transgenic cell, the invention also provides for the expression of 25 PEAMT-specific antisense mRNA. The use of antisense mRNA as a means of controlling or decreasing the amount of a given protein of interest in a cell is well known in the art.

Another aspect of the invention comprises a transgenic plant that expresses a gene or gene segment encoding one or more of the novel polypeptide compositions disclosed herein. As used herein, the term "transgenic plant" is intended to refer to a plant that has 30 incorporated DNA sequences, including but not limited to genes which are perhaps not

normally present, DNA sequences not normally transcribed into RNA or translated into a protein ("expressed"), or any other genes or DNA sequences which one desires to introduce into the non-transformed plant, such as genes which may normally be present in the non-transformed plant but which one desires to either genetically engineer or to have altered 5 expression.

It is contemplated that in some instances the genome of a transgenic plant of the present invention will have been augmented through the stable introduction of one or more PEAMT-encoding transgenes, either native, synthetically modified, or mutated. In some instances, more than one transgene will be incorporated into the genome of the transformed 10 host plant cell. Such is the case when more than one PEAMT-encoding DNA segment is incorporated into the genome of such a plant. In certain situations, it may be desirable to have one, two, three, four, or even more PEAMT polypeptides (either native or recombinantly-engineered) incorporated and stably expressed in the transformed transgenic plant.

15 A preferred gene that may be introduced includes, for example, a PEAMT polypeptide-encoding a DNA sequence from plant origin, such as those illustrated herein, and particularly one or more of those comprising one or more amino acid sequences described in SEQ ID NO:2 or SEQ ID NO:4.

Means for transforming a plant cell and the preparation of a transgenic cell line are 20 well known in the art, and are discussed herein. Vectors, plasmids, cosmids, YACs (yeast artificial chromosomes) and DNA segments for use in transforming such cells will, of course, generally comprise either the operons, genes, or gene-derived sequences of the present invention, either native, or synthetically-derived, and particularly those encoding the disclosed PEAMT polypeptides. These DNA constructs can further include structures such 25 as promoters, enhancers, polylinkers, or even gene sequences that have positively- or negatively-regulating activity upon the particular genes of interest as desired. The DNA segment or gene may encode either a native or modified PEAMT polypeptide, which will be expressed in the resultant recombinant cells, and/or which will impart an improved phenotype to the regenerated plant, in this case, by altering or modulating the biosynthesis or 30 lipid content in a transformed plant cell.

Such transgenic plants may be desirable for increasing lipid biosynthesis in a monocotyledonous or dicotyledonous plant, by incorporating into such a plant, a transgenic DNA segment encoding a PEAMT polypeptide. Particularly preferred plants include grains such as corn, wheat, millet, rye, rice, barley, and oats; legumes such as beans, soybeans, 5 peas; tubers such as potatoes; fiber crops such as flax and cotton; turf and pasture grasses; tobacco, sunflower, safflower, canola, ornamental plants; shrubs; trees; vegetables, berries, citrus, fruits, cacti, succulents, and other commercially-important crops including garden, floral, and houseplants.

In a related aspect, the present invention also encompasses a seed produced by the 10 transformed plant, a progeny from such seed, and a seed produced by the progeny of the original transgenic plant, produced in accordance with the above process. Such progeny and seeds will have one or more PEAMT transgene(s) stably incorporated into its genome, and such progeny plants will inherit the traits afforded by the introduction of a stable transgene in Mendelian fashion. All such transgenic plants having incorporated into their genome 15 transgenic DNA segments encoding one or more PEAMT polypeptides or polypeptides are aspects of this invention. Particularly preferred transgenes for the practice of the invention include nucleic acid segments comprising one or more PEAMT gene(s).

## 2.12 RECOMBINANT PROTEIN EXPRESSION

20 In other embodiments, it is contemplated that certain advantages will be gained by positioning the coding DNA segment under the control of a recombinant, or heterologous, promoter. As used herein, a recombinant or heterologous promoter is intended to refer to a promoter that is not normally associated with a DNA segment encoding a PEAMT polypeptide or peptide in its natural environment. Such promoters may include promoters 25 normally associated with other genes, and/or promoters isolated from any bacterial, viral, eukaryotic, or plant cell. Naturally, it will be important to employ a promoter that effectively directs the expression of the DNA segment in the cell type, organism, or even animal, chosen for expression. The use of promoter and cell type combinations for protein expression is generally known to those of skill in the art of molecular biology; for example, see Sambrook 30 *et al.*, 1989. The promoters employed may be constitutive, or inducible, and can be used

under the appropriate conditions to direct high level expression of the introduced DNA segment, such as is advantageous in modulating lipid biosynthesis or lipid composition of a cell expressing such genetic constructs. Appropriate promoter systems contemplated for use in high-level expression include, but are not limited to, the *Pichia* expression vector system 5 (Pharmacia LKB Biotechnology).

In connection with expression embodiments to prepare recombinant proteins and peptides, it is contemplated that longer DNA segments will most often be used, with DNA segments encoding the entire peptide sequence being most preferred. However, it will be appreciated that the use of shorter DNA segments to direct the expression of PEAMT 10 peptides or epitopic core regions, such as may be used to generate anti-PEAMT antibodies, also falls within the scope of the invention. DNA segments that encode peptide antigens of about 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30 15 or so amino acids, and up to and including those of about 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, or 100 or so amino acids in length, or more preferably, from 9 to about 80 or 20 so amino acids in length, or even more preferably, from about 11 to about 60 or so amino acids in length, or still more preferably, from 13 to about 40 or so amino acids in length, are contemplated to be particularly useful. Such peptide epitopes may be amino acid sequences 15 that comprise contiguous amino acid sequence from SEQ ID NO:2 or SEQ ID NO:4.

Preferably for recombinant protein expression, the polynucleotide comprises at least a 20 first heterologous promoter that directs the expression of the encoded polypeptide in the host cell. The polypeptide may be operably linked to one or more targeting sequences or signal peptides to permit expression of the polypeptide product in a particular cellular location or cell type.

In one embodiment, the invention encompasses a method of using a nucleic acid 25 segment of the present invention that encodes at least a first PEAMT polypeptide. The method generally comprises the steps of: (a) preparing a recombinant vector in which an - encoding nucleic acid segment is positioned under the control of a promoter; (b) introducing the recombinant vector into a host cell; (c) culturing the host cell under conditions effective to allow expression of the polypeptide encoded by the sequence; and (d) obtaining the 30 expressed protein or peptide.

A wide variety of ways are available for introducing a suitable nucleic acid segment into the microorganism or eukaryotic host under conditions that allow for stable maintenance and expression of the nucleic acid segment that encodes the polypeptide. One can provide for DNA constructs that include the transcriptional and translational regulatory signals for 5 expression of the PEAMT sequence, the sequence under their regulatory control and a DNA sequence homologous with a sequence in the host organism, whereby integration will occur, and/or a replication system that is functional in the host, whereby integration or stable maintenance will occur.

The transcriptional initiation signals will preferably include at least a first promoter 10 and at least a first transcriptional initiation start site. In some instances, it may be desirable to provide for regulative expression of the enzyme, where expression of the polypeptide will only occur after release into the environment. This can be achieved with operators or a region binding to an activator or enhancers, which are capable of induction upon a change in the physical or chemical environment of the microorganisms. For example, a temperature 15 sensitive regulatory region may be employed, where the organisms may be grown up in the laboratory without expression of a polypeptide, but upon release into the environment, expression would begin. Other techniques may employ a specific nutrient medium in the laboratory, which inhibits the expression of the polypeptide, where the nutrient medium in the environment would allow for expression of the polypeptide. For translational initiation, a 20 ribosomal binding site and an initiation codon will be present.

Various manipulations may be employed for enhancing the expression of the messenger RNA, particularly by using an active promoter, as well as by employing sequences, which enhance the stability of the messenger RNA. The transcriptional and translational termination region will involve stop codon(s), a terminator region, and 25 optionally, a polyadenylation signal. A hydrophobic "leader" sequence may be employed at the amino terminus of the translated polypeptide sequence in order to promote secretion of the protein across the inner membrane.

In the direction of transcription, namely in the 5' to 3' direction of the coding or sense sequence, the construct will involve the transcriptional regulatory region, if any, and the 30 promoter, where the regulatory region may be either 5' or 3' of the promoter, the ribosomal

binding site, the initiation codon, the structural gene having an open reading frame in phase with the initiation codon, the stop codon(s), the polyadenylation signal sequence, if any, and the terminator region. This sequence as a double strand may be used by itself for transformation of a microorganism host, but will usually be included with a nucleotide sequence involving a marker, where the second nucleotide sequence may be joined to the toxin expression construct during introduction of the nucleotide into the host.

By a marker is intended a structural gene which provides for selection of those hosts which have been modified or transformed. The marker will normally provide for selective advantage, for example, providing for biocide resistance, *e.g.*, resistance to antibiotics or heavy metals; complementation, so as to provide prototropy to an auxotrophic host, or the like. Preferably, complementation is employed, so that the modified host may not only be selected, but may also be competitive in the field. One or more markers may be employed in the development of the constructs, as well as for modifying the host. The organisms may be further modified by providing for a competitive advantage against other wild-type microorganisms in the field. For example, genes expressing metal chelating agents, *e.g.*, siderophores may be introduced into the host along with the structural gene expressing the toxin. In this manner, the enhanced expression of a siderophore may provide for a competitive advantage for the PEAMT-producing host, so that it may effectively compete with the wild-type microorganisms and stably occupy a niche in the environment.

Where no functional replication system is present, the construct will also include a sequence of at least 50 base pairs (bp), preferably at least about 100 bp, more preferably at least about 1000 bp, and usually not more than about 2000 bp of a sequence homologous with a sequence in the host. In this way, the probability of legitimate recombination is enhanced, so that the gene will be integrated into the host and stably maintained by the host. Desirably, the PEAMT gene will be in close proximity to the gene providing for complementation as well as the gene providing for the competitive advantage. Therefore, in the event that a toxin gene is lost, the resulting organism will be likely to also lose the complementing gene and/or the gene providing for the competitive advantage, so that it will be unable to compete in the environment with the gene retaining the intact construct.

A large number of transcriptional regulatory regions are available from a wide variety of microorganism hosts, such as bacteria, bacteriophage, cyanobacteria, algae, fungi, and the like. Various transcriptional regulatory regions include the regions associated with the *trp* gene, *lac* gene, *gal* gene, the  $\lambda_L$  and  $\lambda_R$  promoters, the *tac* promoter, the naturally-occurring 5 promoters associated with the  $\delta$ -endotoxin gene, where functional in the host. See for example, U. S. Patents 4,332,898; 4,342,832; and 4,356,270 (each of which is specifically incorporated herein by reference). The termination region may be the termination region normally associated with the transcriptional initiation region or a different transcriptional initiation region, so long as the two regions are compatible and functional in the host.

10 Where stable episomal maintenance or integration is desired, a plasmid will be employed which has a replication system that is functional in the host. The replication system may be derived from the chromosome, an episomal element normally present in the host or a different host, or a replication system from a virus that is stable in the host. A large number of plasmids are available, such as pBR322, pACYC184, RSF1010, pR01614, and the 15 like. See for example, Olson *et al.* (1982); Bagdasarian *et al.* (1981), Baum *et al.*, 1990, and U. S. Patents 4,356,270; 4,362,817; 4,371,625, and 5,441,884, each incorporated specifically herein by reference.

20 The PEAMT sequence can be introduced between the transcriptional and translational initiation region and the transcriptional and translational termination region, so as to be under the regulatory control of the initiation region. This construct will be included in a plasmid, which will include at least one replication system, but may include more than one, where one replication system is employed for cloning during the development of the plasmid and the second replication system is necessary for functioning in the ultimate host. In addition, one or more markers may be present, which have been described previously. Where integration 25 is desired, the plasmid will desirably include a sequence homologous with the host genome.

The transformants can be isolated in accordance with conventional ways, usually employing a selection technique, which allows for selection of the desired organism as against unmodified organisms or transferring organisms, when present. The transformants then can be tested for enzymatic activity. If desired, unwanted or ancillary DNA sequences 30 may be selectively removed from the recombinant bacterium by employing site-specific

recombination systems, such as those described in U. S. Patent 5,441,884 (specifically incorporated herein by reference).

In accordance with the present invention, nucleic acid sequences include and are not limited to DNA, including and not limited to cDNA and genomic DNA, genes; RNA, including and not limited to mRNA and tRNA; antisense sequences, PNAs (peptide nucleic acids), nucleosides, and suitable nucleic acid sequences such as those set forth herein, as well as alterations in the nucleic acid sequences including alterations, deletions, mutations, and homologs capable of expressing the PEAMT polypeptides and peptide fragments of the present invention.

As such the present invention also concerns DNA segments, that are free from total genomic DNA and that encode the novel plant-derived polypeptides disclosed herein. DNA segments encoding these peptide species may prove to encode proteins, polypeptides, subunits, functional domains, and the like of PEAMT-related or other non-related gene products. In addition these DNA segments may be synthesized entirely *in vitro* using methods that are well known to those of skill in the art.

As used herein, the term "DNA segment" refers to a DNA molecule that has been isolated free of total genomic DNA of a particular species. Therefore, a DNA segment encoding a PEAMT polypeptide or peptide refers to a DNA segment that contains PEAMT coding sequences yet is isolated away from, or purified free from, total genomic DNA of the species from which the DNA segment was obtained. Included within the term "DNA segment", are DNA segments and smaller fragments of such segments, and also recombinant vectors, including, for example, plasmids, cosmids, phagemids, phage, viruses, and the like.

Similarly, a DNA segment comprising an isolated or purified PEAMT polypeptide-encoding gene refers to a DNA segment which may include in addition to peptide encoding sequences, certain other elements such as, regulatory sequences, isolated substantially away from other naturally occurring genes or protein-encoding sequences. In this respect, the term "gene" is used for simplicity to refer to a functional protein-, polypeptide- or peptide-encoding unit. As will be understood by those in the art, this functional term includes both genomic sequences, operon sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides or peptides.

"Isolated substantially away from other coding sequences" means that the gene of interest, in this case, a gene encoding a plant PEAMT polypeptide, forms the significant part of the coding region of the DNA segment, and that the DNA segment does not contain large portions of naturally-occurring coding DNA, such as large chromosomal fragments or other 5 functional genes or operon coding regions. Of course, this refers to the DNA segment as originally isolated, and does not exclude genes, recombinant genes, synthetic linkers, or coding regions later added to the segment by the hand of man.

It will also be understood that amino acid and nucleic acid sequences may include additional residues, such as additional N- or C-terminal amino acids or 5' or 3' sequences, 10 and yet still be essentially as set forth in one of the sequences disclosed herein, so long as the sequence meets the criteria set forth above, including the maintenance of biological protein activity where PEAMT or  $\Delta$ PEAMT protein expression is concerned. The addition of terminal sequences particularly applies to nucleic acid sequences that may, for example, include various non-coding sequences flanking either of the 5' or 3' portions of the PEAMT 15 or  $\Delta$ PEAMT coding region or may include various internal sequences, *i.e.*, introns, which are known to occur within eukaryotic genes.

The nucleic acid segments of the present invention, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other 20 coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol.

Recombinant vectors and isolated DNA segments may therefore variously include the 25 peptide-coding regions themselves, coding regions bearing selected alterations or modifications in the basic coding region, or they may encode larger polypeptides that nevertheless include these peptide-coding regions or may encode biologically functional equivalent proteins or peptides that have variant amino acids sequences.

The DNA segments of the present invention encompass biologically functional, 30 equivalent peptides. Such sequences may arise as a consequence of codon redundancy and

functional equivalency that are known to occur naturally within nucleic acid sequences and the proteins thus encoded. Alternatively, functionally-equivalent proteins or peptides may be created *via* the application of recombinant DNA technology, in which changes in the protein structure may be engineered, based on considerations of the properties of the amino acids 5 being exchanged. Changes designed by man may be introduced through the application of site-directed mutagenesis techniques, *e.g.*, to introduce improvements to the antigenicity of the protein or to test mutants in order to examine activity at the molecular level.

If desired, one may also prepare fusion proteins and peptides, *e.g.*, where the peptide-coding regions are aligned within the same expression unit with other proteins or peptides 10 having desired functions, such as for purification or immunodetection purposes (*e.g.*, proteins that may be purified by affinity chromatography and enzyme label coding regions, respectively).

Recombinant vectors form further aspects of the present invention. Particularly useful vectors are contemplated to be those vectors in which the coding portion of the DNA 15 segment, whether encoding a full-length protein or smaller peptide, is positioned under the control of a promoter. The promoter may be in the form of the promoter that is naturally associated with a gene encoding peptides of the present invention, as may be obtained by isolating the 5' non-coding sequences located upstream of the coding segment or exon, for example, using recombinant cloning and/or PCR<sup>TM</sup> technology, in connection with the 20 compositions disclosed herein.

### 3. BRIEF DESCRIPTION OF THE DRAWINGS

The drawings form part of the present specification and are included to further demonstrate certain aspects of the present invention. The invention may be better understood 25 by reference to one or more of these drawings in combination with the detailed description of specific embodiments presented herein.

**FIG. 1A** illustrates the biogenesis of choline moieties in plants, and the complementation strategy used to identify PEAMT. Shown are the pathways found in leaves and other vegetative tissues of higher plants.

**FIG. 1B** illustrates the biogenesis of choline moieties in *S. pombe*, and the complementation strategy used to identify PEAMT. Shown is the native pathway in *S. pombe* (light arrows) and the bypass route (heavy arrows) that allows PEAMT to substitute for the defective *cho2* gene product.

5 **FIG. 2A** illustrates expression of spinach PEAMT and  $\Delta$ PEAMT cDNAs in extracts of *S. pombe* *cho2*<sup>-</sup> mutant cells. Shown is PEAMT activity in desalted extracts from unsalinized spinach leaves (sp) and strains 972 (h<sup>s</sup>) (wt), Bx22 (*cho2*<sup>-</sup>), and Bx22 expressing PEAMT (P) or  $\Delta$ PEAMT ( $\Delta$ P). Assays contained 50 nmol P-EA, 44 nmol [*methyl*<sup>14</sup>C]AdoMet, and 15-710  $\mu$ g of protein. Data are means  $\pm$ S.E. (n=2 or 3). P-MME is the 10 principal product (>90%) formed in these assays, as shown in the *inset*, which is an autoradiograph of a TLC separation of a [<sup>33</sup>P]P-MME standard (s) and the reaction product from representative assay mixtures.

15 **FIG. 2B** shows a schematic representation of the complementing cDNAs. PEAMT is the full-length cDNA, containing two sets of the methyltransferase motifs I, post-I (p-I), II and III.  $\Delta$ PEAMT is truncated after Gly-286.

20 **FIG. 3** illustrates alignment of the deduced amino acid sequence of the *Spinacia oleracea* (spinach) PEAMT (So) with an *Arabidopsis* homolog (At) deduced from nucleotides 65132-67885 of P1 clone MEB5 (GenBank Accession No. AB019230). The *Arabidopsis* genomic DNA has 13 exons and 12 introns; the identity of the region including 25 amino acids 369-494 is authenticated by alignment with an EST (GenBank Accession No. N65610). The asterisk marks the residue (Gly-286) at which  $\Delta$ PEAMT is truncated.

25 **FIG 4A** illustrates evidence that PEAMT has three methyltransferase activities. Shown is an autoradiograph of a TLC separation of PEAMT reaction products. Extract (130  $\mu$ g protein) from Bx22 expressing PEAMT was incubated with 2 nmol P-EA and 7 nmol (400 nCi) [*methyl*<sup>14</sup>C]AdoMet; samples were removed at intervals, stopped and deproteinized with acetone, fractionated by ion exchange and separated in TLC system 1. The positions of P-MME, P-DME and P-Cho zones and the origin (ori) are indicated.

30 **FIG 4B** shows the progress curve of the PEAMT reaction. Extract (15  $\mu$ g protein) from Bx22 cells expressing PEAMT was incubated with 5 nmol P-EA and 27 nmol (400 nCi) [*methyl*<sup>14</sup>C]AdoMet. Samples were removes at the indicated times, stopped by

freezing, fractionated by ion exchange and separated in TLC system 1. The P-MME, P-DME and P-Cho zones were located by autoradiography, scraped and counted. Radioactivity in P-DME and P-Cho zones was corrected for spillover from the P-MME zone. All data were adjusted for recovery from the TLC plate. ●, P-MME; ○, P-DME; X, P-Cho.

5 **FIG. 5A** illustrates evidence that PEAMT and ΔPEAMT have distinct methyltransferase activities. Shown is an autoradiograph of a TLC separation of the reaction products of wild type PEAMT (P) and the truncated enzyme ΔPEAMT (ΔP). Extracts (200 μg protein) from Bx22 cells expressing PEAMT or ΔPEAMT were incubated with 5 nmol P-EA and 27 nmol (100 nCi) [*methyl*-<sup>14</sup>C]AdoMet for 3 hr. The positions of P-MME and P-10 Cho zones and the origin (ori) are indicated.

10 **FIG. 5B** illustrates that PEAMT AND ΔPEAMT enzyme activity is inhibited by P-Cho. Shown is the PEAMT activity assayed in the presence of increasing P-Cho concentration. Extracts from Bx22 cells expressing PEAMT (25 μg protein) or ΔPEAMT (50 μg protein) were incubated for 10 min with 50 nmol P-EA, 170 nmol [*methyl*-<sup>14</sup>C]AdoMet and various concentrations of the disodium salt of P-Cho. Activities in the absence of P-Cho were 737 ±18 and 200 ±10 pkat mg<sup>-1</sup> protein for PEAMT and ΔPEAMT, respectively. Data are means of duplicates. ●, PEAMT; ○, ΔPEAMT.

15 **FIG. 6** illustrates the effect of salinization on PEAMT expression in spinach leaves. Shown is PEAMT activity, using assays that contained 50 nmol P-EA, 200 nmol [*methyl*-<sup>14</sup>C]AdoMet and 100 μg of protein, and which were incubated for 30 min. Data are means ± S.E. (n = 3).

#### 4. DESCRIPTION OF ILLUSTRATIVE EMBODIMENTS

##### 4.1 TRANSFORMED HOST CELLS AND TRANSFORMATION METHODS

20 In one embodiment, a bacterial cell, a cyanobacterial cell, a fungal cell, a yeast cell, or a plant cell transformed with at least a first PEAMT-encoding gene-containing expression vector represents an important aspect of the present invention. Furthermore, transgenic plants and the progeny and seeds derived from such a transformed or transgenic plant are also important aspects of this invention.

Such transformed host cells are often desirable for use in the expression of the various DNA gene constructs disclosed herein. In some aspects of the invention, it is often desirable to modulate, regulate, or otherwise control the expression of the gene segments disclosed herein. Such methods are routine to those of skill in the molecular genetic arts. Typically, 5 when increased or over-expression of a particular gene is desired, various manipulations may be employed for enhancing the expression of the messenger RNA, particularly by using an active promoter, as well as by employing sequences, which enhance the stability of the messenger RNA in the particular transformed host cell.

Typically, the initiation and translational termination region will involve stop 10 codon(s), a terminator region, and optionally, a polyadenylation signal. In the direction of transcription, namely in the 5' to 3' direction of the coding or sense sequence, the construct will involve the transcriptional regulatory region, if any, and the promoter, where the regulatory region may be either 5' or 3' of the promoter, the ribosomal binding site, the initiation codon, the structural gene having an open reading frame in phase with the initiation 15 codon, the stop codon(s), the polyadenylation signal sequence, if any, and the terminator region. This sequence as a double strand may be used by itself for transformation of a microorganism host, but will usually be included with a DNA sequence involving a marker, where the second DNA sequence may be joined to the expression construct during introduction of the DNA into the host.

Where no functional replication system is present, the construct will also preferably 20 include a sequence of at least about 40 or 50 base pairs (bp) or so, preferably at least about 90 to about 100 or so bp, and usually not more than about 500 to about 1000 or so bp of a sequence homologous with a sequence in the host. In this way, the probability of legitimate recombination is enhanced, so that the gene will be integrated into the host and stably 25 maintained by the host. Desirably, the PEAMT- or  $\Delta$ PEAMT-encoding gene-promoter construct will be in close proximity to the gene providing for complementation as well as the gene providing for the competitive advantage. Therefore, in the event that a PEAMT-or  $\Delta$ PEAMT-encoding gene is lost, the resulting organism will be likely to also lose the PEAMT and  $\Delta$ PEAMT gene and/or the gene providing for the competitive advantage, so that 30 it will be unable to compete in the environment with the gene retaining the intact construct.

The PEAMT- or  $\Delta$ PEAMT-encoding gene can be introduced between the transcriptional and translational initiation region and the transcriptional and translational termination region, so as to be under the regulatory control of the initiation region. This construct will be included in a plasmid, which will include at least one replication system, 5 but may include more than one, where one replication system is employed for cloning during the development of the plasmid and the second replication system is necessary for functioning in the ultimate host. In addition, one or more markers may be present, which have been described previously. Where integration is desired, the plasmid will desirably include a sequence homologous with the host genome.

10 Alternatively, the left and right T-DNA borders from the Ti plasmid may be used when integration is desired using *A. tumefaciens* vectors for plant transformation. The transformants can be isolated in accordance with conventional ways, usually employing a selection technique, which allows for selection of the desired organism as against unmodified organisms or transferring organisms, when present. The transformants then can be tested for 15 presence of the genetic construct.

Genes or other nucleic acid segments, as disclosed herein, can be inserted into host 20 cells using a variety of techniques that are well known in the art. Five general methods for delivering a nucleic segment into cells have been described: (1) chemical methods (Graham and VanDerEb, 1973); (2) physical methods such as microinjection (Capecchi, 1980), electroporation (U. S. Patent No. 5,472,869; Wong and Neumann, 1982; Fromm *et al.*, 1985), microprojectile bombardment (Wang *et al.*, 1988; Tomes *et al.*, 1990; Vain *et al.*, 1993; U. S. Patent No. 5,874,265, specifically incorporated herein by reference in its 25 entirety), “gene gun” (Hilber *et al.*, 1994; Yang *et al.*, 1990); (3) viral vectors (Clapp, 1993; Danos and Heard, 1992; Eglitis and Anderson, 1988); (4) receptor-mediated mechanisms (Curiel *et al.*, 1991; Wagner *et al.*, 1992); and (5) bacterial-mediated delivery such as *A. tumefaciens* transformation (Smith and Hood, 1995).

For example, a large number of cloning vectors comprising a replication system in *E. coli* and a marker that permits selection of the transformed cells are available for preparation 30 for the insertion of foreign genes into higher organisms, including plants. The vectors comprise, for example, plasmids (such as pBR322, pUC series, M13mp series, pACYC184,

etc), cosmids, phage, and/or phagemids and the like. Accordingly, the disclosed polynucleotides can be inserted into a given vector at a suitable restriction site. The resulting plasmid may be used, for example, to transform bacterial cells such as *E. coli* or *A. tumefaciens*. The bacterial cells are then cultivated in a suitable nutrient medium, harvested 5 and lysed. The plasmid is recovered. Sequence analysis, restriction analysis, electrophoresis, and other biochemical-molecular biological methods are generally carried out as methods of analysis. After each manipulation, the DNA sequence used can be cleaved and joined to the next DNA sequence. Each plasmid sequence can be cloned in the same or other plasmids. Depending on the method of inserting desired genes into the plant, other 10 DNA sequences may be necessary.

Methods for DNA transformation of plant cells include *Agrobacterium*-mediated plant transformation, protoplast transformation, gene transfer into pollen, injection into reproductive organs, injection into immature embryos and particle bombardment. Each of these methods has distinct advantages and disadvantages. Thus, one particular method of 15 introducing genes into a particular plant strain may not necessarily be the most effective for another plant strain, but it is well known which methods are useful for a particular plant strain.

Suitable methods are believed to include virtually any method by which DNA can be introduced into a cell, such as by *Agrobacterium* infection, direct delivery of DNA such as, 20 for example, by PEG-mediated transformation of protoplasts (Omirulleh *et al.*, 1993), by desiccation/inhibition-mediated DNA uptake, by electroporation, by agitation with silicon carbide fibers, by acceleration of DNA coated particles, etc. In certain embodiments, acceleration methods are preferred and include, for example, microprojectile bombardment and the like.

25 Technology for introduction of DNA into cells is well known to those of skill in the art, and described hereinbelow in detail. Likewise, a large number of techniques are available for inserting DNA into a plant host cell. Those techniques include transformation with T-DNA using *A. tumefaciens* or *A. rhizogenes* as transformation agent, fusion, injection, or electroporation as well as other possible methods. If agrobacteria are used for the 30 transformation, the DNA to be inserted has to be cloned into special plasmids, namely either

into an intermediate vector or into a binary vector. The intermediate vectors can be integrated into the Ti or Ri plasmid by homologous recombination owing to sequences that are homologous to sequences in the T-DNA. The Ti or Ri plasmid also comprises the *vir* region necessary for the transfer of the T-DNA.

5        Intermediate vectors cannot replicate themselves in agrobacteria. The intermediate vector can be transferred into *A. tumefaciens* by means of a helper plasmid (conjugation). Binary vectors can replicate themselves both in *E. coli* and in agrobacteria. They comprise a selection marker gene and a linker or polylinker that are framed by the right and left T-DNA border regions. They can be transformed directly into agrobacteria (Holsters *et al.*, 1978).

10      The agrobacterium used as host cell is to comprise a plasmid carrying a *vir* region. The *vir* region is necessary for the transfer of the T-DNA into the plant cell. Additional t-DNA may be contained. The bacterium so transformed is used for the transformation of plant cells. Plant explants can advantageously be cultivated with *A. tumefaciens* or *A. rhizogenes* for the transfer of the DNA into the plant cell. Whole plants can then be regenerated from the 15 infected plant material (for example, pieces of leaf, segments of stalk, roots, but also protoplasts or suspension-cultivated cells) in a suitable medium, which may contain antibiotics or biocides for selection. The plants so obtained can then be tested for the presence of the inserted DNA. No special demands are made of the plasmids in the case of injection and electroporation. It is possible to use ordinary plasmids, such as, for example,

20      pUC, pRK, pBluescript, *etc.* and/or their derivatives. If, for example, the Ti or Ri plasmid is used for the transformation of the plant cell, then at least the right border, but often the right and the left border of the Ti or Ri plasmid T-DNA, has to be joined as the flanking region of the genes to be inserted. The use of T-DNA for the transformation of plant cells has been intensively researched and sufficiently described in Eur. Pat. Appl. No. EP 120516; Hockema 25 (1985); An *et al.*, 1985, Herrera-Estrella *et al.*, (1983), Bevan *et al.*, (1983), and Klee *et al.*, (1985).

A particularly useful Ti plasmid cassette vector for transformation of dicotyledonous plants consists of the enhanced CaMV35S promoter (EN-35S) and the 3' end including polyadenylation signals from a soybean gene encoding the  $\alpha'$ -subunit of  $\beta$ -conglycinin.

Between these two elements is a multilinker containing multiple restriction sites for the insertion of genes of interest.

The vector preferably contains a segment of pBR322 which provides an origin of replication in *E. coli* and a region for homologous recombination with the disarmed T-DNA in *Agrobacterium* strain ACO; the *oriV* region from the broad host range plasmid RK1; the streptomycin/spectinomycin resistance gene from Tn7; and a chimeric NPTII gene, containing the CaMV35S promoter and the nopaline synthase (NOS) 3'-end, which provides kanamycin resistance in transformed cells.

Optionally, the enhanced CaMV35S promoter may be replaced with the 1.5-kb mannopine synthase (MAS) promoter (Velten *et al.*, 1984). After incorporation of a DNA construct into the vector, it is introduced into *A. tumefaciens* strain ACO that contains a disarmed Ti plasmid. Cointegrate Ti plasmid vectors are selected and subsequently may be used to transform a dicotyledonous plant.

*A. tumefaciens* ACO is a disarmed strain similar to pTiB6SE described by Fraley *et al.*, (1985). For construction of ACO the starting *Agrobacterium* strain was the strain A208 that contains a nopaline-type Ti plasmid. The Ti plasmid was disarmed in a manner similar to that described by Fraley *et al.* (1985) so that essentially all of the native T-DNA was removed except for the left border and a few hundred base pairs of T-DNA inside the left border. The remainder of the T-DNA extending to a point just beyond the right border was replaced with a novel piece of DNA including (from left to right) a segment of pBR322, the *oriV* region from plasmid RK2, and the kanamycin resistance gene from Tn601. The pBR322 and *oriV* segments are similar to these segments and provide a region of homology for cointegrate formation.

Once the inserted DNA has been integrated in the genome, it is relatively stable there and, as a rule, does not come out again. It normally contains a selection marker that confers on the transformed plant cells resistance to a biocide or an antibiotic, such as kanamycin, G 418, bleomycin, hygromycin, or chloramphenicol, *inter alia*. The individually employed marker should accordingly permit the selection of transformed cells rather than cells that do not contain the inserted DNA.

#### 4.1.1 ELECTROPORATION

The application of brief, high-voltage electric pulses to a variety of animal and plant cells leads to the formation of nanometer-sized pores in the plasma membrane. DNA is taken directly into the cell cytoplasm either through these pores or as a consequence of the 5 redistribution of membrane components that accompanies closure of the pores. Electroporation can be extremely efficient and can be used both for transient expression of clones genes and for establishment of cell lines that carry integrated copies of the gene of interest. Electroporation, in contrast to calcium phosphate-mediated transfection and protoplast fusion, frequently gives rise to cell lines that carry one, or at most a few, integrated 10 copies of the foreign DNA.

The introduction of DNA by electroporation is well-known to those of skill in the art (see *e.g.*, U. S. Patent No. 5,324,253, specifically incorporated herein by reference in its entirety). In this method, certain cell wall-degrading enzymes, such as pectin-degrading enzymes, are employed to render the target recipient cells more susceptible to transformation 15 by electroporation than untreated cells. Alternatively, recipient cells are made more susceptible to transformation, by mechanical wounding. To effect transformation by electroporation one may employ either friable tissues such as a suspension culture of cells (U. S. Patent No. 5,484,956; U. S. Patent No. 5,886,244, each of which is specifically incorporated herein by reference in its entirety), or embryogenic callus (U. S. Patent No. 20 5,405,765, each of which is specifically incorporated herein by reference in its entirety), or alternatively, one may transform immature embryos or other organized tissues directly. One would partially degrade the cell walls of the chosen cells by exposing them to pectin-degrading enzymes (pectolyases) or mechanically wounding in a controlled manner. Such 25 cells would then be the recipient of DNA transfer by electroporation, which may be carried out at this stage, and transformed cells then identified by a suitable selection or screening protocol dependent on the nature of the newly incorporated DNA.

#### 4.1.2 MICROPROJECTILE BOMBARDMENT

A further advantageous method for delivering transforming DNA segments to plant 30 cells is microprojectile bombardment. In this method, particles may be coated with nucleic

acids and delivered into cells by a propelling force. Exemplary particles include those comprised of tungsten, gold, platinum, and the like.

An advantage of microprojectile bombardment, in addition to it being an effective means of reproducibly stably transforming monocots, is that neither the isolation of 5 protoplasts (Cristou *et al.*, 1988) nor the susceptibility to *Agrobacterium* infection is required. An illustrative embodiment of a method for delivering DNA into maize cells by acceleration is a Biolistics Particle Delivery System, which can be used to propel particles coated with DNA or cells through a screen, such as a stainless steel or Nytex screen, onto a filter surface covered with corn cells cultured in suspension. The screen disperses the 10 particles so that they are not delivered to the recipient cells in large aggregates. It is believed that a screen intervening between the projectile apparatus and the cells to be bombarded reduces the size of projectiles aggregate and may contribute to a higher frequency of transformation by reducing damage inflicted on the recipient cells by projectiles that are too large.

15 For the bombardment, cells in suspension are preferably concentrated on filters or solid culture medium. Alternatively, immature embryos or other target cells may be arranged on solid culture medium. The cells to be bombarded are positioned at an appropriate distance below the macroprojectile stopping plate. If desired, one or more screens are also positioned 20 between the acceleration device and the cells to be bombarded. Through the use of techniques set forth herein one may obtain up to 1000 or more foci of cells transiently expressing a marker gene. The number of cells in a focus which express the exogenous gene product 48 h post-bombardment often range from 1 to 10 and average 1 to 3.

In bombardment transformation, one may optimize the prebombardment culturing 25 conditions and the bombardment parameters to yield the maximum numbers of stable transformants. Both the physical and biological parameters for bombardment are important in this technology. Physical factors are those that involve manipulating the DNA/microprojectile precipitate or those that affect the flight and velocity of either the macro- or microprojectiles. Biological factors include all steps involved in manipulation of cells before and immediately after bombardment, the osmotic adjustment of target cells to 30 help alleviate the trauma associated with bombardment, and also the nature of the

transforming DNA, such as linearized DNA or intact supercoiled plasmids. It is believed that pre-bombardment manipulations are especially important for successful transformation of immature embryos.

Accordingly, it is contemplated that one may wish to adjust several of the 5 bombardment parameters in small-scale studies to fully optimize the conditions. One may particularly wish to adjust physical parameters such as gap distance, flight distance, tissue distance, and helium pressure. One may also minimize the trauma reduction factors (TRFs) by modifying conditions which influence the physiological state of the recipient cells and which may therefore influence transformation and integration efficiencies. For example, the 10 osmotic state, tissue hydration and the subculture stage or cell cycle of the recipient cells may be adjusted for optimum transformation. The execution of other routine adjustments will be known to those of skill in the art in light of the present disclosure.

#### 4.1.3 AGROBACTERIUM-MEDIATED TRANSFER

15 *Agrobacterium*-mediated transfer is a widely applicable system for introducing genes into plant cells because the DNA can be introduced into whole plant tissues, thereby bypassing the need for regeneration of an intact plant from a protoplast. The use of *Agrobacterium*-mediated plant integrating vectors to introduce DNA into plant cells is well known in the art. See, for example, the methods described (Fraley *et al.*, 1985; Rogers *et al.*, 20 1988). Further, the integration of the Ti-DNA is a relatively precise process resulting in few rearrangements. The region of DNA to be transferred is defined by the border sequences, and intervening DNA is usually inserted into the plant genome as described (Spielmann *et al.*, 1986; Jorgensen *et al.*, 1987).

25 Modern *Agrobacterium* transformation vectors are capable of replication in *E. coli* as well as *Agrobacterium*, allowing for convenient manipulations as described (Klee *et al.*, 1985). Moreover, recent technological advances in vectors for *Agrobacterium*-mediated gene transfer have improved the arrangement of genes and restriction sites in the vectors to facilitate construction of vectors capable of expressing various polypeptide-coding genes. The vectors described (Eichholtz *et al.*, 1987), have convenient multi-linker regions flanked 30 by a promoter and a polyadenylation site for direct expression of inserted polypeptide coding

genes and are suitable for present purposes. In addition, *Agrobacterium* containing both armed and disarmed Ti genes can be used for the transformations. In those plant strains where *Agrobacterium*-mediated transformation is efficient, it is the method of choice because of the facile and defined nature of the gene transfer.

5        *Agrobacterium*-mediated transformation of leaf disks and other tissues such as cotyledons and hypocotyls appears to be limited to plants that *Agrobacterium* naturally infects. *Agrobacterium*-mediated transformation is most efficient in dicotyledonous plants. Few monocots appear to be natural hosts for *Agrobacterium*, although transgenic plants have been produced in asparagus using *Agrobacterium* vectors as described (Bytebier *et al.*, 1987).

10      Therefore, commercially important cereal grains such as rice, corn, and wheat must usually be transformed using alternative methods (see *e.g.*, U. S. Patent No. 5,610,042, specifically incorporated herein by reference in its entirety).

15      A transgenic plant formed using *Agrobacterium* transformation methods typically contains a single gene on one chromosome. Such transgenic plants can be referred to as being heterozygous for the added gene. However, inasmuch as use of the word "heterozygous" usually implies the presence of a complementary gene at the same locus of the second chromosome of a pair of chromosomes, and there is no such gene in a plant containing one added gene as here, it is believed that a more accurate name for such a plant is an independent segregant, because the added, exogenous gene segregates independently 20 during mitosis and meiosis.

25      More preferred is a transgenic plant that is homozygous for the added structural gene; *i.e.* a transgenic plant that contains two added genes, one gene at the same locus on each chromosome of a chromosome pair. A homozygous transgenic plant can be obtained by sexually mating (selfing) an independent segregant transgenic plant that contains a single added gene, germinating some of the seed produced and analyzing the resulting plants produced for enhanced carboxylase activity relative to a control (native, non-transgenic) or an independent segregant transgenic plant.

30      It is to be understood that two different transgenic plants can also be mated to produce offspring that contain two or more independently segregating added, exogenous genes. Selfing of appropriate progeny can produce plants that are homozygous for both

added, exogenous genes that encode a polypeptide of interest. Backcrossing to a parental plant and out-crossing with a non-transgenic plant are also contemplated.

Transformation of plant protoplasts can be achieved using methods based on calcium phosphate precipitation, polyethylene glycol treatment, electroporation, and combinations of 5 these treatments (see, e.g., Potrykus *et al.*, 1985; Lorz *et al.*, 1985; Fromm *et al.*, 1985; Uchimiya *et al.*, 1986; Callis *et al.*, 1987; Marcotte *et al.*, 1988).

Application of these systems to different plant strains depends upon the ability to regenerate that particular plant strain from protoplasts. Illustrative methods for the 10 regeneration of cereals from protoplasts are described (Fujimura *et al.*, 1985; Toriyama *et al.*, 1986; Yamada *et al.*, 1986; Abdullah *et al.*, 1986).

To transform plant strains that cannot be successfully regenerated from protoplasts, other ways to introduce DNA into intact cells or tissues can be utilized. For example, 15 regeneration of cereals from immature embryos or explants can be effected as described (Vasil, 1988). In addition, "particle gun" or high-velocity microprojectile technology may also be utilized (Vasil, 1992).

Using that latter technology, DNA is carried through the cell wall and into the cytoplasm on the surface of small metal particles as described (Klein *et al.*, 1987; Klein *et al.*, 1988a; 1988b; McCabe *et al.*, 1988). The metal particles penetrate through several layers of cells and thus allow the transformation of cells within tissue explants.

20

#### 4.2 EXPRESSION VECTORS

The present invention also provides an expression vector comprising at least one PEAMT- or  $\Delta$ PEAMT-encoding gene-containing polynucleotide operably linked to an 25 inducible promoter. Thus, in one embodiment an expression vector is an isolated and purified DNA molecule comprising a PEAMT or a  $\Delta$ PEAMT coding region operably linked to a promoter that expresses the gene, which coding region is operatively linked to a transcription-terminating region, whereby the promoter drives the transcription of the coding region.

As used herein, the term "operatively linked" means that a promoter is connected to a 30 nucleic acid region encoding functional RNA in such a way that the transcription of that

functional RNA is controlled and regulated by that promoter. Means for operatively linking a promoter to a nucleic acid region encoding functional RNA are well known in the art.

The choice of which expression vector and ultimately to which promoter a polypeptide coding region is operatively linked depend directly on the functional properties 5 desired, *e.g.*, the location and timing of protein expression, and the host cell to be transformed. These are well known limitations inherent in the art of constructing recombinant DNA molecules. However, a vector useful in practicing the present invention is capable of directing the expression of the functional RNA to which it is operatively linked.

RNA polymerase transcribes a coding DNA sequence through a site where 10 polyadenylation occurs. Typically, DNA sequences located a few hundred base pairs downstream of the polyadenylation site serve to terminate transcription. Those DNA sequences are referred to herein as transcription-termination regions. Those regions are required for efficient polyadenylation of transcribed messenger RNA (mRNA).

A variety of methods have been developed to operatively link DNA to vectors *via* 15 complementary cohesive termini or blunt ends. For instance, complementary homopolymer tracts can be added to the DNA segment to be inserted and to the vector DNA. The vector and DNA segment are then joined by hydrogen bonding between the complementary homopolymeric tails to form recombinant DNA molecules.

#### 20 4.3 DNA SEGMENTS AS HYBRIDIZATION PROBES AND PRIMERS

In another aspect, DNA sequence information provided by the invention allows for the preparation of relatively short DNA (or RNA) sequences having the ability to specifically 25 hybridize to gene sequences of the selected polynucleotides disclosed herein. The ability of such nucleic acid probes to specifically hybridize to all or portions of one or more PEAMT- or  $\Delta$ PEAMT-encoding genes lends them particular utility in a variety of embodiments. Most importantly, the probes may be used in a variety of assays for detecting the presence of complementary sequences in a given sample, and in the identification of new species or genera of  $\Delta$ PEAMT- and PEAMT-encoding genes from a variety of host organisms.

In certain embodiments, it is advantageous to use oligonucleotide primers. The 30 sequence of such primers is designed using a polynucleotide of the present invention for use

in detecting, amplifying or mutating a defined segment of the disclosed PEAMT- and  $\Delta$ PEAMT-encoding genes (e.g., SEQ ID NO:1 and SEQ ID NO:3, respectively) from a sample using PCR<sup>TM</sup> technology. Segments of related  $\Delta$ PEAMT- and PEAMT-encoding genes from other species, and particularly from other related plant species may also be  
5 amplified by PCR<sup>TM</sup> using such primers.

To provide certain of the advantages in accordance with the present invention, a preferred nucleic acid sequence employed for hybridization studies or assays includes sequences that are complementary to at least about 31 to 50 or so long nucleotide stretch of a PEAMT-encoding gene sequence. A size of at least 31 nucleotides in length helps to ensure  
10 that the fragment will be of sufficient length to form a duplex molecule that is both stable and selective. Molecules having complementary sequences over stretches greater than 31 bases in length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-complementary  
15 stretches of about 31 to about 40 or 50 or so nucleotides, or even longer where desired. Such fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, by application of nucleic acid reproduction technology, such as the PCR<sup>TM</sup> technology of U. S. Patent No. 4,683,195, and U. S. Patent No. 4,683,202, (each specifically incorporated herein by reference in its entirety), or by excising selected DNA fragments from  
20 recombinant plasmids containing appropriate inserts and suitable restriction sites.

Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template or where one seeks to isolate  $\Delta$ PEAMT- and PEAMT-encoding gene sequences from related species, functional equivalents, or the like, less stringent hybridization conditions will typically be needed in  
25 order to allow formation of the heteroduplex. In these circumstances, one may desire to employ conditions such as about 0.15 M to about 0.9 M salt, at temperatures ranging from about 25°C to about 60°C. Naturally, these ranges would encompass hybridization conditions that employ temperatures of about 26°C, 27°C, 28°C, 29°C, 30°C, 31°C, 32°C, 33°C, 34°C, 35°C, 36°C, 37°C, 38°C, 39°C, 40°C, 41°C, 42°C, 43°C, 44°C, 45°C, 46°C,  
30 47°C, 48°C, 49°C, 50°C, 51°C, 52°C, 53°C, 54°C, 55°C, 56°C, 57°C, 58°C, 59°C, 60°C,

61°C, 62°C, 63°C, 64°C, 65°C, 66°C, 67°C, 68°C, 69°C, 70°C, 71°C, 72°C, 73°C, or 74°C, and/or conditions that employ a salt concentration of about 0.20 M, 0.25 M, 0.30 M, 0.35 M, 0.40 M, 0.45 M, 0.50 M, 0.55 M, 0.60 M, 0.65 M, 0.70 M, 0.75 M, 0.80 M or 0.85 M.

Cross-hybridizing species can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In any case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

In addition to the use in directing the expression of functional RNA of the present invention, the nucleic acid sequences contemplated herein also have a variety of other uses. For example, they also have utility as probes or primers in nucleic acid hybridization embodiments. As such, it is contemplated that nucleic acid segments that comprise a sequence region that consists of at least a 14 nucleotide long contiguous sequence that has the same sequence as, or is complementary to, a 14 nucleotide long contiguous DNA segment of one or more PEAMT-encoding genes will find particular utility. Longer contiguous identical or complementary sequences, *e.g.*, those of about 20, 21, 22, 23, 24, *etc.*, 30, 31, 32, 33, 34, *etc.*, 40, 41, 42, 43, 44, *etc.*, 50, 51, 52, 53, 54, *etc.*, 100, 200, 300, 400, 500, 600, 700, 800, 900, 1000, 1100, 1300, 1500, 2000, *etc.* (including all intermediate lengths and up to and including full-length sequences will also be of use in certain embodiments.

While the ability of such nucleic acid probes to specifically hybridize to PEAMT-encoding gene sequences makes them ideal for use in detecting the presence of complementary sequences in a given sample, other uses are also envisioned, including the use of the sequence information for the preparation of mutant species primers, synthetic gene sequences, gene fusions, and/or primers for use in preparing other PEAMT-encoding genetic constructs.

The use of a hybridization probe of about 14 or so nucleotides in length allows the formation of a duplex molecule that is both stable and selective. Molecules having contiguous complementary sequences over stretches of about 15, 16, 17, 18, 19, 20, 21, 22,

23, 24, 25, 26, 27, 28, 29, or 30 or more contiguous bases in length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-complementary stretches of about 31, 32, 33, 34, 35, 36, 5 37, 38, 39, or 40 or more contiguous nucleotides in length where desired. When longer polynucleotides are desired, one may employ nucleic acid segments having gene-complementary stretches of about 41, 42, 43, 44, 45, 46, 47, 48, 49, or even 50, 60, 70, 80, 90, or 100 or more contiguous nucleotides in length where desired.

Of course, fragments may also be obtained by other techniques such as, *e.g.*, by 10 mechanical shearing or by restriction enzyme digestion. Small nucleic acid segments or fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, as is commonly practiced using an automated oligonucleotide synthesizer. Also, fragments may be obtained by application of nucleic acid reproduction technology, such as the PCR™ technology of U. S. Patent No. 4,683,195 and U. S. Patent No. 4,683,202 15 (each of which is specifically incorporated herein by reference in its entirety), by introducing selected sequences into recombinant vectors for recombinant production, and by other recombinant DNA techniques generally known to those of skill in the art of molecular biology.

Accordingly, the nucleotide sequences of the invention may be used for their ability 20 to selectively form duplex molecules with complementary stretches of DNA fragments. Depending on the application envisioned, one may employ varying conditions of hybridization to achieve varying degrees of selectivity of probe towards target sequence. For applications requiring high selectivity, one will typically desire to employ relatively stringent conditions to form the hybrids, *e.g.*, one will select relatively low salt and/or high 25 temperature conditions, such as provided by about 0.02 M to about 0.15 M NaCl at temperatures of about 50°C to about 75°C. Naturally, these ranges would encompass hybridization conditions that employ temperatures of about 51°C, 52°C, 53°C, 54°C, 55°C, 56°C, 57°C, 58°C, 59°C, 60°C, 61°C, 62°C, 63°C, 64°C, 65°C, 66°C, 67°C, 68°C, 69°C, 70°C, 71°C, 72°C, 73°C, or 74°C, and/or conditions that employ a salt concentration of 30 about 0.03 M, 0.04 M, 0.05 M, 0.06 M, 0.07 M, 0.08 M, 0.09 M, 0.10 M, 0.11 M, 0.12 M,

0.13 M, or 0.14 M. Such selective conditions tolerate little, if any, mismatch between the probe and the template or target strand, and would be particularly suitable for isolating particular DNA segments that are highly homologous to one or more of the PEAMT sequences disclosed herein.

5 Detection of DNA segments *via* hybridization is well known to those of skill in the art, and the teachings of U. S. Patent No. 4,965,188 and U. S. Patent No. 5,176,995 (each of which is specifically incorporated herein by reference in its entirety) are exemplary of the methods of hybridization analyses. Teachings such as those found in the texts of Maloy *et al.*, 1994; Segal 1976; Prokop and Bajpai, 1991; and Kuby, 1994, are also particularly relevant.

10 In any case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired 15 results.

15 In certain embodiments, it will be advantageous to employ nucleic acid sequences of the present invention in combination with an appropriate means, such as a label, for determining hybridization. A wide variety of appropriate indicator means are known in the art, including fluorescent, radioactive, enzymatic or other ligands, such as avidin/biotin, 20 which are capable of giving a detectable signal. In preferred embodiments, one will likely desire to employ a fluorescent label or an enzyme tag, such as urease, alkaline phosphatase or peroxidase, instead of radioactive or other environmentally undesirable reagents. In the case of enzyme tags, colorimetric indicator substrates are known that can be employed to provide a means visible to the human eye or spectrophotometrically, to identify specific hybridization 25 with complementary nucleic acid-containing samples.

25 In general, it is envisioned that the hybridization probes described herein will be useful both as reagents in solution hybridization as well as in embodiments employing a solid phase. In embodiments involving a solid phase, the test nucleic acid (*e.g.*, DNA, PNA, or RNA) is adsorbed or otherwise affixed to a selected matrix or surface. This fixed, single-stranded nucleic acid is then subjected to specific hybridization with selected probes under

desired conditions. The selected conditions will depend on the particular circumstances based on the particular criteria required (depending upon, *e.g.*, the G+C content, type of target nucleic acid, source of nucleic acid, size of the target sequence, length of the hybridization probe, *etc.*). Following washing of the hybridized surface so as to remove 5 nonspecifically bound probe molecules, specific hybridization is detected, or even quantitated, by means of the label.

#### 4.4 PEAMT AND ΔPEAMT POLYNUCLEOTIDE CONSTRUCTS

An important embodiment of the present invention concerns genetic constructs, such 10 as plasmids, vectors, viruses, cosmids, and the like that comprise one or more of the PEAMT/ΔPEAMT-encoding polynucleotide sequences disclosed herein. Another important embodiment concerns genetic constructs that comprise one or more portions of a PEAMT- or ΔPEAMT-encoding polynucleotide or one or more regulatory regions of a PEAMT/ΔPEAMT-encoding polynucleotide sequence. Particularly important genetic 15 constructs include those used for the preparation of oligonucleotide probes, polynucleotide primers, recombinant proteins, peptides, or peptide epitopes, and those used for sequencing PEAMT/ΔPEAMT-encoding gene sequences and homologous polynucleotides. Other important genetic constructs include transformation vectors, viruses and the like used for the introduction of the disclosed gene sequences into a host cell, plant tissue, or plant for the 20 preparation of transgenic plants or transformed host cells that express the PEAMT/ΔPEAMT polypeptide when cultured under appropriate conditions.

A variety of genetic compositions may be used for preparation and delivery of the disclosed PEAMT/ΔPEAMT genetic constructs to selected recipient host cells, and particularly to selected plant host cells or tissues to ultimately produce transformed plants 25 and plant cell lines in accordance with the present invention. For example, polynucleotides in the form of vectors and plasmids, or linear nucleic acid fragments, in some instances containing only the particular polynucleotide to be expressed in the cell, tissue or animal, and the like, may be employed.

Vectors, plasmids, phagemids, cosmids, viral vectors, shuttle vectors, baculovirus 30 vectors, BACs (bacterial artificial chromosomes), PACs (plant artificial chromosomes),

YACs (yeast artificial chromosomes) and DNA segments for use in transforming cells with a nucleic acid construct of interest, are well known to those of skill in the microbiological and plant molecular biology arts. Typically such constructs generally comprise at least one promoter or other regulatory region that is operably linked to at least one or more of the 5 novel polynucleotides disclosed herein. These polynucleotide constructs may contain a cDNA, or one or more genes which one desires to introduce into a particular cell, cell line, tissue, or other suitable organism. Such polynucleotide constructs may also optionally include one or more structures such as inducible, constitutive, or tissue-specific promoters, one or more enhancers or enhancer elements, one or more polylinkers or multiple cloning 10 sites, or one or more regulatory sequences as may be desired. The polynucleotide segment or gene chosen for cellular introduction may encode the entire PEAMT protein such that the protein may be expressed in the resultant recombinant cells, or, alternatively, the nucleic acid constructs may contain portions of the coding region, or a PEAMT gene regulatory region alone or in combination with other gene sequences, or may even comprise one or more 15 antisense constructs, or ribozyme-encoding regions.

#### **4.5 METHODS FOR PREPARING MUTAGENIZED POLYNUCLEOTIDES**

In certain circumstances, it may be desirable to modify or alter one or more nucleotides in one or more of the sequences disclosed herein for the purpose of altering or 20 changing the transcriptional activity or other property of the sequence region. In general, the means and methods for mutagenizing a polynucleotide are well known to those of skill in the art. Modifications to such polynucleotides may be made by random, or site-specific mutagenesis procedures. The selected polynucleotide may be modified by altering its structure through the addition or deletion of one or more nucleotides within the sequence, or 25 may be modified by the addition of a cloning site, a polylinkers region, or by the preparation of a gene fusion or a protein fusion encoding polynucleotide. Means for preparing mutagenized polynucleotides are exemplified in a number of U. S. patents and in the scientific literature. For example, U. S. Patent No. 6,023,013 (specifically incorporated herein by reference in its entirety) provides a variety of methods for preparing mutagenized 30 polynucleotides.

#### 4.6 EXPRESSION OF TRANSGENES IN PLANTS

In many instances, the level of transcription of a particular transgene in a given host cell is not always indicative of the amount of protein being produced in the transformed host cell. This is often due to post-transcriptional processes, such as splicing, polyadenylation, appropriate translation initiation, and RNA stability that affect the ability of a transcript to produce protein. Such factors may also affect the stability and amount of mRNA produced from the given transgene. As such, it is often desirable to alter the post-translational events through particular molecular biology techniques. The inventors contemplate that in certain instances it may be desirable to alter the transcription and/or expression of the PEAMT/ΔPEAMT-encoding gene constructs of the present invention to increase, decrease, or otherwise regulate or control these constructs in particular host cells and/or transgenic plants.

##### 15 4.6.1 EFFICIENT INITIATION OF PROTEIN TRANSLATION

The 5'-untranslated leader (5'-UTL) sequence of eukaryotic mRNA plays a major role in translational efficiency. Many early chimeric transgenes using a viral promoter used an arbitrary length of viral sequence after the transcription initiation site and fused this to the AUG of the coding region. More recently studies have shown that the 5'-UTL sequence and the sequences directly surrounding the AUG can have a large effect in translational efficiency in host cells and particularly certain plant species and that this effect can be different depending on the particular cells or tissues in which the message is expressed.

In most eukaryotic mRNAs, the point of translational initiation occurs at the AUG codon closest to the 5' cap of the transcript. Comparison of plant mRNA sequences and site directed mutagenesis experiments have demonstrated the existence of a consensus sequence surrounding the initiation codon in plants (Joshi, 1987; Lutcke *et al.*, 1987). However, consensus sequences will be apparent amongst individual plant species. For example, a compilation of sequences surrounding the initiation codon from 85 maize genes yields a consensus of 5'-(C/G)AUGGCG-3' (Luehrsen *et al.*, 1994). In tobacco protoplasts, transgenes encoding β-glucuronidase (GUS) and bacterial chitinase showed a 4-fold and an

eight-fold increase in expression, respectively, when the native sequences of these genes were changed to encode 5'-ACCAUGG-3' (Gallie *et al.*, 1987b; Jones *et al.*, 1988).

When producing chimeric transgenes (*i.e.* transgenes comprising DNA segments from different sources operably linked together), often the 5'-UTL of plant viruses is used.

5 The alfalfa mosaic virus (AMV) coat protein and brome mosaic virus (BMV) coat protein 5'-UTLs have been shown to enhance mRNA translation 8-fold in electroporated tobacco protoplasts (Gallie *et al.*, 1987a; 1987b). A 67-nucleotide derivative ( $\Omega$ ) of the 5'-UTL of tobacco mosaic virus RNA (TMV) fused to the chloramphenicol acetyltransferase (CAT) gene and GUS gene has been shown to enhance translation of reporter genes *in vitro* (Gallie 10 *et al.*, 1987a; 1987b; Sleat *et al.*, 1987; Sleat *et al.*, 1988). Electroporation of tobacco mesophyll protoplasts with transcripts containing the TMV leader fused to reporter genes CAT, GUS, and LUC produced a 33-, 21-, and 36-fold level of enhancement, respectively (Gallie *et al.*, 1987a; 1987b; Gallie *et al.*, 1991). Also in tobacco, an 83-nt 5'-UTL of potato virus X RNA was shown to enhance expression of the neomycin phosphotransferase II 15 (*NptII*) 4-fold (Poogin and Skryabin, 1992).

The effect of a 5'-UTL may be different depending on the plant, particularly between dicots and monocots. The TMV 5'-UTL has been shown to be more effective in tobacco protoplasts (Gallie *et al.*, 1989) than in maize protoplasts (Gallie and Young, 1994). Also, the 5'-UTLs from TMV- $\Omega$  (Gallie *et al.*, 1988), AMV-coat (Gehrke *et al.*, 1983; Jobling and 20 Gehrke, 1987), TMV-coat (Goelet *et al.*, 1982), and BMV-coat (French *et al.*, 1986) worked poorly in maize and inhibited expression of a luciferase gene in maize relative to its native leader (Koziel *et al.*, 1996). However, the 5'-UTLs from the cauliflower mosaic virus (CaMV) 35S transcript and the maize genes glutelin (Boronat *et al.*, 1986), PEP-carboxylase (Hudspeth and Grula, 1989) and ribulose biphosphate carboxylase showed a considerable 25 increase in expression of the luciferase gene in maize relative to its native leader (Koziel *et al.*, 1996).

These 5'-UTLs had different effects in tobacco. In contrast to maize, the TMV  $\Omega$  5'-UTL and the AMV coat protein 5'-UTL enhanced expression in tobacco, whereas the glutelin, maize PEP-carboxylase and maize ribulose-1,5-bisphosphate carboxylase 5'-UTLs 30 did not show enhancement relative to the native luciferase 5'-UTL (Koziel *et al.*, 1996).

Only the CaMV 35S 5'-UTL region enhanced luciferase expression in both maize and tobacco (Koziel *et al.*, 1996). Furthermore, the TMV and BMV coat protein 5'-UTLs were inhibitory in both maize and tobacco protoplasts (Koziel *et al.*, 1996).

#### 5 4.6.2 USE OF INTRONS TO INCREASE EXPRESSION

Including one or more introns in the transcribed portion of a gene has been found to increase heterologous gene expression in a variety of plant systems (Callis *et al.*, 1987; Maas *et al.*, 1991; Mascerenhas *et al.*, 1990; McElroy *et al.*, 1990; Vasil *et al.*, 1989), although not all introns produce a stimulatory effect and the degree of stimulation varies. The enhancing 10 effect of introns appears to be more apparent in monocots than in dicots. Tanaka *et al.* (1990) has shown that use of the catalase intron 1 isolated from castor beans increases gene expression in rice. Likewise, the first intron of the alcohol dehydrogenase 1 (*Adh1*) has been shown to increase expression of a genomic clone of *Adh1* comprising the endogenous 15 promoter in transformed maize cells (Callis *et al.*, 1987; Dennis *et al.*, 1984). Other introns that are also able to increase expression of transgenes which contain them include introns 2 and 6 of *Adh1* (Luehrsen and Walbot, 1991), the catalase intron (Tanaka *et al.*, 1990), intron 20 1 of the maize bronze 1 gene (Callis *et al.*, 1987), the maize sucrose synthase intron 1 (Vasil *et al.*, 1989), intron 3 of the rice actin gene (Luehrsen and Walbot, 1991), rice actin intron 1 (McElroy *et al.*, 1990), and the heat shock protein HSP70 (U. S. Patent No. 5,859,347, specifically incorporated herein by reference in its entirety). Similar results may also be obtained using sequences from certain exons, for example, the maize ubiquitin exon 1 (Christensen *et al.*, 1992).

Generally, to achieve optimal expression, the selected intron(s) should be present in the selected 5' transcriptional unit in the correct orientation with respect to the splice junction 25 sequences (Callis *et al.*, 1987; Maas *et al.*, 1991; Mascerenhas *et al.*, 1990; Oard *et al.*, 1989; Tanaka *et al.*, 1990; Vasil *et al.*, 1989). Intron 9 of *Adh1* has been shown to increase expression of a heterologous gene when placed 3' (or downstream of) the gene of interest (Callis *et al.*, 1987).

#### 4.6.3 USE OF SYNTHETIC GENES TO INCREASE GENE EXPRESSION

When introducing a prokaryotic gene into a eukaryotic host, or when expressing a eukaryotic gene in a non-native host, the sequence of the gene must often be altered or modified to allow efficient translation of the transcript(s) derived from the gene. Significant experience in using synthetic genes to increase expression of a desired protein has been achieved in the expression of *B. thuringiensis*-derived genes in plants. Native *B. thuringiensis* genes are expressed only at low levels in dicots and not at all in monocots (Koziel *et al.*, 1996). Codon usage in the native genes is considerably different from that found in typical plant genes, which have a higher G+C content. Strategies to increase expression of these genes in plants generally alter the overall G+C content of the genes. For example, synthetic *B. thuringiensis* δ-endotoxin encoding genes have resulted in significant improvements in expression of the δ-endotoxins in various crops including cotton (Perlak *et al.*, 1990; Wilson *et al.*, 1992), tomato (Perlak *et al.*, 1991), potato (Perlak *et al.*, 1993), rice (Cheng *et al.*, 1998), and maize (Koziel *et al.*, 1993).

In a similar fashion the inventors contemplate that the PEAMT/ΔPEAMT-encoding sequences of the present invention, may in certain circumstances, be altered to increase or decrease the expression of the genes in particular eukaryotic host cells and/or transgenic plants that comprise such constructs. Using molecular biology techniques that are well known to those of skill in the art, one may alter the coding or non-coding sequences of the particular PEAMT/ΔPEAMT gene(s) to optimize or facilitate its expression in transformed plant cells at suitable levels.

#### 4.6.4 CHLOROPLAST SEQUESTERING AND TARGETING

Methods of targeting proteins to the chloroplast have been developed utilizing the pea chloroplast transit peptide, to target the enzymes of the polyhydroxybutyrate synthesis pathway to the chloroplast (Nawrath *et al.*, 1994). Also, this technique negated the necessity of modification of the coding region other than to add an appropriate targeting sequence.

U. S. Patent No. 5,576,198 (specifically incorporated herein by reference in its entirety) discloses compositions and methods useful for genetic engineering of plant cells to provide a method of controlling the timing or tissue pattern of expression of foreign DNA

sequences inserted into the plant plastid genome. Constructs include those for nuclear transformation that provide for expression of a viral single subunit RNA polymerase in plant tissues, and targeting of the expressed polymerase protein into plant cell plastids. Also included are plastid expression constructs comprising a viral gene promoter region which is 5 specific to the RNA polymerase expressed from the nuclear expression constructs described above and a heterologous gene of interest to be expressed in the transformed plastid cells.

#### **4.6.5 EFFECTS OF 3' REGIONS ON TRANSGENE EXPRESSION**

The 3'-end regions of transgenes have been found to have a large effect on transgene 10 expression in plants (Ingelbrecht *et al.*, 1989). In this study, different 3' ends were operably linked to the neomycin phosphotransferase II (*NptII*) reporter gene and expressed in transgenic tobacco. The different 3' ends used were obtained from the octopine synthase gene, the 2S seed protein from *Arabidopsis*, the small subunit of *rbcS* from *Arabidopsis*, extension form carrot, and chalcone synthase from *Antirrhinum*. In stable tobacco 15 transformants, there was about a 60-fold difference between the best-expressing construct (small subunit *rbcS* 3' end) and the lowest expressing construct (chalcone synthase 3' end).

**TABLE 1**  
**PLANT PROMOTERS**

<b>Promoter</b>	<b>Reference<sup>a</sup></b>
<b>Viral</b>	
Figwort Mosaic Virus (FMV)	U. S. Patent No. 5,378,619
Cauliflower Mosaic Virus (CaMV)	U. S. Patent No. 5,530,196
	U. S. Patent No. 5,097,025
	U. S. Patent No. 5,110,732
<b>Plant</b>	
Elongation Factor	U. S. Patent No. 5,177,011
Tomato Polygalacturonase	U. S. Patent No. 5,442,052
<i>Arabidopsis</i> Histone H4	U. S. Patent No. 5,491,288
Phaseolin	U. S. Patent No. 5,504,200
Group 2	U. S. Patent No. 5,608,144
Ubiquitin	U. S. Patent No. 5,614,399
P119	U. S. Patent No. 5,633,440
$\alpha$ -amylase	U. S. Patent No. 5,712,112
Wheat starch branching enzyme	U. S. Patent No. 5,866,793
Osmotin	U. S. Patent No. 5,874,626
<b>Viral enhancer/Plant promoter</b>	
CaMV 35S enhancer/mannopine synthase promoter	U. S. Patent No. 5,106,739

<sup>a</sup>Each reference is specifically incorporated herein by reference in its entirety.

**TABLE 2**  
**TISSUE SPECIFIC PLANT PROMOTERS**

<b>Tissue Specific Promoter</b>	<b>Tissue(s)</b>	<b>Reference<sup>a</sup></b>
Blec	Epidermis	U. S. Patent No. 5,646,333
Malate synthase	Seeds; seedlings	U. S. Patent No. 5,689,040
Isocitrate lyase	Seeds; seedlings	U. S. Patent No. 5,689,040
Patatin	Tuber	U. S. Patent No. 5,436,393
ZRP2	Root	U. S. Patent No. 5,633,363
ZRP2(2.0)	Root	U. S. Patent No. 5,633,363
ZRP2(1.0)	Root	U. S. Patent No. 5,633,363
RB7	Root	U. S. Patent No. 5,459,252
	Root	U. S. Patent No. 5,401,836
	Fruit	U. S. Patent No. 4,943,674
	Meristem	U. S. Patent No. 5,589,583
	Guard cell	U. S. Patent No. 5,538,879
	Stamen	U. S. Patent No. 5,589,610
SodA1	Pollen; middle layer; stomium of anthers	Van Camp <i>et al.</i> , 1996
SodA2	Vascular bundles; stomata; axillary buds; pericycle; stomium; pollen	Van Camp <i>et al.</i> , 1996
CHS15	Flowers; root tips	Faktor <i>et al.</i> , 1996
Psam-1	Phloem tissue; cortex; root tips	Vander <i>et al.</i> , 1996
ACT11	Elongating tissues and organs; pollen; ovules	Huang <i>et al.</i> , 1997
ZmGBS	Pollen; endosperm	Russell and Fromm, 1997
zmZ27	Endosperm	Russell and Fromm, 1997
OsAGP	Endosperm	Russell and Fromm, 1997
osGT1	Endosperm	Russell and Fromm, 1997

**TABLE 2**  
**TISSUE SPECIFIC PLANT PROMOTERS**

<b>Tissue Specific Promoter</b>	<b>Tissue(s)</b>	<b>Reference<sup>a</sup></b>
RolC	Phloem tissue; bundle sheath; vascular parenchyma	Graham <i>et al.</i> , 1997
Sh	Phloem tissue	Graham <i>et al.</i> , 1997
CMD	Endosperm	Grosset <i>et al.</i> , 1997
Bnm1	Pollen	Treacy <i>et al.</i> , 1997
rice tungro bacilliform virus	Phloem	Yin <i>et al.</i> , 1997a; 1997b
S2-RNase	Pollen	Ficker <i>et al.</i> , 1998
LeB4	Seeds	Baumlein <i>et al.</i> , 1991
gf-2.8	Seeds; seedlings	Berna and Bernier, 1997

<sup>a</sup>Each reference is specifically incorporated herein by reference in its entirety.

The ability to express genes in a tissue specific manner in plants has led to the production of male and female sterile plants. Generally, the production of male sterile plants 5 involves the use of anther-specific promoters operably linked to heterologous genes that disrupt pollen formation (U. S. Patent No. 5,689,051; U. S. Patent No. 5,689,049; and U. S. Patent No. 5,659,124, each of which is specifically incorporated herein by reference in its entirety). U. S. Patent No. 5,633,441 (specifically incorporated herein by reference in its entirety) discloses a method of producing plants with female genetic sterility. The method 10 comprises the use of style-cell, stigma-cell, or style- and stigma-cell specific promoters that express polypeptides that, when produced in the cells of the plant kill or significantly disturbs the metabolism, functioning or development of the cells.

**TABLE 3**  
**INDUCIBLE PLANT PROMOTERS**

Promoter	Reference <sup>a</sup>
Heat shock promoter	U. S. Patent No. 5,447,858
Em	U. S. Patent No. 5,139,954
Adh1	Kyozuka <i>et al.</i> , 1991
HMG2	U. S. Patent No. 5,689,056
Cinnamyl alcohol dehydrogenase	U. S. Patent No. 5,633,439
Asparagine synthase	U. S. Patent No. 5,595,896
GST-II-27	U. S. Patent No. 5,589,614

<sup>a</sup>Each reference is specifically incorporated herein by reference in its entirety.

5    4.7        **GENE EXPRESSION IN PLANTS**

Although great progress has been made in recent years with respect to preparation of transgenic plants that express PEAMT/ΔPEAMT-polypeptides and DNA binding proteins such as the *lac* operator, the results of expressing heterologous genes in particular plant species are often disappointing. Unlike microbial genetics, little was known by early plant 10 geneticists about the factors that affected heterologous expression of foreign genes in plants. In recent years, however, several potential factors have been implicated as responsible in varying degrees for the level of protein expression from a particular coding sequence. For example, scientists now know that maintaining a significant level of a particular mRNA in the cell is indeed a critical factor. Unfortunately, the causes for low steady state levels of 15 mRNA encoding foreign proteins are many. First, full length RNA synthesis may not occur at a high frequency. This could, for example, be caused by the premature termination of RNA during transcription or due to unexpected mRNA processing during transcription. Second, full length RNA may be produced in the plant cell, but then processed (splicing, polyA addition) in the nucleus in a fashion that creates a nonfunctional mRNA. If the RNA 20 is not properly synthesized, terminated and polyadenylated, it cannot move to the cytoplasm for translation. Similarly, in the cytoplasm, if mRNAs have reduced half-lives (which are determined by their primary or secondary sequence) insufficient protein product will be

produced. In addition, there is an effect, whose magnitude is uncertain, of translational efficiency on mRNA half-life. In addition, every RNA molecule folds into a particular structure, or perhaps family of structures, which is determined by its sequence. The particular structure of any RNA might lead to greater or lesser stability in the cytoplasm.

5 Structure *per se* is probably also a determinant of mRNA processing in the nucleus. Unfortunately, it is impossible to predict, and nearly impossible to determine, the structure of any RNA (except for tRNA) *in vitro* or *in vivo*. However, it is likely that dramatically changing the sequence of an RNA will have a large effect on its folded structure. It is likely that structure *per se* or particular structural features also have a role in determining RNA

10 stability.

To overcome these limitations in heterologous gene expression, researchers have identified particular sequences and signals in RNAs that have the potential for having a specific effect on RNA stability. In certain embodiments of the invention, therefore, there is a desire to optimize expression of the disclosed nucleic acid segments *in planta*. One

15 particular method of doing so is by alteration of the native gene to remove sequences or motifs that decrease expression in a particular plant species. The process of engineering a coding sequence for optimal expression *in planta* is often referred to as “optimizing” a DNA sequence.

The present invention provides a method for preparing PEAMT- and ΔPEAMT-  
20 encoding genes that express their polypeptide product at sufficiently high levels in a heterologous transformed plant, so as to alter lipid content or alter lipid biosynthesis in the transformed plant.

#### 4.8 SYNTHETIC OLIGONUCLEOTIDES FOR MUTAGENESIS

25 When oligonucleotides are used in the mutagenesis, it is desirable to maintain the proper amino acid sequence and reading frame, without introducing common restriction sites such as *Bgl*II, *Hind*III, *Sac*I, *Kpn*I, *Eco*RI, *Nco*I, *Pst*I and *Sal*I into the modified gene. These restriction sites are found in poly-linker insertion sites of many cloning vectors. Of course, the introduction of new polyadenylation signals, ATTAA sequences or consecutive stretches  
30 of more than five A+T or G+C, should also be avoided. The preferred size for the

oligonucleotides is about 40 to about 50 bases, but fragments ranging from about 18 to about 100 bases have been utilized. In most cases, a minimum of about 5 to about 8 base pairs of homology to the template DNA on both ends of the synthesized fragment are maintained to insure proper hybridization of the primer to the template. The oligonucleotides should avoid sequences longer than five base pairs A+T or G+C. Codons used in the replacement of wild-type codons should preferably avoid the TA or CG doublet wherever possible. Codons are selected from a plant preferred codon table (such as Table 4 below) so as to avoid codons which are rarely found in plant genomes, and efforts should be made to select codons to preferably adjust the G+C content to about 50%.

10

**TABLE 4**  
**PREFERRED CODON USAGE IN PLANTS**

<b>Amino Acid</b>	<b>Codon</b>	<b>% Usage in Plants</b>
ARG	CGA	7
	CGC	11
	CGG	5
	CGU	25
	AGA	29
	AGG	23
LEU	CUA	8
	CUC	20
	CUG	10
	CUU	28
	UUA	5
	UUG	30
SER	UCA	14
	UCC	26
	UCG	3

**TABLE 4**  
**PREFERRED CODON USAGE IN PLANTS**

<b>Amino Acid</b>	<b>Codon</b>	<b>% Usage in Plants</b>
SER	UCU	21
	AGC	21
	AGU	15
THR	ACA	21
	ACC	41
	ACG	7
	ACU	31
PRO	CCA	45
	CCC	19
	CCG	9
	CCU	26
ALA	GCA	23
	GCC	32
	GCG	3
	GCU	41
GLY	GGA	32
	GGC	20
	GGG	11
	GGU	37
ILE	AUA	12
	AUC	45
	AUU	43

**TABLE 4**  
**PREFERRED CODON USAGE IN PLANTS**

<b>Amino Acid</b>	<b>Codon</b>	<b>% Usage in Plants</b>
VAL	GUA	9
	GUC	20
	GUG	28
	GUU	43
LYS	AAA	36
	AAG	64
ASN	AAC	72
	AAU	28
GLN	CAA	64
	CAG	36
HIS	CAC	65
	CAU	35
GLU	GAA	48
	GAG	52
ASP	GAC	48
	GAU	52
TYR	UAC	68
	UAU	32
CYS	UGC	78

**TABLE 4**  
**PREFERRED CODON USAGE IN PLANTS**

<b>Amino Acid</b>	<b>Codon</b>	<b>% Usage in Plants</b>
	UGU	22
PHE	UUC	56
	UUU	44
MET	AUG	100
TRP	UGG	100

Regions with many consecutive A+T bases or G+C bases are predicted to have a higher likelihood to form hairpin structures due to self-complementarity. Disruption of these regions by the insertion of heterogeneous base pairs is preferred and should reduce the 5 likelihood of the formation of self-complementary secondary structures such as hairpins which are known in some organisms to inhibit transcription (transcriptional terminators) and translation (attenuators).

Alternatively, a completely synthetic gene for a given amino acid sequence can be prepared, with regions of five or more consecutive A+T or G+C nucleotides being avoided. 10 Codons are selected avoiding the TA and CG doublets in codons whenever possible. Codon usage can be normalized against a plant preferred codon usage table (such as Table 4) and the G+C content preferably adjusted to about 50%. The resulting sequence should be examined to ensure that there are minimal putative plant polyadenylation signals and ATTAA sequences. Restriction sites found in commonly used cloning vectors are also preferably 15 avoided. However, placement of several unique restriction sites throughout the gene is useful for analysis of gene expression or construction of gene variants.

#### 4.9        OPTIMIZED GENE CONSTRUCTS

The expression of a plant gene that exists in double-stranded DNA form involves transcription of messenger RNA (mRNA) from one strand of the DNA by RNA polymerase enzyme, and the subsequent processing of the mRNA primary transcript inside the nucleus.

5        This processing may involve a 3' non-translated region that adds polyadenylated nucleotides to the 3' end of the RNA, or may also involve removal of introns and 5' capping. Transcription of DNA into mRNA is regulated by a region of DNA usually referred to as the "promoter." The promoter region contains a sequence of bases that signals RNA polymerase to associate with the DNA and to initiate the transcription of mRNA using one of the DNA  
10      strands as a template to make a corresponding strand of RNA.

A number of promoters that are active in plant cells have been described in the literature. These include the nopaline synthase (NOS) and octopine synthase (OCS) promoters (which are carried on tumor-inducing plasmids of *A. tumefaciens*), the Cauliflower Mosaic Virus (CaMV) 19S and 35S promoters, the light-inducible promoter from the small  
15      subunit of ribulose bis-phosphate carboxylase (ssRUBISCO, a very abundant plant polypeptide) and the mannopine synthase (MAS) promoter (Velten *et al.*, 1984 and Velten and Schell, 1985). All of these promoters have been used to create various types of DNA constructs that have been expressed in plants (see *e.g.*, Int. Pat. Appl. Publ. No. WO 84/02913).

20        Promoters that are known or are found to cause transcription of RNA in plant cells can be used in the present invention. Such promoters may be obtained from plants or plant viruses and include, but are not limited to, the CaMV35S promoter and promoters isolated from plant genes such as ssRUBISCO genes. As described below, it is preferred that the particular promoter selected should be capable of causing sufficient expression to result  
25      in the production of an effective amount of protein.

The promoters used in the DNA constructs (*i.e.* chimeric plant genes) of the present invention may be modified, if desired, to affect their control characteristics. For example, the CaMV35S promoter may be ligated to the portion of the ssRUBISCO gene that represses the expression of ssRUBISCO in the absence of light, to create a promoter which is  
30      active in leaves but not in roots. The resulting chimeric promoter may be used as described

herein. For purposes of this description, the phrase "CaMV35S" promoter thus includes variations of CaMV35S promoter, *e.g.*, promoters derived by means of ligation with operator regions, random or controlled mutagenesis, *etc.* Furthermore, the promoters may be altered to contain multiple "enhancer sequences" to assist in elevating gene expression.

5 The RNA produced by a DNA construct of the present invention also contains a 5' non-translated leader sequence. This sequence can be derived from the promoter selected to express the gene, and can be specifically modified so as to increase translation of the mRNA. The 5' non-translated regions can also be obtained from RNAs, suitable eukaryotic genes, or from a synthetic gene sequence. The present invention is not limited to constructs,  
10 as presented in the following examples. Rather, the non-translated leader sequence can be part of the 5' end of the non-translated region of the coding sequence for the virus coat protein, or part of the promoter sequence, or can be derived from an unrelated promoter or coding sequence. In any case, it is preferred that the sequence flanking the initiation site conform to the translational consensus sequence rules for enhanced translation initiation  
15 reported by Kozak (1984).

The DNA constructs of the present invention may also contain one or more modified or fully synthetic structural coding sequences which have been changed to enhance the performance of the gene in a particular species of plant. The structural genes of the present invention may optionally encode a fusion protein comprising an amino-terminal  
20 chloroplast transit peptide or secretory signal sequence.

The DNA construct also contains a 3' non-translated region. The 3' non-translated regions contain a polyadenylation signal which functions in plants to cause the addition of polyadenylate nucleotides to the 3' end of the viral RNA. Examples of suitable 3' regions are  
25 (1) the 3' transcribed, non-translated regions containing the polyadenylation signal of *Agrobacterium* tumor-inducing (Ti) plasmid genes, such as the nopaline synthase (NOS) gene, and (2) plant genes like the soybean storage protein (7S) genes and the small subunit of the RuBP carboxylase (E9) gene.

#### 4.10 METHODS FOR ALTERING PEAMT ACTIVITY IN TRANSGENIC PLANTS

By transforming a suitable host cell, such as a plant cell, with a recombinant PEAMT/ΔPEAMT-encoding polynucleotide segment, the expression of the PEAMT/ΔPEAMT-encoding polynucleotide under the control of an inducible promoter can 5 result in the formation of transgenic plants in which the alteration of PEAMT/ΔPEAMT activity and subsequent modulation of lipid content may be achieved.

By way of example, one may utilize an expression vector containing a coding region for a PEAMT or ΔPEAMT polynucleotide and an appropriate selectable marker to transform a suspension of embryonic plant cells, such as wheat or corn cells using a method such as 10 particle bombardment (Maddock *et al.*, 1991; Vasil *et al.*, 1992) to deliver the DNA coated on microprojectiles into the recipient cells. Transgenic plants are then regenerated from transformed embryonic calli that express the encoded polypeptide.

The formation of transgenic plants may also be accomplished using other methods of cell transformation that are known in the art such as *Agrobacterium*-mediated DNA transfer 15 (Fraley *et al.*, 1983). Alternatively, DNA can be introduced into plants by direct DNA transfer into pollen (U. S. Patent No. 5,629,183; Zhou *et al.*, 1983; Hess, 1987; Luo *et al.*, 1988), by injection of the DNA into reproductive organs of a plant (Pena *et al.*, 1987), or by direct injection of DNA into the cells of immature embryos followed by the rehydration of desiccated embryos (Neuhaus *et al.*, 1987; Benbrook *et al.*, 1986).

20 Methods for the regeneration, development, and cultivation of plants from single plant protoplast transformants or from various transformed explants are well known in the art (Weissbach and Weissbach, 1988). This regeneration and growth process typically includes the steps of selection of transformed cells, culturing those individualized cells through the usual stages of embryonic development through the rooted plantlet stage. Transgenic 25 embryos and seeds are similarly regenerated. The resulting transgenic rooted shoots are thereafter planted in an appropriate plant growth medium such as soil.

The development or regeneration of plants containing the foreign, exogenous gene that encodes a polypeptide of interest introduced by *Agrobacterium* from leaf explants can be achieved by methods well known in the art such as described (Horsch *et al.*, 1985). In this 30 procedure, transformants are cultured in the presence of a selection agent and in a medium

that induces the regeneration of shoots in the plant strain being transformed as described (Fraley *et al.*, 1983).

This procedure typically produces shoots within two to four months and those shoots are then transferred to an appropriate root-inducing medium containing the selective agent 5 and an antibiotic to prevent bacterial growth. Shoots that rooted in the presence of the selective agent to form plantlets are then transplanted to soil or other media to allow the production of roots. These procedures vary depending upon the particular plant strain employed, such variations being well known in the art.

Preferably, the regenerated plants are self-pollinated to provide homozygous 10 transgenic plants, as discussed before. Otherwise, pollen obtained from the regenerated plants is crossed to seed-grown plants of agronomically important, preferably inbred lines. Conversely, pollen from plants of those important lines is used to pollinate regenerated plants. A transgenic plant of the present invention containing a desired polypeptide is cultivated using methods well known to one skilled in the art.

Such plants can form germ cells and transmit the transformed trait(s) to progeny 15 plants. Likewise, transgenic plants can be grown in the normal manner and crossed with plants that have the same transformed hereditary factors or other hereditary factors. The resulting hybrid individuals have the corresponding phenotypic properties. A transgenic plant of this invention thus has an increased amount of a coding region that encodes the 20 PEAMT or  $\Delta$ PEAMT polypeptide of interest. A preferred transgenic plant is an independent segregant and can transmit that gene and its activity to its progeny. A more preferred transgenic plant is homozygous for that gene, and transmits that gene to each of its offspring on sexual mating.

Seed from a transgenic plant may be grown in the field or greenhouse, and resulting 25 sexually mature transgenic plants are self-pollinated to generate true breeding plants. The progeny from these plants become true breeding lines that are evaluated for, by way of example, increased lipid biosynthesis, preferably in the field, under a range of environmental conditions. The inventors contemplate that the present invention will find particular utility in the creation of transgenic plants of commercial interest including various grains, grasses,

fibers, tubers, legumes, ornamental plants, cacti, succulents, fruits, berries, and vegetables, as well as a number of nut- and fruit-bearing trees and plants.

#### **4.11 PLANT CELLS AND TRANSGENIC PLANTS EXPRESSING PEAMT**

5 In one embodiment, the invention provides a transgenic plant having incorporated into its genome a transgene that encodes a PEAMT or  $\Delta$ PEAMT polypeptide. A further aspect of the invention is a transgenic plant having incorporated into its genome a transgene that encodes such a polypeptide. Other embodiments of the invention also concern the progeny of such a transgenic plant, as well as its seed, the progeny from such seeds, and 10 seeds arising from the second and subsequent generation plants derived from such a transgenic plant.

15 The invention also discloses and claims host cells, both native, and genetically engineered, which express one or more genes encoding all or substantially all of a PEAMT polypeptide to produce the encoded polypeptide(s) in a suitably transformed host cell, and in particular, in a transformed plant cell.

In yet another aspect, the present invention provides methods for producing a transgenic plant that expresses such a nucleic acid segment. The process of producing transgenic plants is well known in the art. In general, the method comprises transforming a suitable host cell with one or more DNA segments that contain a promoter operatively linked 20 to a coding region that encodes one or more PEAMT polypeptides. Such a coding region is generally operatively linked to a transcription-terminating region, whereby the promoter is capable of driving the transcription of the coding region in the cell, and hence providing the cell the ability to produce the recombinant protein *in vivo*. Alternatively, in instances where it is desirable to control, regulate, or decrease the amount of a particular recombinant protein 25 expressed in a particular transgenic cell, the invention also provides for the expression of an antisense oligonucleotide or other nucleic acid sequences that are complementary to the mRNA that encodes the expressed polypeptide. The use of antisense mRNA as a means of controlling or decreasing the amount of a given protein of interest in a cell is well known in the art.

As used herein, the term "transgenic plant" is intended to refer to a plant that has incorporated DNA sequences, including but not limited to genes which are perhaps not normally present, DNA sequences not normally transcribed into RNA or translated into a protein ("expressed"), or any other genes or DNA sequences which one desires to introduce 5 into the non-transformed plant, such as genes which may normally be present in the non-transformed plant but which one desires to either genetically engineer or to have altered expression.

It is contemplated that in some instances the genome of a transgenic plant of the present invention will have been augmented through the stable introduction of one or more 10 transgenes, either native, synthetically modified, or mutated. In some instances, more than one transgene will be incorporated into the genome of the transformed host plant cell. Such is the case when more than one DNA segment is incorporated into the genome of such a plant. In certain situations, it may be desirable to have one, two, three, four, or even more 15 PEAMT proteins (either native or recombinantly-engineered) incorporated and stably expressed in the transformed transgenic plant.

A preferred gene that may be introduced includes, for example, a DNA sequence from a plant that encodes a PEAMT or a  $\Delta$ PEAMT polypeptide, and particularly one or more of those described in SEQ ID NO:2 or SEQ ID NO:4.

Means for transforming a plant cell and the preparation of a transgenic cell line are 20 well known in the art, and are discussed herein. Vectors, plasmids, cosmids, bacterial artificial chromosomes (BACs), plant artificial chromosomes (PACs), yeast artificial chromosomes (YACs), and DNA segments for use in transforming such cells will, of course, generally comprise either the operons, genes, or gene-derived sequences of the present invention, either native, or synthetically-derived, and particularly those encoding the 25 disclosed PEAMT polypeptides. These nucleic acid constructs can further include structures such as promoters, enhancers, polylinkers, or even gene sequences that have positively- or negatively-regulating activity upon the particular genes of interest as desired. The nucleic acid segment or gene may encode either a native or modified protein, which will be expressed in the resultant recombinant cells, and/or which will impart an improved 30 phenotype to the regenerated plant

Such transgenic plants may be desirable for modulating lipid biosynthesis in a population of monocotyledonous or dicotyledonous plants. Particularly preferred plants include grains such as corn, wheat, rye, rice, barley, and oats; legumes such as beans, soybeans; tubers such as potatoes; fiber crops such as flax and cotton; turf and pasture 5 grasses; ornamental plants; shrubs; trees; vegetables; berries; citrus crops, including oranges, tangerines, grapefruit, limes, lemons, and the like; fruits, cacti, succulents, and other commercially-important crops including greenhouse, garden and houseplants.

In a related aspect, the present invention also encompasses a seed produced by the transformed plant, a progeny from such seed, and a seed produced by the progeny of the 10 original transgenic plant, produced in accordance with the above process. Such progeny and seeds will have one or more PEAMT-encoding transgene(s) stably incorporated into its genome, and such progeny plants will inherit the traits afforded by the introduction of a stable transgene in Mendelian fashion. All such transgenic plants having incorporated into 15 their genome transgenic DNA segments encoding one or more PEAMT polypeptides are aspects of this invention.

#### **4.12 ISOLATING HOMOLOGOUS GENE AND GENE FRAGMENTS ENCODING PEAMT**

The polynucleotide sequences of the subject invention include not only full-length sequences but also fragments of these sequences, (including *e.g.*, fusion proteins), which 20 retain the PEAMT or  $\Delta$ PEAMT enzymatic activity of the sequences specifically exemplified herein in SEQ ID NO:2 and SEQ ID NO:4, respectively.

It should be apparent to a person skilled in this art that the various genetic constructs encoding PEAMT/ $\Delta$ PEAMT polypeptides can be identified and obtained through several means. The PEAMT/ $\Delta$ PEAMT-encoding genes, or portions thereof, may be obtained from a 25 culture depository, or constructed synthetically, for example, by use of a gene machine. Variations of these genes may be readily constructed using standard techniques for making point mutations. Also, fragments of these genes can be made using commercially available exonucleases or endonucleases according to standard procedures. For example, enzymes 30 such as *Bal*31 or site-directed mutagenesis can be used to systematically cut off nucleotides from the ends of these genes. Also, genes or gene fragments that encode biologically active

polypeptides may be obtained using a variety of other restriction enzymes. Proteases may be used to directly obtain active fragments of these constructs.

Equivalent polypeptides and/or polynucleotides encoding these equivalent polypeptides can also be isolated from DNA libraries using the teachings provided herein.

5 For example, antibodies to the polypeptides disclosed and claimed herein can be used to identify and isolate other similar or related polypeptides from a mixture of proteins. These antibodies can then be used to specifically identify equivalent polypeptides possessing the desired characteristics by a variety of methodologies including, *e.g.*, immunoprecipitation, enzyme linked immunoassay (ELISA), and/or Western blotting.

10 A further method for identifying the polypeptides and polynucleotides of the subject invention is through the use of oligonucleotide probes. These probes are nucleotide sequences having a detectable label. As is well known in the art, if the probe molecule and nucleic acid sample hybridize by forming a strong bond between the two molecules, it can be reasonably assumed that the probe and sample are essentially identical. The probe's detectable label provides a means for determining in a known manner whether hybridization 15 has occurred. Such a probe analysis provides a rapid method for identifying genes of the subject invention.

20 The nucleotide segments that are used as probes according to the invention may be synthesized by use of nucleic acid synthesizers using standard procedures. In the use of the nucleotide segments as probes, the particular probe is labeled with any suitable label known to those skilled in the art, including radioactive and non-radioactive labels. Typical radioactive labels include  $^{32}\text{P}$ ,  $^{125}\text{I}$ ,  $^{131}\text{I}$ ,  $^{35}\text{S}$ ,  $^{14}\text{C}$ ,  $^3\text{H}$ , or the like. A probe labeled with a radioactive isotope can be constructed from a nucleotide sequence complementary to the 25 DNA sample by a conventional nick translation reaction, using a DNase and DNA polymerase. The probe and sample can then be combined in a hybridization buffer solution and held at an appropriate temperature until annealing occurs. Thereafter, the membrane is washed free of extraneous materials, leaving the sample and bound probe molecules typically detected and quantified by autoradiography and/or liquid scintillation counting.

30 Non-radioactive labels include, for example, ligands such as biotin or thyroxine, as well as enzymes such as hydrolases or peroxidases, or the various chemiluminescers such as

luciferin, or fluorescent compounds like fluorescein and its derivatives. The probe may also be labeled at both ends with different types of labels for ease of separation, as, for example, by using an isotopic label at the end mentioned above and a biotin label at the other end.

Duplex formation and stability depend on substantial complementarity between the 5 two strands of a hybrid, and, as noted above, a certain degree of mismatch can be tolerated. Therefore, the probes of the subject invention include mutations (both single and multiple), deletions, insertions of the described sequences, and combinations thereof, wherein said mutations, insertions and deletions permit formation of stable hybrids with the target 10 polynucleotide of interest. Mutations, insertions, and deletions can be produced in a given polynucleotide sequence in many ways, by methods currently known to an ordinarily skilled artisan, and perhaps by other methods which may become known in the future.

The potential variations in the probes listed are due, in part, to the redundancy of the genetic code. Because of the redundancy of the genetic code, *i.e.* more than one coding nucleotide triplet (codon) can be used for most of the amino acids used to make proteins. 15 Therefore different nucleotide sequences can code for a particular amino acid. Thus, the amino acid sequences of the disclosed polypeptides can be prepared by equivalent nucleotide sequences encoding the same amino acid sequence of the protein or peptide. Accordingly, the subject invention includes such equivalent nucleotide sequences. Also, inverse or complement sequences are an aspect of the subject invention and can be readily used by a 20 person skilled in this art. In addition it has been shown that proteins of identified structure and function may be constructed by changing the amino acid sequence if such changes do not alter the protein secondary structure (Kaiser and Kezdy, 1984). Thus, the subject invention includes mutants of the amino acid sequence depicted herein that do not alter the protein secondary structure, or if the structure is altered, the biological activity is substantially 25 retained. Further, the invention also includes mutants of organisms hosting all or part of one or more of the DNA constructs of the invention. Such mutants can be made by techniques well known to persons skilled in the art. For example, UV irradiation can be used to prepare mutants of host organisms. Likewise, such mutants may include asporogenous host cells that also can be prepared by procedures well known in the art.

#### 4.13 PEPTIDE NUCLEIC ACID COMPOSITIONS

In certain embodiments, the inventors contemplate the use of peptide nucleic acids (PNAs) in the practice of the methods of the invention. PNAs are DNA analogs that mimic the structure of the polynucleotide, in which the nucleobases are attached to a pseudopeptide backbone (Good and Nielsen, 1997). PNAs can be utilized in a number of methods that traditionally have used RNAs or DNAs (U. S. Patent No. 5,786,461; U. S. Patent No. 5,773,571, U. S. Patent No. 5,766,855; U. S. Patent No. 5,736,336; U. S. Patent No. 5,719,262; and U. S. Patent No. 5,539,082, each of which is specifically incorporated herein by reference in its entirety). Often PNA sequences perform better in techniques than the corresponding RNA or DNA sequences and have utilities that are not inherent to RNA or DNA. Methods of making, and using PNAs are also found in Corey (1997).

PNAs when delivered within cells have the potential to be general sequence-specific regulators of gene expression. Reviews of PNAs and their use as antisense and anti-gene agents exist (Nielsen *et al.*, 1993; Hanvey *et al.*, 1992; and Good and Nielsen, 1997). Other applications of PNAs include use in DNA strand invasion (Nielsen *et al.*, 1991), antisense inhibition (Hanvey *et al.*, 1992), mutational analysis (Orum *et al.*, 1993), enhancers of transcription (Mollegaard *et al.*, 1994), nucleic acid purification (Orum *et al.*, 1995), isolation of transcriptionally active genes (Boffa *et al.*, 1995), blocking of transcription factor binding (Vickers *et al.*, 1995), genome cleavage (Veselkov *et al.*, 1996), biosensors (Wang *et al.*, 1996), *in situ* hybridization (Thisted *et al.*, 1996), and in an alternative to Southern blotting (Perry-O'Keefe, 1996).

#### 4.14 PEAMT-SPECIFIC ANTIBODY COMPOSITIONS AND FORMULATIONS THEREOF

Means for preparing and characterizing antibodies are well known in the art (see, *e.g.*, Harlow and Lane, 1988; incorporated herein by reference). The methods for generating monoclonal antibodies (mAbs) generally begin along the same lines as those for preparing polyclonal antibodies. Briefly, a polyclonal antibody is prepared by immunizing an animal with an immunogenic composition in accordance with the present invention and collecting antisera from that immunized animal. A wide range of animal species can be used for the production of antisera. Typically the animal used for production of anti-antisera is a rabbit, a

mouse, a rat, a hamster, a guinea pig or a goat. Because of the relatively large blood volume of rabbits, a rabbit is a preferred choice for production of polyclonal antibodies.

As is well known in the art, a given composition may vary in its immunogenicity. It is often necessary therefore to boost the host immune system, as may be achieved by 5 coupling a peptide or polypeptide immunogen to a carrier. Exemplary and preferred carriers are keyhole limpet hemocyanin (KLH) and bovine serum albumin (BSA). Other albumins such as ovalbumin, mouse serum albumin or rabbit serum albumin can also be used as carriers. Means for conjugating a polypeptide to a carrier protein are well known in the art and include glutaraldehyde, *m*-maleimidobencoyl-N-hydroxysuccinimide ester, carbodiimide 10 and bis-biazotized benzidine.

As is also well known in the art, the immunogenicity of a particular immunogen composition can be enhanced by the use of non-specific stimulators of the immune response, known as adjuvants. Exemplary and preferred adjuvants include complete Freund's adjuvant (a non-specific stimulator of the immune response containing killed *Mycobacterium tuberculosis*), incomplete Freund's adjuvants and aluminum hydroxide adjuvant. 15

The amount of immunogen composition used in the production of polyclonal antibodies varies upon the nature of the immunogen as well as the animal used for immunization. A variety of routes can be used to administer the immunogen (subcutaneous, intramuscular, intradermal, intravenous and intraperitoneal). The production of polyclonal 20 antibodies may be monitored by sampling blood of the immunized animal at various points following immunization. A second, booster, injection may also be given. The process of boosting and titering is repeated until a suitable titer is achieved. When a desired level of immunogenicity is obtained, the immunized animal can be bled and the serum isolated and stored, and/or the animal can be used to generate mAbs.

25 mAbs that are specific for a PEAMT or  $\Delta$ PEAMT peptide epitope, peptide, or polypeptide may be readily prepared through use of well-known techniques, such as those exemplified in U. S. Patent 4,196,265, incorporated herein by reference. Typically, this technique involves immunizing a suitable animal with a selected immunogen composition, *e.g.*, a purified or partially purified protein, polypeptide or peptide. The immunizing 30 composition is administered in a manner effective to stimulate antibody-producing cells.

Rodents such as mice and rats are preferred animals, however, the use of rabbit, sheep frog cells is also possible. The use of rats may provide certain advantages (Goding, 1986), but mice are preferred, with the BALB/c mouse being most preferred as this is most routinely used and generally gives a higher percentage of stable fusions.

5 Following immunization, somatic cells with the potential for producing antibodies, specifically B-lymphocytes (B-cells), are selected for use in the mAb generating protocol. These cells may be obtained from biopsied spleens, tonsils or lymph nodes, or from a peripheral blood sample. Spleen cells and peripheral blood cells are preferred, the former because they are a rich source of antibody producing cells that are in the dividing plasmablast stage, and the latter because peripheral blood is easily accessible. Often, a panel of animals 10 will have been immunized and the spleen of animal with the highest antibody titer removed and the spleen lymphocytes obtained by homogenizing the spleen with a syringe. Typically, a spleen from an immunized mouse contains approximately about  $5 \times 10^7$  to about  $2 \times 10^8$  lymphocytes.

15 The antibody-producing B-lymphocytes from the immunized animal are then fused with cells of an immortal myeloma cell, generally one of the same species as the animal that was immunized. Myeloma cell lines suited for use in hybridoma-producing fusion procedures preferably are non-antibody-producing, have high fusion efficiency, and enzyme deficiencies that render them incapable of growing in certain selective media which support 20 the growth of only the desired fused cells (hybridomas).

Any one of a number of myeloma cells may be used, as are known to those of skill in the art (Goding, 1986; Campbell, 1984). For example, where the immunized animal is a mouse, one may use P3-X63/Ag8, X63-Ag8.653, NS1/1.Ag 4 1, Sp210-Ag14, FO, NSO/U, MPC-11, MPC11-X45-GTG 1.7 and S194/5XX0 Bul; for rats, one may use R210.RCY3, 25 Y3-Ag 1.2.3, IR983F and 4B210; and U-266, GM1500-GRG2, LICR-LON-HMy2 and UC729-6 are all useful in connection with human cell fusions.

One preferred murine myeloma cell is the NS-1 myeloma cell line (also termed P3-NS-1-Ag4-1), which is readily available from the NIGMS Human Genetic Mutant Cell Repository by requesting cell line repository number GM3573. Another mouse myeloma

cell line that may be used is the 8-azaguanine-resistant mouse murine myeloma SP2/0 non-producer cell line.

Methods for generating hybrids of antibody-producing spleen or lymph node cells and myeloma cells usually comprise mixing somatic cells with myeloma cells in a 2:1 ratio, 5 though the ratio may vary from about 20:1 to about 1:1, respectively, in the presence of an agent or agents (chemical or electrical) that promote the fusion of cell membranes. Fusion methods using Sendai virus have been described (Kohler and Milstein, 1975; 1976), and those using polyethylene glycol (PEG), such as 37% (vol./vol.) PEG, by Gefter *et al.* (1977). The use of electrically induced fusion methods is also appropriate (Goding, 1986).

10 Fusion procedures usually produce viable hybrids at low frequencies, about  $1 \times 10^{-6}$  to about  $1 \times 10^{-8}$ . However, this does not pose a problem, as the viable, fused hybrids are differentiated from the parental, unfused cells (particularly the unfused myeloma cells that would normally continue to divide indefinitely) by culturing in a selective medium. The selective medium is generally one that contains an agent that blocks the *de novo* synthesis of 15 nucleotides in the tissue culture media. Exemplary and preferred agents are aminopterin, methotrexate, and azaserine. Aminopterin and methotrexate block *de novo* synthesis of both purines and pyrimidines, whereas azaserine blocks only purine synthesis. Where aminopterin or methotrexate is used, the media is supplemented with hypoxanthine and thymidine as a source of nucleotides (HAT medium). Where azaserine is used, the media is 20 supplemented with hypoxanthine.

The preferred selection medium is HAT. Only cells capable of operating nucleotide salvage pathways are able to survive in HAT medium. The myeloma cells are defective in key enzymes of the salvage pathway, *e.g.*, hypoxanthine phosphoribosyl transferase (HPRT), and they cannot survive. The B-cells can operate this pathway, but they have a limited life 25 span in culture and generally die within about two wk. Therefore, the only cells that can survive in the selective media are those hybrids formed from myeloma and B-cells.

This culturing provides a population of hybridomas from which specific hybridomas are selected. Typically, selection of hybridomas is performed by culturing the cells by single-clone dilution in microtiter plates, followed by testing the individual clonal 30 supernatants (after about two to three wk) for the desired reactivity. The assay should be

sensitive, simple and rapid, such as radioimmunoassays, enzyme immunoassays, cytotoxicity assays, plaque assays, dot immunobinding assays, and the like.

The selected hybridomas would then be serially diluted and cloned into individual antibody-producing cell lines, which clones can then be propagated indefinitely to provide 5 mAbs. The cell lines may be exploited for mAb production in two basic ways. A sample of the hybridoma can be injected (often into the peritoneal cavity) into a histocompatible animal of the type that was used to provide the somatic and myeloma cells for the original fusion. The injected animal develops tumors secreting the specific monoclonal antibody produced by the fused cell hybrid. The body fluids of the animal, such as serum or ascites fluid, can then 10 be tapped to provide mAbs in high concentration. The individual cell lines may also be cultured *in vitro*, where the mAbs are naturally secreted into the culture medium from which they can be readily obtained in high concentrations. mAbs produced by either means may be further purified, if desired, using filtration, centrifugation and various chromatographic methods such as HPLC or affinity chromatography.

15

#### 4.15 EPITOPE CORE SEQUENCES

The present invention is also directed to PEAMT/ΔPEAMT polypeptide compositions, free from total cells and other polypeptides, which comprise a purified PEAMT/ΔPEAMT polypeptide which incorporates an epitope that is immunologically cross-reactive with one or more of the PEAMT/ΔPEAMT-specific antibodies of the present 20 invention.

As used herein, the term "incorporating an epitope(s) that is immunologically cross-reactive with one or more anti-PEAMT/ΔPEAMT antibodies" is intended to refer to a peptide or protein antigen which includes a primary, secondary or tertiary structure similar to an 25 epitope located within a PEAMT/ΔPEAMT polypeptide. The level of similarity will generally be to such a degree that monoclonal or polyclonal antibodies directed against the PEAMT/ΔPEAMT polypeptide will also bind to, react with, or otherwise recognize, the cross-reactive peptide or protein antigen. Various immunoassay methods may be employed in conjunction with such antibodies, such as, for example, Western blotting, ELISA, RIA, 30 and the like, all of which are known to those of skill in the art.

The identification of PEAMT/ΔPEAMT epitopes and/or their functional equivalents, suitable for use in vaccines is a relatively straightforward matter. For example, one may employ the methods of Hopp, as taught in U. S. Patent 4,554,101, incorporated herein by reference, which teaches the identification and preparation of epitopes from amino acid sequences on the basis of hydrophilicity. The methods described in several other papers, and software programs based thereon, can also be used to identify epitopic core sequences (see, for example, Jameson and Wolf, 1988; Wolf *et al.*, 1988; U. S. Patent 4,554,101). The amino acid sequence of these "epitopic core sequences" may then be readily incorporated into peptides, either through the application of peptide synthesis or recombinant technology.

Preferred peptides for use in accordance with the present invention will generally be on the order of about 5 to about 25 amino acids in length, and more preferably about 8 to about 20 amino acids in length. It is proposed that shorter antigenic peptide sequences will provide advantages in certain circumstances, for example, in the preparation of vaccines or in immunological detection assays. Exemplary advantages include the ease of preparation and purification, the relatively low cost and improved reproducibility of production, and advantageous biodistribution.

It is proposed that particular advantages of the present invention may be realized through the preparation of synthetic peptides that include modified and/or extended epitopic/immunogenic core sequences that result in a "universal" epitopic peptide directed to PEAMT/ΔPEAMT-related sequences. It is proposed that these regions represent those that are most likely to promote T-cell or B-cell stimulation in an animal, and, hence, elicit specific antibody production in such an animal.

An epitopic core sequence, as used herein, is a relatively short stretch of amino acids that is "complementary" to, and therefore will bind, antigen-binding sites on PEAMT/ΔPEAMT epitope-specific antibodies. Additionally or alternatively, an epitopic core sequence is one that will elicit antibodies that are cross-reactive with antibodies directed against the peptide compositions of the present invention. It will be understood that in the context of the present disclosure, the term "complementary" refers to amino acids or peptides that exhibit an attractive force towards each other. Thus, certain epitope core sequences of the present invention may be operationally defined in terms of their ability to compete with

or perhaps displace the binding of the desired protein antigen with the corresponding protein-directed antisera.

In general, the size of the polypeptide or peptide antigen is not believed to be particularly crucial, so long as it is at least large enough to carry the identified core sequence or sequences. The smallest useful core sequence expected by the present disclosure would generally be on the order of about 5 amino acids in length, with sequences on the order of 8 or 25 being more preferred. Thus, this size will generally correspond to the smallest peptide antigens prepared in accordance with the invention. However, the size of the antigen may be larger where desired, so long as it contains a basic epitopic core sequence.

10 The identification of epitopic core sequences is known to those of skill in the art, for example, as described in U. S. Patent 4,554,101, incorporated herein by reference, which teaches the identification and preparation of epitopes from amino acid sequences on the basis of hydrophilicity. Moreover, numerous computer programs are available for use in predicting antigenic portions of proteins (see *e.g.*, Jameson and Wolf, 1988; Wolf *et al.*, 15 1988). Computerized peptide sequence analysis programs (*e.g.*, DNASTar™ software, DNASTar, Inc., Madison, WI) may also be useful in designing synthetic epitopes and epitope analogs in accordance with the present disclosure.

In certain embodiments, particular advantages may be realized through the preparation of synthetic PEAMT/ΔPEAMT peptides that include epitopic/immunogenic core 20 sequences. These epitopic core sequences may be identified as hydrophilic and/or mobile regions of the polypeptides or those that include a T cell motif. It is known in the art that such regions represent those that are most likely to promote B cell or T cell stimulation, and, hence, elicit specific antibody production.

To confirm that a polypeptide is immunologically cross-reactive with, or a biological 25 functional equivalent of, one or more epitopes of the disclosed PEAMT/ΔPEAMT peptides is also a straightforward matter. This can be readily determined using specific assays, *e.g.*, of a single proposed epitopic sequence, or using more general screens, *e.g.*, of a pool of randomly generated synthetic peptides or protein fragments. The screening assays may be employed to identify either equivalent antigens or cross-reactive antibodies. In any event, the principle is 30 the same, *i.e.* based upon competition for binding sites between antibodies and antigens.

Suitable competition assays that may be employed include protocols based upon immunohistochemical assays, ELISAs, RIAs, Western or dot blotting and the like. In any of the competitive assays, one of the binding components, generally the known element, such as a PEAMT or PEAMT-derived peptide, or a known antibody, will be labeled with a detectable 5 label and the test components, that generally remain unlabeled, will be tested for their ability to reduce the amount of label that is bound to the corresponding reactive antibody or antigen.

As an exemplary embodiment, to conduct a competition study between PEAMT/ΔPEAMT and any test antigen, one would first label PEAMT/ΔPEAMT with a detectable label, such as, *e.g.*, biotin or an enzymatic, radioactive or fluorogenic label, to 10 enable subsequent identification. One would then incubate the labeled antigen with the other, test, antigen to be examined at various ratios (*e.g.*, 1:1, 1:10 and 1:100) and, after mixing, one would then add the mixture to a known antibody. Preferably, the known antibody would be immobilized, *e.g.*, by attaching to an ELISA plate. The ability of the mixture to bind to the antibody would be determined by detecting the presence of the 15 specifically bound label. This value would then be compared to a control value in which no potentially competing (test) antigen was included in the incubation.

The assay may be any one of a range of immunological assays based upon hybridization, and the reactive antigens would be detected by means of detecting their label, *e.g.*, using streptavidin in the case of biotinylated antigens or by using a chromogenic 20 substrate in connection with an enzymatic label or by simply detecting a radioactive or fluorescent label.

The reactivity of the labeled antigen, *e.g.*, a PEAMT/ΔPEAMT-derived peptide, in the absence of any test antigen would be the control high value. The control low value would be obtained by incubating the labeled antigen with an excess of unlabeled antigen, when 25 competition would occur and reduce binding. A significant reduction in labeled antigen reactivity in the presence of a test antigen is indicative of a test antigen that is "cross-reactive," *i.e.* that has binding affinity for the same antibody. "A significant reduction," in terms of the present application, may be defined as a reproducible (*i.e.* consistently observed) reduction in binding.

In addition to the peptidyl compounds described herein, the inventors also contemplate that other sterically similar compounds may be formulated to mimic the key portions of the peptide structure. Such compounds, which may be termed peptidomimetics, may be used in the same manner as the peptides of the invention and hence are also 5 functional equivalents. The generation of a structural functional equivalent may be achieved by the techniques of modeling and chemical design known to those of skill in the art. It will be understood that all such sterically similar constructs fall within the scope of the present invention.

Syntheses of epitopic sequences, or peptides which include an antigenic epitope 10 within their sequence, are readily achieved using conventional synthetic techniques such as the solid phase method (*e.g.*, through the use of a commercially-available peptide synthesizer such as an Applied Biosystems Model 430A Peptide Synthesizer). Peptide antigens synthesized in this manner may then be aliquoted in predetermined amounts and stored in conventional manners, such as in aqueous solutions or, even more preferably, in a powder or 15 lyophilized state pending use.

In general, due to the relative stability of peptides, they may be readily stored in aqueous solutions for fairly long periods of time if desired, *e.g.*, up to six months or more, in virtually any aqueous solution without appreciable degradation or loss of antigenic activity. However, where extended aqueous storage is contemplated it will generally be desirable to 20 include agents including buffers such as Tris or phosphate buffers to maintain a pH of about 7.0 to about 7.5. Moreover, it may be desirable to include agents that will inhibit microbial growth, such as sodium azide or Merthiolate. For extended storage in an aqueous state it will be desirable to store the solutions at 4°C, or more preferably, frozen. Of course, where the peptides are stored in a lyophilized or powdered state, they may be stored virtually 25 indefinitely, *e.g.*, in metered aliquots that may be rehydrated with a predetermined amount of water (preferably distilled) or buffer prior to use.

#### **4.16 ANTISENSE OLIGONUCLEOTIDES TARGETED TO mRNA**

In certain embodiments, the inventors contemplate the use of antisense compositions 30 to negatively regulate the expression of a gene encoding PEAMT or ΔPEAMT in a host cell.

The end result of the flow of genetic information is the synthesis of protein. DNA is transcribed by polymerases into messenger RNA and translated on the ribosome to yield a folded, functional protein. Thus, even from this simplistic description of an extremely complex set of reactions, it is obvious that there are several steps along the route where 5 protein synthesis can be inhibited. The native DNA segment encoding PEAMT or  $\Delta$ PEAMT has two strands: a sense strand and an antisense strand held together by hydrogen bonding. The messenger RNA encoding PEAMT or  $\Delta$ PEAMT has the same nucleotide sequence as the sense DNA strand except that the DNA thymidine is replaced by uridine. Thus, antisense nucleotide sequences will bind to the mRNA encoding the PEAMT or  $\Delta$ PEAMT 10 polypeptides and inhibit production of the corresponding protein.

The targeting of antisense oligonucleotides to bind mRNA is one mechanism to shut down protein synthesis. For example, the synthesis of polygalacturonase and the muscarine type-2 acetylcholine receptor are inhibited by antisense oligonucleotides directed to their respective mRNA sequences (U. S. Patent 5,739,119 and U. S. Patent 5,759,829, U. S. Patent 15 5,801,154; U. S. Patent 5,789,573; U. S. Patent 5,718,709 and U. S. Patent 5,610,288, each specifically incorporated herein by reference in its entirety).

In illustrative embodiments, antisense oligonucleotides may be prepared which are complementary nucleic acid sequences that can recognize and bind to target genes or the transcribed mRNA, resulting in the arrest and/or inhibition of deoxyribonucleic acid (DNA) 20 transcription or translation of the messenger ribonucleic acid (mRNA). These oligonucleotides can be expressed within a host cell that normally expresses PEAMT-specific mRNA to reduce or inhibit the expression of this mRNA. Thus, the oligonucleotides may be useful for reducing the level of PEAMT polypeptide in a suitably transformed host cell or transgenic plant.

25 The native nucleic acid segment encoding PEAMT has, as do all such plant DNAs, two strands: a sense strand and an antisense strand held together in a duplex formation by hydrogen bonding. The messenger RNA (mRNA) encoding PEAMT has the same nucleotide sequence as the sense DNA strand except that the thymidine in DNA is replaced by uridine in DNA. Thus, preferred antisense oligonucleotide compositions for use in the 30 practice of the present invention are those sequences that specifically bind to the mRNA

coding for PEAMT and that inhibit or reduce the expression of the PEAMT polypeptide encoding by that mRNA.

The present invention provides a composition comprising at least a first oligonucleotide of at least about 9 to about 45 or so bases in length, wherein the 5 oligonucleotide specifically binds to a portion of mRNA expressed from a gene encoding a plant PEAMT polypeptide, and further wherein binding of the oligonucleotide to the mRNA is effective in decreasing the activity of or reducing the quantity of the PEAMT enzyme in a host plant cell expressing the mRNA.

In certain aspects of the invention, the oligonucleotide comprises deoxyribonucleic 10 acid, ribonucleic acid, or peptide-nucleic acid. In particular embodiments, the oligonucleotide comprises a sequence of at least nine, at least ten, at least eleven, at least twelve, at least thirteen, or at least fourteen, up to and including the full-length contiguous sequences from SEQ ID NO:1 or SEQ ID NO:3. When longer antisense molecules are required, one may employ an oligonucleotide that comprises a sequence of at least fifteen, at 15 least sixteen, at least seventeen, at least eighteen, at least nineteen, or at least twenty, up to and including the full-length contiguous sequences from SEQ ID NO:1 or SEQ ID NO:3. Such antisense molecules may comprise even longer contiguous nucleotide sequences, such as those comprising about 21, about 22, about 23, about 24, about 25, about 26, about 27, about 28, about 29, or about 30 or so contiguous nucleotides from SEQ ID NO:1 or SEQ ID 20 NO:3.

#### 4.17 DEFINITIONS

In accordance with the present invention, nucleic acid sequences include and are not limited to DNA (including and not limited to genomic or extragenomic DNA), genes, RNA 25 (including and not limited to mRNA and tRNA), nucleosides, and suitable nucleic acid segments either obtained from native sources, chemically synthesized, modified, or otherwise prepared by the hand of man. The following words and phrases have the meanings set forth below.

**A, an:** In accordance with long standing patent law convention, the words "a" and 30 "an" when used in this application, including the claims, denotes "one or more".

**Expression:** The combination of intracellular processes, including transcription and translation undergone by a coding DNA molecule such as a structural gene to produce a polypeptide.

5 **Promoter:** A recognition site on a DNA sequence or group of DNA sequences that provide an expression control element for a structural gene and to which RNA polymerase specifically binds and initiates RNA synthesis (transcription) of that gene.

**Regeneration:** The process of growing a plant from a plant cell (*e.g.*, plant protoplast or explant).

10 **Structural gene:** A gene that is expressed to produce a polypeptide.

**Transformation:** A process of introducing an exogenous DNA sequence (*e.g.*, a vector, a recombinant DNA molecule) into a cell or protoplast in which that exogenous DNA is incorporated into a chromosome or is capable of autonomous replication.

**Transformed cell:** A cell whose DNA has been altered by the introduction of an exogenous DNA molecule into that cell.

15 **Transgenic cell:** Any cell derived or regenerated from a transformed cell or derived from a transgenic cell. Exemplary transgenic cells include plant calli derived from a transformed plant cell and particular cells such as leaf, root, stem, *e.g.*, somatic cells, or reproductive (germ) cells obtained from a transgenic plant.

20 **Transgenic plant:** A plant or progeny thereof derived from a transformed plant cell or protoplast, wherein the plant DNA contains an introduced exogenous DNA molecule not originally present in a native, non-transgenic plant of the same strain. The terms "transgenic plant" and "transformed plant" have sometimes been used in the art as synonymous terms to define a plant whose DNA contains an exogenous DNA molecule. However, it is thought more scientifically correct to refer to a regenerated plant or callus obtained from a 25 transformed plant cell or protoplast as being a transgenic plant, and that usage will be followed herein.

**Vector:** A DNA molecule capable of replication in a host cell and/or to which another DNA segment can be operatively linked so as to bring about replication of the attached segment. A plasmid is an exemplary vector.

The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides: (a) "reference sequence," (b) "comparison window," (c) "sequence identity," (d) "percentage of sequence identity," and (e) "substantial identity."

(a) As used herein, "reference sequence" is a defined sequence used as a basis for 5 sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.

(b) As used herein, "comparison window" makes reference to a contiguous and specified segment of a, polynucleotide sequence, wherein the polynucleotide sequence in the 10 comparison window may comprise additions or deletions (*i.e.* gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 60, 70, 80, 90, or 100 contiguous nucleotides, or even longer depending upon the particular analysis. Those of skill in the art understand that 15 to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

Methods of alignment of sequences for comparison are well known in the art. Optimal alignment of sequences for comparison may be conducted by the local homology 20 algorithm of Smith *et al.*, (1981); by the homology alignment algorithm of Needleman *et al.*, (1970); by the search for similarity method of Pearson *et al.* (1988); by computerized implementations of these algorithms, including, but not limited to: CLUSTAL in the PC/Gene program by Intelligenetics (Mountain View, CA); GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer 25 Group (Madison, WI); the CLUSTAL program is well described by Higgins *et al.* (1988); Higgins *et al.* (1989); Corpet *et al.* (1988); Huang *et al.* (1992), and Person *et al.* (1994); preferred computer alignment methods also include the BLASTP, BLASTN, and BLASTX algorithms (Altschul *et al.*, 1990). Alignment is also often performed by inspection and manual alignment.

(c) As used herein, “sequence identity” or “identity” in the context of two nucleic acid or polypeptide sequences makes reference to the residues in the two sequences that are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that 5 residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (*e.g.*, charge or hydrophobicity) and therefore do not change the functional properties of the molecule. When sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature 10 of the substitution. Sequences that differ by such conservative substitutions are said to have “sequence similarity” or “similarity.” Means for making this adjustment are well known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non- 15 conservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, *e.g.*, as implemented in the program PC/GENE (Intelligenetics, Mountain View, CA).

(d) As used herein, “percentage of sequence identity” means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion 20 of the polynucleotide sequence in the comparison window may comprise additions or deletions (*i.e.* gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, 25 dividing the number of matched positions by the total number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

(e) The term “substantial identity” of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least about 70% sequence identity, preferably at least about 80%, more preferably at least about 90%, and most preferably at 30 least about 95% to about 99%, compared to a reference sequence using one of the alignment

programs described using standard parameters. One of skill in the art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like. Substantial identity of amino acid 5 sequences for these purposes normally means sequence identity of at least about 60%, more preferably at least about 70%, at least about 80%, at least about 90%, and most preferably at least about 95% to about 99%.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. Generally, stringent conditions 10 are selected to be about 5°C to about 20°C lower than the thermal melting point ( $T_m$ ) for the specific sequence at a defined ionic strength and pH. The  $T_m$  is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typically, stringent wash conditions are those in which the salt concentration is about 0.02 M at pH 7 and the temperature is at least about 50°C, about 55°C, 15 or even at least about 60°C, about 65°C, or at least about 70°C or 75°C so. However, nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides they encode are substantially identical. This may occur, *e.g.*, when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two nucleic acid sequences are substantially identical is 20 when the polypeptide encoded by the first nucleic acid is immunologically cross reactive with the polypeptide encoded by the second nucleic acid.

(e)(ii) The term "substantial identity" in the context of a peptide indicates that a peptide comprises a sequence with at least about 70% sequence identity to a reference sequence, preferably at least about 80%, more preferably at least about 85%, most preferably 25 at least about 90% or at least about 95% to about 99% or so sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman *et al.* (1970). An indication that two peptide sequences are substantially identical is that one peptide is immunologically reactive with antibodies raised against the second peptide. Thus, a peptide 30 is substantially identical to a second peptide, for example, where the two peptides differ only

by a conservative substitution. Peptides that are "substantially similar" share sequences as noted above except that residue positions that are not identical may differ by conservative amino acid changes.

The proteins of the invention may be altered in various ways including amino acid 5 substitutions, deletions, truncations, and insertion. Methods for such manipulations are generally known in the art. For example, amino acid sequence variants of the proteins can be prepared by mutations in the DNA that encode them. Methods for mutagenesis and nucleotide sequence alterations are well known in the art (see *e.g.*, Kunkel, 1985; Kunkel *et al.*, 1987; U. S. Patent No. 4,873,192; Walker and Gaastra, 1983, each of which is 10 specifically incorporated herein by reference in its entirety).

Thus, the genes and nucleotide sequences of the invention include both the naturally occurring sequences as well as mutant forms. Likewise, the proteins of the invention encompass both naturally occurring proteins as well as variations and modified forms thereof. Such variants will continue to possess the desired PEAMT or  $\Delta$ PEAMT activity. 15 Obviously, the mutations that will be made in the DNA encoding the variant must not place the sequence out of reading frame and preferably will not create complementary regions that could produce secondary mRNA structure (see *e.g.*, Eur. Pat. Appl. Publ. No. 75,444, specifically incorporated herein by reference in its entirety).

"Seed-specific" promoters of the invention may also include embryo-specific 20 promoters. Such promoters may include, but are not limited to, globulin 1, cruciferin, napin,  $\beta$ -conglycinin, phaseolin, and the like, as well as other promoters associated with storage proteins or involved in fatty acid or lipid biosynthesis.

The polynucleotides of the invention may be provided in one or more expression 25 cassettes or genetic constructs to facilitate introduction and stable integration into the plant genome. Such expression cassettes may comprise one or more transcriptional initiation regions linked to a coding sequence or antisense sequence of the particular PEAMT or  $\Delta$ PEAMT sequence. Such an expression cassette is generally provided with a plurality of restriction sites for insertion of the sequence to be under the transcriptional regulation of the regulatory regions. The expression cassette may additionally contain one or more selectable 30 marker genes.

The transcriptional initiation region, the promoter, may be native (*i.e.* analogous) or foreign (*i.e.* heterologous) to the plant host. Additionally, the promoter may be a synthetic sequence. By “foreign,” it is intended that the transcriptional initiation region not be found in the native plant into which the transcriptional initiation region is introduced. For example, 5 a spinach PEAMT gene would be consider a “foreign” gene if introduced into the genome of a non-spinach plant, such as corn or soybeans.

The transcriptional cassette may include in the 5' to 3' direction of transcription, a transcriptional and translational initiation region, a PEAMT or  $\Delta$ PEAMT coding sequence, and a transcriptional and translational termination region functional in the particular plant 10 species into which the construct is introduced. The termination region may be native with the transcriptional initiation region, may be native with the DNA sequence of interest, or may be derived from another source. Convenient termination regions are available from the Ti-plasmid of *A. tumefaciens* such as the octopine synthase and nopaline synthase termination 15 regions (Guerineau *et al.*, 1991; Proudfoot, 1991; Sanfacon *et al.*, 1991; Mogen *et al.*, 1990; Munroe *et al.*, 1990; Ballas *et al.*, 1989; Joshi *et al.*, 1987, each of which is specifically incorporated herein by reference in its entirety).

In preparing the expression cassette, the various polynucleotide fragment(s) may be manipulated, so as to provide for the DNA sequences in the proper orientation and, as appropriate, in the proper reading frame. Toward this end, adapters or linkers may be 20 employed to join the polynucleotides or other manipulations may be involved to provide for convenient restriction sites, removal of superfluous DNA, removal of restriction sites, or the like. For this purpose, *in vitro* mutagenesis, primer repair, restriction, annealing, resubstitutions (*e.g.*, transitions and transversions), may be involved.

The modulation of lipid biosynthesis can be achieved in any plant of interest. Of 25 particular interest are plants useful for human foodstuffs and domestic animal feedstock. Such plants include forage and seed crop plants, and preferably crops such as cereals and oilseed crops. Of particular interest are plants where the seed is produced in high amounts, or the seed or a seed part is edible. Seeds of interest include the oilseeds, such as from *Brassica*, cotton, soybean, safflower, canola, sunflower, coconut, palm, *etc.*; grain seeds such 30 as wheat, rice, corn, *etc.*; other seeds including oats, pumpkin, squash, poppy, sesame,

peanut, peas, beans and other legumes, cocoa, coffee, *etc.*; and tree nuts such as walnuts, pecans, almonds, *etc.* Especially preferred plants are corn, soybean, legumes, safflower, sunflower, canola, *Brassica*, wheat, rye, rice, millet, sorghum, alfalfa, and the like.

The modified plant may be grown into plants in accordance with conventional ways  
5 (McCorrick *et al.*, 1986). These plants may then be grown, and either pollinated with the same transformed strain or different strains, and the resulting hybrid having the desired phenotypic characteristic identified. Two or more generations may be grown to ensure that the subject phenotypic characteristic is stably maintained and inherited into the progeny and subsequent generations of the transformed plant. Likewise, the seeds from the transformed  
10 plant or from a progeny or subsequent generation of the plant may be harvested and assayed to ensure the desired phenotype has been achieved in the progeny and the seeds from the transgenic plant and its offspring.

## 5. EXAMPLES

15 The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventors to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure,  
20 appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

### 5.1 EXAMPLE 1 – CLONING AND CHARACTERIZATION OF PEAMT

#### 25 5.1.1 EXPERIMENTAL PROCEDURES

##### 5.1.1.1 CHEMICALS

30  $[^{32}\text{P}]$ dCTP (3000 mCi mmol<sup>-1</sup>) and [*methyl*-<sup>14</sup>C]AdoMet (59 mCi mmol<sup>-1</sup>) were purchased from NEN; the specific activity of AdoMet was adjusted to the desired value with unlabeled compound (Sigma, St. Louis, MO). Chiral HPLC (Beaudouin *et al.*, 1993) showed that [*methyl*-<sup>14</sup>C]AdoMet was ≥99% in the *S,S* (biologically active) form. Unlabeled AdoMet

was 85% in the *S,S* form and 15% in the *R,S* (inactive) form; specific activities were calculated using the *S,S*-AdoMet content. [<sup>33</sup>P]P-MME and [<sup>14</sup>C]P-Cho were made as described (McNeil *et al.*, 2000). Restriction and modification enzymes, and oligonucleotides were from Gibco-BRL, NEB or Boehringer Mannheim. P-Cho (Na<sub>2</sub> salt) was from TCI America (Portland, OR). P-EA and all other biochemicals were from Sigma. AG-50 (HR<sup>+</sup>) ion exchange resin was from BioRad. Silica gel G (0.25-mm) TLC plates were from Merck.

#### 5.1.1.2 PLANT MATERIALS

*Spinacia oleracea* L. plants (cv. Savoy Hybrid 612, Harris Moran Seeds) were grown in coarse vermiculite with an 8-hr day at 24°C (photosynthetic photon flux density 325  $\mu\text{mol m}^{-2}\text{s}^{-1}$ ) and 16-hr night at 19°C. Plants were irrigated daily with 0.5X Hoagland's solution. Prior to harvest, plants were held at nighttime conditions for 40 hr and then placed under day conditions for 8 hr; salinized plants were irrigated with 200 mM NaCl at the onset of the 8-hr day period (Smith *et al.*, 1999). Harvested leaves were frozen in liquid N<sub>2</sub> and stored at -80°C.

#### 5.1.1.3 YEAST STRAINS AND GROWTH CONDITIONS

The *S. pombe* strains used were 972h- (*h<sup>-</sup>S*) and Bx22 (*h<sup>-</sup>S cho2-20 ade6-M210 leu1-32*) (Kanipes, 1997). Both were maintained on YEA medium (Kanipes *et al.*, 1998) supplemented with 40  $\mu\text{M}$  *myo*-inositol. SD medium was as described (Kanipes *et al.*, 1998) except that it contained 110  $\mu\text{M}$  *myo*-inositol, 1 g l<sup>-1</sup> of CSM-leu (Bio101, Vista, CA), plus 250 mg l<sup>-1</sup> each of lysine, adenine, uracil, and histidine, 1 mM EA and, for plates only, 20 mg l<sup>-1</sup> phloxine B. All liquid media also contained 0.5 g l<sup>-1</sup> asparagine.

#### 5.1.1.4 cDNA LIBRARY CONSTRUCTION

A salinized spinach cDNA expression library was constructed in the *S. pombe* vector pREP3, which contains a thiamine-repressible expression cassette driven by the *nmt* promoter (Maundrell, 1993). To prepare pREP3 for library construction, it was digested with *MscI* and *SalI*, dephosphorylated with shrimp alkaline phosphatase, ligated to an adapter made by hybridizing the oligonucleotides 5'-CTCGAGATCTG-3' (SEQ ID NO:5) and 5'-

TCGACAGATCTCGAG-3' (SEQ ID NO:6), digested with *Xba*I, gel-purified and religated. This replaced the *Msc*I site with *Xba*I and *Bgl*II sites in the polylinker, creating pREP3A. The cDNA library was directionally cloned in the *Bgl*II/*Sal*I sites; to ensure these sites were accessible, a 2.2-kb *Bgl*II/*Sal*I fragment from pJD301 (Luehrsen *et al.*, 1992) was ligated to 5 pREP3A creating pREP3B. Total RNA was isolated from salinized spinach leaves as described (Rathinasabapathi *et al.*, 1997). Poly(A)<sup>+</sup> RNA was isolated on poly-U Sephadex (Hondred *et al.*, 1987) and used to construct cDNA with the Stratagene ZAP-cDNA synthesis kit, replacing the kit's *Eco*RI adapter with a *Bgl*II adapter made by hybridizing the oligonucleotides 5'-CTCGTGCCA-3' (SEQ ID NO:7) and 5'-GATCTGGCACGAG-3 (SEQ 10 ID NO:8). Size-selected cDNAs (average length 1.2 kb) were ligated to pREP3B *Bgl*II/*Sal*I under optimized conditions and transformed into XL1-Blue MRF' cells by electroporation. The expression library ( $3.2 \times 10^6$  colony forming units) was amplified as described (Alexander, 1987).

15 **5.1.1.5 COMPLEMENTATION OF A CHO2<sup>-</sup> MUTANT**

Plasmid DNA from the spinach expression library was prepared and transformed into Bx22 cells by standard procedures (Kanipes *et al.*, 1998). Transformants were selected on SSD medium and then replica-plated onto SD medium containing 1 mM EA to select for colonies displaying the complemented phenotype. Complementing plasmids were rescued 20 and transformed into *E. coli* DH10B cells for analysis. Sequencing was carried out using the ABI Prism dye terminator cycle sequencing Ready Reaction (PE Applied Biosystems, Foster City, CA) and an ABI model 373 sequencer.

**5.1.1.6 CONSTRUCTION OF  $\Delta$ PEAMT**

25 The plasmid encoding PEAMT, designated pREP3-PEAMT, was digested with *Bam*HI and end-polished with T4 DNA polymerase. The 3'-sequence of PEAMT was removed by digestion with *Msc*I; the remaining sequence plus vector was gel-purified and religated to give  $\Delta$ PEAMT. In  $\Delta$ PEAMT the native coding sequence terminates at Gly-286, and the pREP3 polylinker adds the sequence Ile Pro Gly to the carboxyl-terminus.

### 5.1.1.7 RNA GEL BLOT ANALYSIS

Total RNA was prepared from unstressed and salinized spinach leaves using the RNAeasy plant mini kit (Qiagen, Valencia, CA). Ten- $\mu$ g samples of RNA were separated in formaldehyde/1.5% agarose gels and transferred to supported nitrocellulose membrane (NitroPure, MSI, Westborough, MA), hybridized and washed according to the manufacturer's protocols. The PEAMT probe template was a 1.2-kb *Bam*HI/*Bgl*II fragment of pREP3-PEAMT. The rRNA probe template was a 0.9-kb *Sma*I fragment of a *Zamia pumila* rRNA clone (Nairn and Ferl, 1988). Probes were labeled with [<sup>32</sup>P]dCTP by the random primer method. Hybridization was detected by autoradiography.

10

### 5.1.1.8 ENZYME ISOLATION AND MOLECULAR MASS DETERMINATION

Cells were grown to an OD<sub>600</sub> of  $\approx$ 1 in 50 ml of medium at 30°C, shaking at 250 rpm. The media were SD containing 250 mg l<sup>-1</sup> leucine for Bx22 cells, and SSD for wild type cells and Bx22 cells harboring pREP3-PEAMT or pREP3- $\Delta$ PEAMT. Subsequent operations were at 0-4°C. Cells were harvested by centrifugation (5000  $\times$  g, 10 min), washed twice with water and once with 100 mM HEPES-KOH, pH 7.8, resuspended in 0.5 ml of lysis buffer (100 mM HEPES-KOH, pH 7.8, 2 mM Na<sub>2</sub>EDTA, 5 mM dithiothreitol, 10% glycerol) and transferred to 2-ml microfuge tubes containing  $\approx$ 1.5 g of acid-washed glass beads (425-600  $\mu$ m). Cells were broken by vortexing at maximum speed for 2 min. The brei was centrifuged (16,000  $\times$  g, 2 min), the supernatant was removed and the beads were washed with 3  $\times$  1 ml of lysis buffer. The combined supernatants were clarified by centrifugation (16,000  $\times$  g, 15 min) and desalted on a PD-10 column (Amersham Pharmacia Biotech, Piscataway, NJ) equilibrated with storage buffer (10 mM HEPES-KOH, pH 7.8, 2 mM Na<sub>2</sub>EDTA, 5 mM dithiothreitol, 10% glycerol). The extract was aliquoted, frozen in liquid N<sub>2</sub> and stored at -80°C. Spinach leaf extracts were prepared as described (Summers and Weretilnyk, 1993). Protein was determined using the Bio-Rad dye reagent (Hercules, CA). Native molecular mass was estimated using a Waters 626 HPLC system (Milford, MA) equipped with a Superdex 200 HR 10/30 column (Amersham Pharmacia Biotech) as described (Weretilnyk *et al.*, 1995).

30

### 5.1.1.9 ENZYME ASSAYS

PEAMT activity was measured by a modification of published methods (Datko and Mudd, 1988b; Summers and Weretilnyk, 1993), under conditions in which product formation was proportional to enzyme concentration and time. Unless otherwise indicated, assays (final volume 100  $\mu$ l) contained 10  $\mu$ l 10X assay buffer (1 M HEPES-KOH, 20 mM Na<sub>2</sub>EDTA, pH 8.6), 10% glycerol, 100 nCi [*methyl*-<sup>14</sup>C]AdoMet, P-EA and enzyme extract as specified in the text, and were incubated at 30°C for 15 min. The final pH in the assays was 8.1. Reactions were stopped by adding 1 ml of ice-cold water; each was applied to a 1-ml AG-50 (HR<sup>+</sup>) column held at 4°C and the assay tube was rinsed with 1 ml of water, which was also applied to the column. Products were eluted with 10 ml of 0.1 N HCl, and 2 ml of the eluate was mixed with 3 ml of Ready Gel scintillation fluid (Beckman, Palo Alto, CA) and counted. For assay blanks, enzyme was omitted during incubation and added just before the ion exchange step. Product recovery was determined to be 84% by spiking unlabeled reaction mixtures with [<sup>14</sup>C]P-Cho, and experimental values were corrected accordingly. Reaction products were separated by TLC on silica gel G plates developed in methanol:acetone:concentrated HCl (90:10:4, vol./vol./vol.) (TLC system 1), and detected by autoradiography.

AdoMet:EA *N*-methyltransferase activity was assayed as above except that reactions contained 200 nmol EA and 160 nmol (40 nCi) of [*methyl*-<sup>14</sup>C]AdoMet. The [*methyl*-<sup>14</sup>C]AdoMet was removed by treating with activated charcoal and centrifuging (Cook and Wagner, 1984), and <sup>14</sup>C-incorporation into freebase products was measured by counting a sample of the supernatant. TLC tests confirmed that the charcoal did not bind free bases. For assay blanks, EA was omitted. AdoMet:Ptd-EA *N*-methyltransferase activity was measured essentially as described (Datko and Mudd, 1988b). Assays (final volume 100  $\mu$ l) contained 9  $\mu$ l 10X assay buffer, 0.8 nmol (47 nCi) [*methyl*-<sup>14</sup>C]AdoMet, 10  $\mu$ l of a Ptd-EA emulsion in 1X assay buffer (10 mg ml<sup>-1</sup>, sonicated for 3  $\times$  30 sec), and enzyme extract; incubation was for 30 min at 30°C. The phospholipid fraction was isolated for <sup>14</sup>C quantification as described (Datko and Mudd, 1988b) except that the methanol:chloroform:water proportions were 12:5:1 (vol./vol./vol.). Data were corrected for the <sup>14</sup>C incorporation in control assays containing extract from Bx22 cells.

## 5.1.2 RESULTS

### 5.1.2.1 PEAMT CLONING BY COMPLEMENTATION OF A *S. POMBE CHO2* MUTATION

Cho moieties are synthesized *de novo* in *S. pombe* via three sequential 5 methylations of Ptd-EA. The *cho2* gene product mediates the first of these, Ptd-EA → Ptd-MME (FIG. 1B), and *cho2*<sup>-</sup> mutants require MME, DME or Cho for growth (Kanipes and Henry, 1997). *S. pombe* incorporates pre-formed Cho or other free bases into phospholipids *via* the CDP-base or Kennedy pathway, *i.e.* base → P-base → CDP-base → Ptd-base (FIG. 1B) (Kanipes and Henry, 1997). These features of Cho metabolism in *S. pombe* suggested that PEAMT could be cloned by complementation of a *cho2*<sup>-</sup> mutant, 10 because expression of PEAMT would restore Cho prototrophy by installing a bypass to the blocked Ptd-EA methylation step (FIG. 1B).

A cDNA expression library was therefore constructed in the *S. pombe* expression vector pREP3 using mRNA from salinized spinach leaves, salinization being known to 15 increase PEAMT activity (Weretilnyk *et al.*, 1995). The pREP3 plasmid contains the *leu2* gene for selection and a thiamine-repressible expression cassette (Maundrell, 1993). The amplified library was transformed into *S. pombe* strain Bx22, a *leu1*<sup>-</sup> *cho2*<sup>-</sup> mutant (Kanipes and Henry, 1997). Transformants were selected first for leucine prototrophy on medium 20 containing thiamine and Cho, then replica plated onto medium without thiamine and with EA in place of Cho. Screening ≈30,000 transformants in this way identified 24 complemented colonies that harbored plasmids with the same 2.2-kb insert, as judged by restriction analysis and sequencing. No complementation was obtained with the vector alone and retransformation of Bx22 with rescued plasmid conferred Cho prototrophy, establishing that 25 the complementation is due to the encoded plant protein. The complemented strains lacked detectable activity with Ptd-EA or EA as substrates (<1 pkat mg<sup>-1</sup> protein) but had high PEAMT activity (FIG. 2A). TLC analysis of the PEAMT reaction mixtures confirmed that P-MME was formed (FIG. 2A, inset). The specific activity of PEAMT in extracts of complemented cells was up to 80-fold greater than that in salinized spinach leaves (FIG. 2A).

Because sequence analysis showed that PEAMT has two methyltransferase 30 domains (see below), a convenient *MscI* site was used to remove almost all of the C-terminal

domain (FIG. 2B). The truncated construct ( $\Delta$ PEAMT) remained able to complement the *cho2* mutation and specified a protein with PEAMT activity (FIG. 2A).

#### 5.1.2.2 ANALYSIS OF THE PEAMT cDNA SEQUENCE

5 The PEAMT cDNA comprises a 494-residue open reading frame (FIG. 3) flanked by long 5'- and 3'-untranslated regions (253 and 496 bp, respectively). The deduced polypeptide (56.4-kDa) can be divided in the center (FIG. 3, arrowhead) into two sequences that share significant homology (21% identity, 53% similarity), each of which contains the consensus sequences (Kagan and Clarke, 1994; Gary *et al.*, 1996) for methyltransferase 10 motifs I, post-I, II and III (FIG. 2B and FIG. 3). PEAMT thus appears to contain two distinct but related methyltransferase domains. This is consistent with its size, which is roughly double that typical of small molecule methyltransferases (Fujioka, 1992). The deduced PEAMT sequence has no recognizable N-terminal signal sequence, consistent with biochemical evidence that the spinach enzyme is cytosolic (Weretilnyk *et al.*, 1995).

15 The novel, bipartite structure of PEAMT prompted a search for homologous sequences in GenBank™, to determine whether similar proteins occur in other plants. PEAMT homologs were found in species representing three other families: *Malvaceae* (e.g., cotton), *Brassicaceae* (e.g., *Arabidopsis* and canola) and *Gramineae* (e.g., barley and rice). The *Arabidopsis* protein was predicted from genomic DNA sequence and was complete 20 (FIG. 3); those for other species were fragments deduced from expressed sequence tags (ESTs) spanning the junction between the domains.

#### 5.1.2.3 PROPERTIES OF RECOMBINANT PEAMT

25 The native molecular mass was estimated by size exclusion chromatography to be 57-kDa, indicating that the enzyme exists as a monomer. PEAMT activity showed a broad pH optimum in the region 8.1-8.5; the  $K_m$  values for P-EA and AdoMet were 96 and 140  $\mu$ M, respectively. In view of the bipartite structure of PEAMT and the finding that highly purified (but not homogenous) PEAMT preparations from spinach catalyze the methylation 30 of P-MME and P-DME as well as of P-EA (Smith *et al.*, 1999), it was of interest to determine whether the recombinant enzyme mediates more than the first methylation of

P-EA. The products formed when PEAMT is incubated for long periods with a small amount of P-EA and  $\geq 3.5$ -fold excess of [*methyl-<sup>14</sup>C*]AdoMet were analyzed. In these conditions, <sup>14</sup>C appears successively in P-MME, P-DME and P-Cho, as shown qualitatively in FIG. 4A. As incubation proceeds, the molar quantities of P-MME and P-DME peak and 5 then decline while P-Cho continues to accumulate (FIG. 4B). These results establish that PEAMT catalyzes all three methylations required to convert P-EA to P-Cho, and also show that the intermediates P-MME and P-DME do not invariably remain bound to the enzyme because both accumulate transiently during the reaction.

10 **5.1.2.4 PRODUCTS OF THE REACTION CATALYZED BY THE N-TERMINAL  
PEAMT DOMAIN**

Deleting the C-terminal domain of PEAMT did not abolish complementation or enzyme activity (FIG. 2A). The N-terminal domain thus catalyzes the first methylation of EA – but not necessarily the others. The reaction products of the truncated ( $\Delta$ PEAMT) and 15 wild type enzymes were therefore compared. After a 3-hr incubation with a small quantity of P-EA and a 5-fold excess of [*methyl-<sup>14</sup>C*]AdoMet,  $\Delta$ PEAMT yields only [<sup>14</sup>C]P-MME whereas the wild type enzyme yields, as expected, almost solely [<sup>14</sup>C]P-Cho (FIG. 5A). This result demonstrates that the N-terminal methyltransferase domain mediates only the first 20 N-methylation of P-EA, and strongly implies that the C-terminal domain mediates the other two. In this connection it is noteworthy that the kinetics of product formation for the wild type enzyme (FIG. 4B) show that P-MME accumulates to a higher level than P-DME. This is consistent with the second and third methylation steps occurring at the same catalytic site, from which the intermediate P-DME does not exit very freely.

25 **5.1.2.5 SENSITIVITY TO P-CHO AND OTHER METABOLITES**

Recombinant wild type PEAMT was strongly inhibited by its product P-Cho (FIG. 5B), as reported for the activity extracted from spinach or *Lemna* plants (Smith *et al.*, 1999; Mudd and Datko, 1989a). Activity was reduced by 80% at 2 mM and by 95% at 10 mM P-Cho, 10 mM being approximately the P-Cho concentration in the cytosol of spinach 30 leaves (Bligny *et al.*, 1990; Winter *et al.*, 1994). This effect appeared specific inasmuch as

Cho, GlyBet, MME and DME were not inhibitory at 2 mM. The truncated  $\Delta$ PEAMT enzyme remained sensitive to P-Cho, although significantly ( $P = 0.05$ ) less so than the wild-type enzyme, retaining twice as much of its initial activity at the physiological P-Cho concentration of 10 mM (FIG. 5B).

5

#### 5.1.2.6 EFFECT OF SALINIZATION ON PEAMT mRNA LEVEL

RNA gel blot analyses revealed an approximately 2.5-kb PEAMT mRNA in spinach leaves, which was consistent with the size of the cDNAs that were isolated. Upon salinization, there was an approximately 10-fold increase in PEAMT mRNA abundance, and 10 this was accompanied by an 8-fold rise in enzyme activity (FIG. 6). This indicates that the salt-induction of PEAMT activity reported previously (Summers and Weretilnyk, 1993; Weretilnyk *et al.*, 1995) results principally, if not solely, from increased gene expression.

#### 5.1.3 DISCUSSION

15 The complementation strategy used to clone PEAMT was not narrowly specific because in principle the *cho2* mutation (in Ptd-EA *N*-methyltransferase) could have been complemented by plant genes encoding *N*-methyltransferases acting on Ptd-EA or free EA as well as on P-EA. It is therefore significant that all 24 complemented *S. pombe* strains that were recovered harbored a plasmid encoding PEAMT, for this implies that 20 *N*-methyltransferases acting on Ptd-EA or EA are either absent or expressed at far lower levels. This evidence reinforces the conclusion from *in vivo* radiolabeling studies (Rhodes and Hanson, 1993; Hanson and Rhodes, 1983; Summers and Weretilnyk, 1993; Datko and Mudd, 1988a) and enzyme assays (Datko and Mudd, 1988b; Summers and Weretilnyk, 1993; Weretilnyk *et al.*, 1995) that in plant leaves the first methylation in the synthesis of Cho 25 moieties takes place exclusively at the phosphobase level.

The large size and tandem-domain structure of PEAMT indicate that it is the product of a fusion between two related methyltransferase genes. That this fusion is an ancient one that predates the radiation of the Angiosperms is shown by the occurrence of PEAMT homologs in species from four diverse families (*Chenopodiaceae*, *Malvaceae*, 30 *Brassicaceae* and *Gramineae*) whose progenitors diverged >125 million years (Myr) ago

(Crane *et al.*, 1995). In view of the probable evolutionary age of the fused enzyme it is particularly interesting that at least one of its domains retains what is presumably its ancestral activity (conversion of P-EA to P-MME) when expressed separately.

Spinach PEAMT is a novel protein, and there is no precedent for the synthesis of Cho moieties *via* an enzyme composed of two fused *N*-methyltransferases. Three separate enzymes are required to convert P-EA to P-Cho in nerve tissues (Mukherjee *et al.*, 1995), and two separate enzymes carry out the Ptd-EA → Ptd-MME and Ptd-MME →→ Ptd-Cho steps in fungi (Kanipes and Henry, 1997). *R. sphaeroides* and liver have enzymes that mediate all three methylations in Ptd-Cho synthesis, but these are small (22-23 kDa) proteins that appear to have only one methyltransferase domain (Vance *et al.*, 1997; Arondel *et al.*, 1993). It is, however, noteworthy that the genome of the nematode *Caenorhabditis elegans* includes a hypothetical protein (GenBank Accession No. AAB04824, 437 residues) that shares homology throughout its length with PEAMT and has two methyltransferase domains. Other organisms may therefore have two-domain phosphobase *N*-methyltransferases that remain to be discovered. It is also noteworthy that soybean cell extracts mediate only the first methylation of P-EA (Datko and Mudd, 1988b; Rhodes and Hanson, 1993; Hanson and Rhodes, 1983; Summers and Weretilnyk, 1993; Datko and Mudd, 1988a) and that spinach leaves have P-MME and P-DME *N*-methyltransferase activities that appear to be independent of PEAMT activity (Smith *et al.*, 1999; Weretilnyk *et al.*, 1995). Plants may therefore have other phosphobase *N*-methyltransferases besides PEAMT.

These data show for the first time that PEAMT is regulated at the gene level, because the mRNA is strongly induced in leaves by salinization. They also confirm that PEAMT is regulated at the enzyme level, because the activity is sensitive to the reaction product P-Cho. The activity was 95% inhibited at a P-Cho concentration of 10 mM, the physiological level in spinach leaf cytosol (Bligny *et al.*, 1990). As this inhibition was measured in the presence of a saturating P-EA concentration, the degree of inhibition *in vivo* is probably greater. Together, these findings support the view that PEAMT is the committing step in the synthesis of Cho moieties in plants and that it exerts major control over the flux to P-Cho and its metabolites (Datko and Mudd, 1988a; Mudd and Datko, 1989a; Mudd and Datko, 1989b). Specifically in spinach, the salt-induction of PEAMT fits

with the increased demand for Cho to support GlyBet accumulation, for which the two biosynthetic enzymes are also induced at the gene level by salinity (Rhodes and Hanson, 1993; Rathinasabapathi *et al.*, 1997).

The cloning and characterization of PEAMT was driven by the need to understand the 5 pathway and regulation of Cho synthesis in plants, in order to engineer an enhanced Cho supply to support GlyBet synthesis (Nuccio *et al.*, 1998; Nuccio *et al.*, 1999). In this context, it is convenient that a single gene encodes all three *N*-methyltransferase activities needed to produce P-Cho. However, the strong feedback inhibition of native PEAMT by P-Cho could render raising the level of PEAMT relatively ineffective in increasing the flux to P-Cho. The 10 fact that the N-terminal domain expressed separately is less sensitive to feedback control may therefore be valuable in engineering. Since most if not all plants have the capacity to carry out the second and third methylations of Cho synthesis at the phosphatidylbase level (Datko and Mudd, 1988b; Rhodes and Hanson, 1993), and there may also be phosphobase *N*-methyltransferases specific for these steps (Smith *et al.*, 1999; Weretilnyk *et al.*, 1995), a 15 partially desensitized enzyme able to carry out just the first methylation could prove more effective than native PEAMT in enhancing Cho biogenesis.

## 5.2 EXAMPLE 2 – SEQUENCE OF PEAMT POLYPEPTIDES AND POLYNUCLEOTIDES

### 5.2.1 SEQUENCE OF SPINACH PEAMT-ENCODING DNA (SEQ ID NO:1)

20 CATTCAATTGAAGCGTGGAAAGTAGTAGTTGTGGTAGAGTGAATTGATACTCCTACTGCTCATGCGGCAGAG  
AGGCAGGGCTTCGAACCGTAGATCCAGGACTTTCTCGTTCTGCATTGCCATTGAGGGTCACTAATACTTT  
AACTATCTCCTCTTTCTTCCCACAATTCTCGCTTACGCACATTAATCTCACCTATTTCTAGCTTC  
TTCATTTCTCAATCAATCTCTCGTGTATTATGGCCGCTTCAGCTATGGGAGTGGTCAAGAGAGAGAGGTGT  
TCAAGAAATACTGGATTGAACACTCTGTGATTGACTGTTGAGGCTATGATGCTTGAAGAGCTTCAGAT  
15 CTTGACAAAGTGGAGCGACCTGAGGTACTTCCATGCTTCCACCTTATGAAGGAAGTCTGCTTGAAGAACTCGG  
TGCTGGTATTGGCTTTACTGGTGAATTGGCCGAGAAAGCTAGCCAGGTCACTGCTCTGGATTTCATTGAGA  
GTGTTATAAAGAAGAATGAAAGCATAAATGGCATACAAAAATGTGAAGTTATGTGTGCTGATGTGACATCT  
CCAAGTCTCAACATTCACCAAATCCGTGGATATCATATTCTCCAATTGGCTACTCATGTATCTTCTGATGA  
AGAGGTTGAGCGTCTGGTTGAAAGGATGTTGAATGGTGAAGGCCAGGAGGATACATTCTCAGAGAATCTT  
30 GTTTCAATCAGGAGATCACAAGCGAAAAGCAATCCAACCCACTACCGTGAACCTAGGTTCTACACCAAG  
ATCTTCAAAAGAATGCCATATGCAAGATGATTCTGGGACTCCTATGAGCTCTCCCTAATTGGCTGCAAATGTAT  
TGGAGCTTATGTCAAAAGCAAGAAGAATCAGAACAGATAAGCTGGTTATGGCAGAAAGTTGATTGAGGGATG  
ACAAGGGTTCCAGCGATTCTGGATTCTAGTCAATACAAGTTAACAGCATACTGCGTTATGAGCGTGTATT  
GGCCTGGTTATGTAGTACCGGAGGACTGAAACAAACCAAGGAGTTGTATCAAAGCTTGACTGAAAGCCTGG  
35 CCAGAAGGTCTAGATGTGGTTGTGGCATAGGTGGAGGTGATTTTACATGGCAGAGAACTATGATGTTGAGG  
TTGTTGGAATTGATCTCTCCATTAATATGATTCTTTGCCCTTGAGCGCTCAATTGGCTCAAATGTGCTGTT  
GAGTTTGAGGTGGCAGATTGCAACCAAGAAAGATTACCCCTGAAAACCTCTTGTATGTCATCTACAGCCGTGATAC  
CATTCTGCATATTCAAGGACAAACCTGCTTATTAGATCCTTCCACAAATGGTGAACAGGAGGATGACTTCATTGATG  
40 TTATTAGTGAACAGAGTGTGGTACACCTCAGCTGAATTGCTGCATACATCAGGCAGAGGGGATAT  
GATCTCCACGATGTGAAGGCATATGGCAAGATGCTTAAAGATGCTGGATTGTTGAGGTTATGCTGAGAATAG  
GACTGACCAGTTCAAGTCTGCAGAAGGAACTAGATGCTTGTGAACAGGAGAAGGGATGACTTCATTGATG  
ATTCTCTGAGGAGGATTATAACGACATAGTTGATGGTTGGAAGGCCAAGTGGTGGAGGACTACAGAGGGTGG

5 CAACAATGGGTTTGTTCATTGCCAAGAAAATGTGAAGAATGAGCTGGTGAAGCAGCACGGTGCCTTTCTA  
 GTATTAGTTATCAATGTATTTCAGTTCATGGACTGTATATGCAAAACTACCAATAAGCTGTGAGTTGCAA  
 CTGAAAGATGATTCTTATAGTCACCTCTGAATTAGCACAAGCAGTGAAGTTCGATAAGAAACTGAAGGGAAC  
 TCATGGAGTTGCAGACGAAATCATAAAACGGCAGAACCCACTCTCTATATAGAGATCTAGGGTTAAGTTATG  
 10 15 TGTGGATTGAAGTCGAAACTCTTGTAGTAATGTATTAGTTGAAAGTGGAACTTGCACACACTTATT  
 CACAAGTGTGTAGGGAAATATGGATTTGTATTAGTATGTACTGACTTAGTTGTAAAAGGAACTTCCTACG  
 TTTCTCTGGCA

10 **5.2.2 SEQUENCE OF SPINACH PEAMT POLYPEPTIDE (SEQ ID NO:2)**

15 MAASAMGVLQEREVFKKYWIEHSVDLTVEAMMLDSQASDLKVERPEVLSMLPPYEGKSVLELGAGIGRFTGEL  
 AEKASQVIALDFIESVIKKNESINGHYKNVKFMCAVTSPSLNISPNSVDIIFSNWLLMYLSDEEVERLVERML  
 KWLKPGGYIFFRESCFHQSGDHKRKSNPHTYREPRFYTKIFKECHMQDDSGNSYELSLIGCKCIGAYVKSKKNQ  
 NOISWLWQKVDSEDDKGQFLDSSQYKFNSILRYERVFGPGYVSTGGLETTKEFVSKLDLKGQKVLDVGC  
 20 25 GGGDFYMAENYDVEVVGIDLSINMISALERSIGLKCAFEVADCTKDYPENSFDVIYSRDTILHIQDKPAL  
 FRSFHKWLKPGGKVLIISDYCSAGTPSAEFAAYIRQRGYDLHDVKAYGKMLKDAGFVEVIAENRTDQFQVLQK  
 ELDALEQEKDDFIDDFSEEDYNDIVDGWKAALKVRTTEGEQQWGLFIAKKM

20 **5.2.3 SEQUENCE OF GENE ENCODING SPINACH  $\Delta$ PEAMT**

25 **POLYPEPTIDE (SEQ ID NO:3)**

25 CATTCAATTGAAGCGTGGAAAGTAGTAGTTGTGGTAGAGTGAATTGATACTCCTACTGCTCATGCGGCAGAG  
 AGGCAGGGCTTCGAACCGTAGATCCAGGACTTTCTCGTTCTCGCATGCCATTGAGGGTCACTAATACTTT  
 AACTATCCTCTCTTTCTTCCCACAATTCTCGTTTACGCACATTAATCACCTATTCTAGCTTC  
 30 35 TTCATTTCTCAATCAATCTCTCGTTATTATGCCGCTTCAGCTATGGAGTGTGCAAGAGAGAGAGGTGT  
 TCAAGAAATACTGGATTGAACACTCTGTTGACTGTTGAGGCTATGATGCTGATTACAAGCTTCAGAT  
 CTTGACAAAGTGGAGCGACCTGAGGTACTTCCATGCTCCACCTATGAAGGAAAGTCTGCTTAACTCGG  
 TGCTGGTATTGGCTTTACTGGTGAATTGCCAGAAAGCTAGCCAGGTATCGCTCTGGATTCATTGAGA  
 GTGTTATAAAGAAGAATGAAAGCATAAATGGCATTACAAAATGTGAAGTTATGTGCTGATGTGACATCT  
 CCAAGTCTCAACATTCACCAAATTCCGGATATCATATTCTCAATTGGCTACTCATGTATCTTCTGATGA  
 40 45 AGAGGTTGAGCGCTGGTTGAAAGGATGTTGAAATGGTGAAGCCAGGAGGATACATTTCTCAGAGAATCTT  
 GTTTCATCAATCAGGAGATCACAAGCGAAAAGCAATCCAACCCACTACCGTGAACCTAGGTTACACCAAG  
 ATCTTCAAGAATGCCATATGCAAGATGATTCTGGGAACTCCTATGAGCTCTCCCTAATTGGCTGCAAATGTAT  
 TGGAGCTATGTCAAAGAAGAATCAGAACAGATAAGCTGGTTATGGCAGAAAGTTGATTCAAGGATG  
 ACAAGGGGTTCCAGCGATTCTGGATTCTAGTCAATACAAGTTAACAGCATACTCGCTTATGAGCGTGTATT  
 50 55 GGTCTGGTTATGTAGTACCGGAGGACTCGAAACAACCAAGGAGTTGTATCAAAGCTTGACTTGAAGCCTGG  
 GATCCCCGG

5.2.4 SEQUENCE OF SPINACH  $\Delta$ PEAMT POLYPEPTIDE (SEQ ID NO:4)

MAASAMGVLQEREVFKKYWIEHSVDLTVEAMMLDSQASDLKVERPEVLSMLPPYEGKSVLELGAGIGRFTGEL  
 40 AEKASQVIALDFIESVIKKNESINGHYKNVKFMCAVTSPSLNISPNSVDIIFSNWLLMYLSDEEVERLVERML  
 KWLKPGGYIFFRESCFHQSGDHKRKSNPHTYREPRFYTKIFKECHMQDDSGNSYELSLIGCKCIGAYVKSKKNQ  
 NOISWLWQKVDSEDDKGQFLDSSQYKFNSILRYERVFGPGYVSTGGLETTKEFVSKLDLKGQIPG

6. REFERENCES

45 The following literature citations as well as those cited above are incorporated in  
 pertinent part by reference herein for the reasons cited in the above text:

U. S. Patent No. 4,196,265, issued Apr. 1, 1980.

- U. S. Patent No. 4,342,832, issued Aug. 3, 1982.
- U. S. Patent No. 4,356,270, issued Oct. 26, 1982.
- U. S. Patent No. 4,362,817, issued Dec. 7, 1982.
- U. S. Patent No. 4,371,625, issued Feb. 1, 1983.
- 5 U. S. Patent No. 4,554,101, issued Nov. 19, 1985.
- U. S. Patent No. 4,683,195, issued Jul. 28, 1987.
- U. S. Patent No. 4,683,202, issued Jul. 28, 1987.
- U. S. Patent No. 4,800,159, issued Jan. 24, 1989.
- U. S. Patent No. 4,873,192, issued Oct. 10, 1989.
- 10 U. S. Patent No. 4,943,674, issued Jul. 24, 1990.
- U. S. Patent No. 4,965,188, issued Oct. 23, 1990.
- U. S. Patent No. 5,097,025, issued Mar. 17, 1992.
- U. S. Patent No. 5,106,739, issued Apr. 21, 1992.
- U. S. Patent No. 5,110,732, issued May 5, 1992.
- 15 U. S. Patent No. 5,139,954, issued Aug. 19, 1992.
- U. S. Patent No. 5,176,995, issued Oct. 15, 1991.
- U. S. Patent No. 5,177,011, issued Jan. 5, 1993.
- U. S. Patent No. 5,324,253, issued Jun. 28, 1994.
- U. S. Patent No. 5,378,619, issued Jan. 3, 1995.
- 20 U. S. Patent No. 5,399,680, issued Mar. 21, 1995.
- U. S. Patent No. 5,401,836, issued Mar. 28, 1995.
- U. S. Patent No. 5,405,765, issued Apr. 11, 1995.
- U. S. Patent No. 5,436,393, issued Jul. 25, 1995.
- U. S. Patent No. 5,441,884, issued Aug. 15, 1995.
- 25 U. S. Patent No. 5,442,052, issued Aug. 15, 1995.
- U. S. Patent No. 5,447,858, issued Sep. 5, 1995.
- U. S. Patent No. 5,459,252, issued Oct. 17, 1995.
- U. S. Patent No. 5,472,869, issued Dec. 5, 1995.
- U. S. Patent No. 5,484,956, issued Jan. 16, 1996.
- 30 U. S. Patent No. 5,491,288, issued Feb. 13, 1996.

- U. S. Patent No. 5,504,200, issued Apr. 2, 1996.
- U. S. Patent No. 5,530,196, issued Jun. 25, 1996.
- U. S. Patent No. 5,538,879, issued Jul. 23, 1996.
- U. S. Patent No. 5,539,082, issued Jul. 23, 1996.
- 5 U. S. Patent No. 5,576,198, issued Nov. 19, 1996.
- U. S. Patent No. 5,589,583, issued Dec. 31, 1996.
- U. S. Patent No. 5,589,610, issued Dec. 31, 1996.
- U. S. Patent No. 5,589,614, issued Dec. 31, 1996.
- U. S. Patent No. 5,595,896, issued Jan. 21, 1997.
- 10 U. S. Patent No. 5,608,144, issued Mar. 4, 1997.
- U. S. Patent No. 5,610,042, issued Mar. 11, 1997.
- U. S. Patent No. 5,610,288, issued Mar. 11, 1997.
- U. S. Patent No. 5,614,399, issued Mar. 25, 1997.
- U. S. Patent No. 5,629,183, issued May 13, 1997.
- 15 U. S. Patent No. 5,633,363, issued May 27, 1997.
- U. S. Patent No. 5,633,439, issued May 27, 1997.
- U. S. Patent No. 5,633,440, issued May 27, 1997.
- U. S. Patent No. 5,633,441, issued May 27, 1997.
- U. S. Patent No. 5,646,333, issued Jul. 8, 1997.
- 20 U. S. Patent No. 5,659,124, issued Aug. 19, 1997.
- U. S. Patent No. 5,689,040, issued Nov. 18, 1997.
- U. S. Patent No. 5,689,049, issued Nov. 18, 1997.
- U. S. Patent No. 5,689,051, issued Nov. 18, 1997.
- U. S. Patent No. 5,689,056, issued Nov. 18, 1997.
- 25 U. S. Patent No. 5,712,112, issued Jan. 27, 1998.
- U. S. Patent No. 5,718,709, issued Feb. 17, 1998.
- U. S. Patent No. 5,719,262, issued Feb. 17, 1998.
- U. S. Patent No. 5,736,336, issued Apr. 17, 1998.
- U. S. Patent No. 5,739,119, issued Apr. 14, 1998.
- 30 U. S. Patent No. 5,759,829, issued Jun. 2, 1998.

U. S. Patent No. 5,766,855, issued Jun. 16, 1998.

U. S. Patent No. 5,773,571, issued Jun. 30, 1998.

U. S. Patent No. 5,786,461, issued Jul. 28, 1998.

U. S. Patent No. 5,789,573, issued Aug. 4, 1998.

5 U. S. Patent No. 5,801,154, issued Sept. 1, 1998.

U. S. Patent No. 5,859,347, issued Jan. 12, 1999.

U. S. Patent No. 5,866,793, issued Feb. 2, 1999.

U. S. Patent No. 5,874,265, issued Feb. 23, 1999.

U. S. Patent No. 5,874,626, issued Feb. 23, 1999.

10 U. S. Patent No. 5,886,244, issued Mar. 23, 1999.

U. S. Patent No. 6,023,013, issued Feb. 8, 2000.

Eur. Pat. Appl. Publ. No. EP 0120516.

Eur. Pat. Appl. Publ. No. EP 75444.

Int. Pat. Appl. Publ. No. WO 84/02913.

15 Abdullah *et al.*, *Biotechnology*, 4:1087, 1986.

Alexander, *Methods Enzymol.*, 154:41-64, 1987.

Altschul *et al.*, "Basic local alignment search tool," *J. Mol. Biol.*, 215:403-410, 1990.

An *et al.*, *EMBO J.*, 4:277-287, 1985.

Andriamampary, Freysz, Kanfer, Dreyfus and Massarelli, *J. Neurochem.*, 56:1845-1850,

20 1991.

Arondel, Benning and Somerville, *J. Biol. Chem.*, 268:16002-16008, 1993.

Arz *et al.*, *Biochim. Biophys. Acta*, 1218(3):447-452, 1994.

Bagdasarian, Lurz, Ruckert, Franklin, Bagdasarian, Frey, Timmis, "Specific-purpose plasmid cloning vectors. II. Broad host range, high copy number, RSF1010-derived

25 vectors, and a host-vector system for gene cloning in *Pseudomonas*," *Gene*, 16(1-3):237-247, 1981.

Ballas *et al.*, *Nucl. Acids Res.*, 17:7991-7903, 1989.

Baum *et al.*, *Appl. Environ. Microbiol.*, 56:3420-3428, 1990.

Baumlein, Boerjan, Nagy, Panitz, Inze, Wobus, "Upstream sequences regulating legumin gene expression in heterologous transgenic plants," *Mol. Gen. Genet.*, 225(1):121-128, 1991.

Beaudouin, Haurat, Lafitte and Renaud, *J. Neurochem.*, 61:928-935, 1993.

5 Benbrook *et al.*, In: *Proceedings Bio Expo 1986*, Butterworth, Stoneham, MA, pp. 27-54, 1986.

Berna and Bernier, "Regulated expression of a wheat germin gene in tobacco: oxalate oxidase activity and apoplastic localization of the heterologous protein," *Plant Mol. Biol.*, 33(3):417-429, 1997.

10 Bevan *et al.*, *Nature*, 304:184, 1983.

Bligny, Gardestrom, Roby and Douce, *J. Biol. Chem.*, 265:1319-1326, 1990.

Boffa, Carpaneto, Allfrey, *Proc. Natl. Acad. Sci. USA*, 92:1901-1905, 1995.

Boronat, Martinez, Reina, Puigdomenech, Palau, "Isolation and sequencing of a 28 kd gluteline-2 gene from maize: Common elements in the 5' flanking regions among 15 zein and glutelin genes," *Plant Sci.*, 47:95-102, 1986.

Bytebier *et al.*, "t-DNA organization in tumor cultures and transgenic plants of the monocotyledon *apsparagus-officinalis*," *Proc. Natl. Acad. Sci. USA*, 84(15):5345-5349, 1987.

Callis, Fromm, Walbot, "Introns increase gene expression in cultured maize cells," *Genes 20 Devel.*, 1:1183-1200, 1987.

Campbell, In: *Monoclonal Antibody Technology, Laboratory Techniques in Biochemistry and Molecular Biology*, Burden and Von Knippenberg (Eds.), Vol. 13, pp. 75-83, Elsevier, Amsterdam, 1984.

Capecchi, "High efficiency transformation by direct microinjection of DNA into cultured 25 mammalian cells," *Cell*, 22(2):479-488, 1980.

Cheng *et al.*, *Proc. Natl. Acad. Sci. USA*, 95(6):2767-2772, 1998.

Christensen, Sharrock, Quail, "Maize polyubiquitin genes: Structure, thermal perturbation of expression and transcript splicing, and promoter activity following transfer to protoplasts by electroporation," *Plant Mol. Biol.*, 18:675-689, 1992.

Clapp, "Somatic gene therapy into hematopoietic cells. Current status and future implications," *Clin. Perinatol.*, 20(1):155-168, 1993.

Conway and Wickens, *In: RNA Processing*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, p. 40, 1988.

5 Cook and Wagner, *Proc. Natl. Acad. Sci. USA*, 81:3631-3634, 1984.

Corey, *Trends Biotechnol.*, 15(6):224-229, 1997.

Corpet, *Nucleic Acids Res.*, 16:10881-10890, 1988.

Crane, Friis and Pedersen, *Nature*, 374:27-33, 1995.

Cristou *et al.*, *Plant Physiol.*, 87:671-674, 1988.

10 Curiel, Agarwal, Wagner, Cotten, "Adenovirus enhancement of transferrin-polylysine-mediated gene delivery," *Proc. Natl. Acad. Sci. USA*, 88(19):8850-8854, 1991.

Danos and Heard, "Recombinant retroviruses as tools for gene transfer to somatic cells," *Bone Marrow Transplant*, 9(Suppl. 1):131-138, 1992.

Datko and Mudd, *Plant Physiol.*, 88:1338-1348, 1988b.

15 Datko and Mudd, *Plant Physiol.*, 88:854-861, 1988a.

Dean *et al.*, *Nucl. Acids Res.*, 14(5):2229-2240, 1986.

Dennis, Gerlach, Pryor, Bennetzen, Inglis, Llewellyn, Sachs, Ferl, Peacock, "Molecular analysis of the alcohol dehydrogenase (*Adh1*) gene of maize," *Nucl. Acids Res.*, 12:3983-4000, 1984.

20 Eglitis and Anderson, "Retroviral vectors for introduction of genes into mammalian cells," *Biotechniques*, 6(7):608-614, 1988.

Eichholtz *et al.*, "Expression of mouse dihydrofolate reductase gene confers methotrexate resistance in transgenic petunia plants," *Somat. Cell Mol. Genet.*, 13(1):67-76, 1987.

25 Faktor, Kooter, Dixon, Lamb, "Functional dissection of a bean chalcone synthase gene promoter in transgenic tobacco plants reveals sequence motifs essential for floral expression," *Plant Mol. Biol.*, 32(5):849-859, 1996.

Ficker, Kirch, Eijlander, Jacobsen, Thompson, "Multiple elements of the S2-RNase promoter from potato (*Solanum tuberosum* L.) are required for cell type-specific expression in transgenic potato and tobacco," *Mol. Gen. Genet.*, 257(2):132-142, 1998.

30

Fraley *et al.*, *Bio/Technology*, 3:629-635, 1985.

Fraley, Rogers, Horsch, Sanders, Flick, Adams, Bittner, Brand, Fink, Fry, Galluppi, Goldberg, Hoffmann, Woo, "Expression of bacterial genes in plant cells," *Proc. Natl. Acad. Sci. USA*, 80(15):4803-4087, 1983.

5 French, Janda, Ahlquist, "Bacterial gene inserted in an engineered RNA virus: efficient expression in monocotyledonous plant cells," *Science*, 231:1294-1297, 1986.

Fromm, Taylor, Walbot, "Expression of genes transferred into monocot and dicot plant cells by electroporation," *Proc. Natl. Acad. Sci. USA*, 82(17):5824-5828, 1985.

Fujimura *et al.*, *Plant Tissue Cult. Lett.*, 2:74, 1985.

10 Fujioka, *Int. J. Biochem.*, 24:1917-1924, 1992.

Gallie and Young, "The regulation of expression in transformed maize aleurone and endosperm protoplasts," *Plant Physiol.*, 106:929-939, 1994.

Gallie, Feder, Schimke, Walbot, "Post-transcriptional regulation in higher eukaryotes: the role of the reporter gene in controlling expression," *Mol. Gen. Genet.*, 228:258-264, 1991.

15 Gallie, Lucas, Walbot, "Visualizing mRNA expression in plant protoplasts: factors influencing efficient mRNA uptake and translation," *Plant Cell*, 1:301-311, 1989.

Gallie, Sleat, Turner, Wilson, "Mutational analysis of the tobacco mosaic virus 5'-leader for altered ability to enhance translation," *Nucl. Acids Res.*, 16:883-893, 1988.

20 Gallie, Sleat, Watts, Turner, Wilson, "A comparison of eukaryotic viral 5'-leader sequences as enhancers of mRNA expression *in vivo*," *Nucl. Acids Res.*, 15:8693-8711, 1987b.

Gallie, Sleat, Watts, Turner, Wilson, "The 5'-leader sequence of tobacco mosaic virus RNA enhances the expression of foreign gene transcripts *in vitro* and *in vivo*," *Nucl. Acids Res.*, 15:3257-3273, 1987a.

25 GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Drive, Madison, Wisconsin, USA.

Gary, Lin, Yang, Herschman and Clarke, *J. Biol. Chem.*, 271:12585-12594, 1996.

Gefter, Margulies, Scharff, "A simple method for polyethylene glycol-promoted hybridization of mouse myeloma cells," *Somat. Cell Genet.*, 3(2):231-236, 1977.

Gehrke, Auron, Quigley, Rich, Sonenberg, "5'-Conformation of capped alfalfa mosaic virus ribonucleic acid 4 may reflect its independence of the cap structure or of cap-binding protein for efficient translation," *Biochemistry*, 22:5157-5164, 1983.

5 Gil and Proudfoot, "A sequence downstream of AAUAAA is required for rabbit beta-globin mRNA 3'-end formation," *Nature*, 312(5993):473-474, 1984.

Goding, "Monoclonal Antibodies: Principles and Practice," pp. 60-74. 2nd Edition, Academic Press, Orlando, FL, 1986.

10 Goelet, Lomonossoff, Butler, Akam, Gait, Karn, "Nucleotide sequence of tobacco mosaic virus RNA," *Proc. Natl. Acad. Sci. USA*, 79:5818-5822, 1982.

Good and Nielsen, *Antisense Nucleic Acid Drug Dev.*, 7(4):431-437, 1997.

Gothel *et al.*, *Eur. J. Biochem.*, 244(1):59-65, 1997.

Graham and van der Eb, "Transformation of rat cells by DNA of human adenovirus 5," *Virology*, 54(2):536-539, 1973.

15 Graham, Craig, Waterhouse, "Expression patterns of vascular-specific promoters ROI C and Sh in transgenic potatoes and their use in engineering PLRV-resistant plants," *Plant Mol. Biol.*, 33(4):729-735, 1997.

Grosset, Alary, Gautier, Menossi, Martinez-Izquierdo, Joudrier, "Characterization of a barley gene coding for an alpha-amylase inhibitor subunit (Cmd protein) and analysis of its promoter in transgenic tobacco plants and in maize kernels by microprojectile bombardment," *Plant Mol. Biol.*, 34(2):331-338, 1997.

20 Guerineau *et al.*, *Mol. Gen. Genet.*, 262:141-144, 1991.

Hanson and Rhodes, *Plant Physiol.*, 71:692-700, 1983.

25 Hanvey *et al.*, *Science*, 258:1481-1485, 1992.

Harlow and Lane, *In: Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1988.

Herrera-Estrella *et al.*, *Nature*, 303:209, 1983.

Hess, *Intern Rev. Cytol.*, 107:367, 1987.

30 Higgins *et al.*, *Gene*, 73:237-244, 1988.

Higgins *et al.*, CABIOS, 5:151-153, 1989.

Hilber *et al.*, "Biostatic transformation of *Botryotinia fuckeliana*," *Curr. Genet.*, 25(2):124-127, 1994.

Hockema, *In: The Binary Plant Vector System*, Offset-durkkerij, Kanters B.V.,  
5 Alblasserdam, Chapter 5, 1985.

Holsters *et al.*, *Mol. Gen. Genet.*, 163:181-187, 1978.

Hondred, Wadle, Titus and Becker, *Plant Mol. Biol.*, 9:259-275, 1987.

Horsch *et al.*, *Science*, 227:1229-1231, 1985.

Huang *et al.*, *Computer Applications in the Biosciences*, 8:155-165, 1992.

10 Huang, An, McDowell, McKinney, Meagher, "The *Arabidopsis* ACT11 action gene is strongly expressed in tissues of the emerging inflorescence, pollen and developing ovules," *Plant Mol. Biol.*, 33(1):125-139, 1997.

Huang, Hirji, Adam, Rozwadowski, Hammerlindl, Keller and Selvaraj, *Plant Physiol.*, in press, 2000.

15 Hudspeth and Grula, "Structure and expression of the maize gene encoding the phosphoenolpyruvate carboxylase isozyme involved in C4 photosynthesis," *Plant Mol. Biol.*, 12:579-589, 1989.

Ingelbrecht, Herman, Dekeyser, Van Montagu, Depicker, "Different 3' end regions strongly influence the level of gene expression in plant cells," *Plant Cell*, 1:671-680, 1989.

20 Jameson and Wolf, "The Antigenic Index: A Novel Algorithm for Predicting Antigenic Determinants", *Compu. Appl. Biosci.*, 4(1):181-6, 1988.

Jobling and Gehrke, "Enhanced translation of chimaeric messenger RNAs containing a plant viral untranslated leader sequence," *Nature*, 325:622-625, 1987.

25 Jones, Dean, Gidoni, Gilbert, Bond-Nutter, Lee, Bedbrook, Dunsmuir, "Expression of bacterial chitinase protein in tobacco leaves using two photosynthetic gene promoters," *Mol. Gen. Genet.*, 212:536-542, 1988.

Jorgensen *et al.*, *Mol. Gen. Genet.*, 207:471, 1987.

Joshi *et al.*, *Nucleic Acids Res.*, 15:9627-9639, 1987.

Kagan and Clarke, *Arch. Biochem. Biophys.*, 310:417-427, 1994.

30 Kaiser and Kezdy, *Science*, 223:249-255, 1984.

Kanipes and Henry, *Biochim. Biophys. Acta*, 1348:134-141, 1997.

Kanipes, Hill and Henry, *Genetics*, 150:553-562, 1998.

Kanipes, Ph.D Thesis. Carnegie Mellon University, 1997.

Klee *et al.*, *Bio/Technology*, 3:637-642, 1985.

5 Klein *et al.*, "Factors influencing gene delivery into Zea mays cells by high velocity microparticles," *Bio/Tech.*, 6:559-563, 1988a.

Klein *et al.*, *Nature*, 327:70, 1987.

Klein *et al.*, *Plant Physiol.*, 91:440-444, 1988b.

Klein *et al.*, *Proc. Natl. Acad. Sci. USA*, 85:4305-4309, 1988c.

10 Klein *et al.*, *Proc. Natl. Acad. Sci. USA*, 85:8502-8505, 1988d.

Kohler and Milstein, "Continuous cultures of fused cells secreting antibody of predefined specificity," *Nature*, 256(5517):495-497, 1975.

Kohler and Milstein, "Derivation of specific antibody-producing tissue culture and tumor lines by cell fusion," *Eur. J. Immunol.*, 6(7):511-519, 1976.

15 Kozak, *Nature*, 308:241-246, 1984.

Koziel *et al.*, *Plant Mol. Biol.*, 32:393-405, 1996.

Koziel, Beland, Bowman, Carozzi, Crenshaw, Crossland, Dawson, Desai, Hill, Kadwell, Launis, Lewis, Maddox, McPherson, Meghji, Merlin, Rhodes, Warren, Wright, Evola, "Field performance of elite transgenic maize plants expressing an 20 insecticidal protein derived from *Bacillus thuringiensis*," *Bio/technology*, 11:194-200, 1993.

Kuby, *Immunology 2nd Edition*, W. H. Freeman & Company, NY, 1994.

Kunkel, *Proc. Natl. Acad. Sci. USA*, 82:488-492, 1985.

Kunkel, Roberts, Zabour, *Methods Enzymol.*, 154:367-382, 1987.

25 Kyozuka *et al.*, "Anaerobic induction and tissue-specific expression of maize Adh1 promoter in transgenic rice plants and their progeny," *Mol. Gen. Genet.*, 228(1-2):40-48, 1991.

Lorz *et al.*, *Mol. Gen. Genet.*, 199:178, 1985.

Luehrsen and Walbot, "Intron enhancement of gene expression and the splicing efficiency of 30 introns in maize cells," *Mol. Gen. Genet.*, 225:81-93, 1991.

Luehrsen, De Wit and Walbot, *Methods Enzymol.*, 216:397-414, 1992.

Luehrsen, In: *The Maize Handbook*, Springer-Verlag New York, Inc., New York, NY, 1994.

Luo *et al.*, *Plant Mol. Biol. Report.*, 6:165, 1988.

Lutcke, Chow, Mickel, Moss, Kern, Scheele, "Selection of AUG initiation codons differs in 5 plants and animals," *EMBO J.*, 6:43-48, 1987.

Maas, Laufs, Grant, Korfhage, Werr, "The combination of a novel stimulatory element in the first exon of the maize *shrunken-1* gene with the following intron enhances reporter gene expression 1000-fold," *Plant Mol. Biol.*, 16:199-207, 1991.

Maddock *et al.*, *Third International Congress of Plant Molecular Biology*, Abstract 372, 10 1991.

Maloy *et al.*, In: *Microbial Genetics*, 2nd Ed., Jones and Bartlett Publishers, Boston, MA, 1994.

Marcotte *et al.*, *Nature*, 335:454, 1988.

Mascerenhas, Mettler, Pierce, Lowe, "Intron mediated enhancement of heterologous gene 15 expression in maize," *Plant Mol. Biol.*, 15:913-920, 1990.

Maundrell, *Gene*, 123:127-130, 1993.

McCabe *et al.*, *Biotechnology*, 6:923-926, 1988.

McCornick *et al.*, *Plant Cell Reports*, 5:81-84, 1986.

McDevitt, Imperiale, Ali, Nevins, "Requirement of a downstream sequence for generation of 20 a poly(A) addition site," *Cell*, 37(3):993-999, 1984.

McElroy, Zhang, Cao, Wu, "Isolation of an efficient promoter for use in rice transformation," *Plant Cell*, 2(2):163-171, 1990.

McNeil, Rhodes, Nuccio and Hanson, *Plant Physiol.*, submitted, 2000.

Mogen *et al.*, *Plant Cell*, 2:1261-1272, 1990.

Mollegaard, Buchardt, Egholm, Nielsen, "Peptide nucleic acid. DNA strand displacement 25 loops as artificial transcription promoters," *Proc. Natl. Acad. Sci. USA*, 91(9):3892-3895, 1994.

Mudd and Datko, *Plant Physiol.*, 90:296-305, 1989a.

Mudd and Datko, *Plant Physiol.*, 90:306-310, 1989b.

Mudd *et al.*, *Plant Physiol.*, 93:623-630, 1990.

Mukherjee, Freysz and Kanfer, *Neurochem. Res.*, 20:1233-1237, 1995.

Munroe *et al.*, *Gene*, 91:151-158, 1990.

Nairn and Ferl, *J. Mol. Evol.*, 27:133-141, 1988.

Nawrath, Poirier, Somerville, "Targeting of the polyhydroxybutyrate biosynthetic pathway to  
5 the plastids of *Arabidopsis thaliana* results in high levels of polymer accumulation," *Proc. Natl. Acad. Sci. USA*, 91:12760-12764, 1994.

Needleman *et al.*, *J. Mol. Biol.*, 48:443, 1970.

Neuhaus *et al.*, *Theor. Appl. Genet.*, 75:30, 1987.

Nielsen *et al.*, "Peptide nucleic acids (PNAs): potential antisense and anti-gene agents,"  
10 *Anticancer Drug Des.*, 8(1):53-63, 1993.

Nielsen, Egholm, Berg, Buchardt, "Sequence-selective recognition of DNA by strand  
displacement with a thymine-substituted polyamide," *Science*, 254(5037):1497-  
1500, 1991.

Nuccio, Rhodes, McNeil and Hanson, *Curr. Opin. Plant Biol.*, 2:128-134, 1999.

15 Nuccio, Russell, Nolte, Rathinasabapathi, Gage and Hanson, *Plant J.*, 16:487-496, 1998.

Oard, Paige, Dvorak, "Chimeric gene expression using maize intron in cultured cells of  
breadwheat," *Plant Cell. Rep.*, 8:156-160, 1989.

Olson, Porter, Rubinstein, Silver, "Mercuric reductase enzyme from a mercury-volatilizing  
strain of *Thiobacillus ferrooxidans*," *J. Bacteriol.*, 151(3):1230-1236, 1982.

20 Omirulleh, Abraham, Golovkin, Stefanov, Karabaev, Mustardy, Morocz, Dudits, "Activity of  
a chimeric promoter with the doubled CaMV 35S enhancer element in protoplast-  
derived cells and transgenic plants in maize," *Plant Molecular Biology*,  
21(3):415-428, 1993.

Orum, Nielsen, Egholm, Berg, Buchardt, Stanley, "Single base pair mutation analysis by  
25 PNA directed PCR<sup>TM</sup> clamping," *Nucl. Acids Res.*, 21(23):5332-5336, 1993.

Orum, Nielsen, Jorgensen, Larsson, Stanley, Koch, "Sequence-specific purification of  
nucleic acids by PNA-controlled hybrid selection," *BioTechniques*, 19(3):472-  
480, 1995.

Pearson *et al.*, *Proc. Natl. Acad. Sci. USA*, 85:2444, 1988.

30 Pena *et al.*, *Nature*, 325:274, 1987.

Perlak, Deaton, Armstrong, Fuchs, Sims, Greenplate, Fischhoff, "Insect resistant cotton plants," *Bio/technology*, 8:939-943, 1990.

Perlak, Fuchs, Dean, McPherson, Fischhoff, "Modification of the coding sequence enhances plant expression of insect control protein genes," *Proc. Natl. Acad. Sci. USA*, 88:3324-3328, 1991.

5 Perlak, Stone, Muskopf, Peterson, Parker, McPherson, Wyman, Love, Reed, Biever, Fischhoff, "Genetically improved potatoes: protection from damage by Colorado potato beetles," *Plant Mol. Biol.*, 22:313-321, 1993.

10 Perry-O'Keefe, Yao, Coull, Fuchs, Egholm, "Peptide nucleic acid pre-gel hybridization: an alternative to southern hybridization," *Proc. Natl. Acad. Sci. USA*, 93(25):14670-14675, 1996.

Person *et al.*, *Methods of Molecular Biology*, 24:307-331, 1994.

15 Poogin and Skryabin, "The 5' untranslated leader sequence of potato virus X RNA enhances the expression of the heterologous gene *in vivo*," *Mol. Gen. Genet.*, 234:329-331, 1992.

Potrykus, Paszkowski, Saul, Petruska, Shillito, "Molecular and general genetics of a hybrid foreign gene introduced into tobacco by direct gene transfer," *Mol. Gen. Genet.*, 199(2):169-177, 1985.

20 Prokop and Bajpai, "Recombinant DNA Technology I," *Ann. N. Y. Acad. Sci.*, Vol. 646, 1991.

Proudfoot, *Cell*, 64:671-674, 1991.

Rathinasabapathi, Burnet, Russell, Gage, Liao, Nye, Scott, Golbeck and Hanson, *Proc. Natl. Acad. Sci. USA*, 94:3454-3458, 1997.

Rhodes and Hanson, *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, 44:357-384, 1993.

25 Rogers *et al.*, *In: Methods For Plant Molecular Biology*, Weissbach and Weissbach, Eds., Academic Press Inc., San Diego, CA, 1988.

Russell and Fromm, "Tissue-specific expression in transgenic maize for four endosperm promoters from maize and rice," *Transgenic Res.*, 6(2):157-168, 1997.

122

Sadofsky and Alwine, "Sequences on the 3' side of hexanucleotide AAUAAA affect efficiency of cleavage at the polyadenylation site," *Molec. Cell. Biol.*, 4(8):1460-1468, 1984.

5 Sambrook *et al.*, In: *Molecular Cloning: A Laboratory Manual*, 2nd Ed.; Cold Spring Harbor Laboratory Press, Plainview, New York, 1989.

Sanfacon *et al.*, *Genes Dev.*, 5:141-149, 1991.

Segal, "Biochemical Calculations" 2nd Edition, John Wiley & Sons, New York, 1976.

Shaw and Kamen, "A conserved AU sequence from the 3' untranslated region of GM-CSF mRNA mediates selective mRNA degradation," *Cell*, 46(5):659-667, 1986.

10 Shaw and Kamen, In: *RNA Processing*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, p. 220, 1987.

Sleat, Gallie, Jefferson Bevan, Turner, Wilson, "Characterization of the 5'-leader sequence of tobacco mosaic virus RNA as a general enhancer of translation *in vitro*," *Gene*, 217:217-225, 1987.

15 Sleat, Hull, Turner, Wilson, "Studies on the mechanism of translational enhancement by the 5'-leader sequence of tobacco mosaic virus RNA," *Eur. J. Biochem.*, 175:75-86, 1988.

Smith and Hood, "*Agrobacterium tumefaciens* transformation of monocotyledons," *Crop Science*, 35:301-309, 1995.

20 Smith *et al.*, *Adv. Appl. Math.*, 2:482, 1981.

Smith, Summers and Weretilnyk, *Physiol. Plant.*, in press, 1999.

Speich *et al.*, *Microbiol.*, 140(Pt6):1273-1284, 1994.

Spielmann *et al.*, *Mol. Gen. Genet.*, 205:34, 1986.

Summers and Weretilnyk, *Plant Physiol.*, 103:1269-1276, 1993.

25 Tanaka, Mita, Ohta, Kyozuka, Shimamoto, Nakamura, "Enhancement of foreign gene expression by a dicot intron in rice but not in tobacco is correlated with an increased level of mRNA and an efficient splicing of the intron," *Nucl. Acids Res.*, 18:6767-6770, 1990.

Thisted, Just, Petersen, Hyldig-Nielsen, Godtfredsen, *Cell Vision*, 3:358-363, 1996.

Tomes *et al.*, "Transgenic tobacco plants and their progeny derived by microprojectile bombardment of tobacco leaves," *Plant Mol. Biol.*, 14:261-268, 1990.

Toriyama *et al.*, *Theor Appl. Genet.*, 73:16, 1986.

Treacy, Hattori, Prud'homme, Barbour, Boutilier, Basczynski, Huang, Johnson, Miki,  
5 "Bnm1, a *Brassica* pollen-specific gene," *Plant Mol. Biol.*, 34(4):603-611, 1997.

Uchimiya *et al.*, *Mol. Gen. Genet.*, 204:204, 1986.

Vain *et al.*, "Osmotic pretreatment enhances particle bombardment-mediated transient and stable transformation of maize," *Plant Cell Rep.*, 12:84-88, 1993.

Van Camp, Herouart, Willekens, Takahashi, Saito, Van Montagu, Inze, "Tissue-specific  
10 activity of two manganese superoxide dismutase promoters in transgenic tobacco," *Plant Physiol.*, 112(2):525-535, 1996.

Vance, Walkey and Cui, *Biochim. Biophys. Acta*, 1348:142-150, 1997.

Vander, Van Montagu, Inze, Boerjan, "Tissue-specific expression conferred by the S-adenosyl-L-methionine synthetase promoter of *Arabidopsis thaliana* in transgenic  
15 poplar," *Plant Cell Physiol.*, 37(8):1108-1115, 1996.

Vasil *et al.*, "Herbicide-resistant fertile transgenic wheat plants obtained by microprojectile bombardment of regenerable embryogenic callus," *Biotechnology*, 10:667-674, 1992.

Vasil, *Biotechnology*, 6:397, 1988.

20 Vasil, Clancy, Ferl, Vasil, Hannah, "Increased gene expression by the first intron of maize *shrunken-1* locus in grass species," *Plant Physiol.*, 91:1575-1579, 1989.

Velten and Schell, "Selection-expression plasmid vectors for use in genetic transformation of higher plants," *Nucl. Acids Res.*, 13(19):6981-6998, 1985.

Velten *et al.*, *EMBO J.*, 3:2723-2730, 1984.

25 Veselkov, Demidov, Nielsen, Frank-Kamenetskii, "A new class of genome rare cutters," *Nucl. Acids Res.*, 24(13):2483-2487, 1996.

Vickers, Griffith, Ramasamy, Risen, Freier, "Inhibition of NF-kappa B specific transcriptional activation by PNA strand invasion," *Nucl. Acids Res.*, 23(15):3003-3008, 1995.

Wagner, Zatloukal, Cotten, Kirlappos, Mechtler, Curiel, Birnstiel, "Coupling of adenovirus to transferrin-polylysine/DNA complexes greatly enhances receptor-mediated gene delivery and expression of transfected genes," *Proc. Natl. Acad. Sci. USA*, 89(13):6099-6103, 1992.

5 Walker and Gaastra, Eds., *Techniques in Molecular Biology*, MacMillan Publishing Company, New York, 1983.

Wang *et al.*, "Transient expression of foreign genes in rice, wheat and soybean cells following particle bombardment," *Plant Mol. Biol.*, 11:433-439, 1988.

Wang, *J. Am. Chem. Soc.*, 118:7667-7670, 1996.

10 Weissbach and Weissbach, Eds., *Methods for Plant Molecular Biology*, Academic Press, Inc., San Diego, CA, 1988.

Weretilnyk, Smith, Wilch and Summers, *Plant Physiol.*, 109:1085-1091, 1995.

Wickens and Stephenson, "Role of the conserved AAUAAA sequence: four AAUAAA point mutants prevent messenger RNA 3' end formation," *Science*, 226(4678):1045-1051, 1984.

15 Wickens *et al.*, In: *RNA Processing*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, p. 9, 1987.

Wilson, Flint, Deaton, Fischhoff, Perlak, Armstrong, Fuchs, Berberich, Parks, Stapp, "Resistance of cotton lines containing a *Bacillus thuringiensis* toxin to pink bollworm (Lepidopteran: Gelechiidae) and other insects," *J. Econ. Entomol.*, 85(4):1516-1521, 1992.

20 Winter, Robinson and Heldt, *Planta*, 193:530-535, 1994.

Wolf *et al.*, "An Integrated Family of Amino Acid Sequence Analysis Programs," *Comput. Appl. Biosci.*, 4(1):187-91, 1988.

25 Wong and Neumann, "Electric field mediated gene transfer," *Biochim. Biophys. Res. Commun.*, 107(2):584-587, 1982.

Yamada *et al.*, *Plant Cell Rep.*, 4:85, 1986.

Yang *et al.*, *Proc. Natl. Acad. Sci. USA*, 87:4144-4148, 1990.

30 Yin, Chen, Beachy, "Promoter elements required for phloem-specific gene expression from the RTBV promoter in rice," *Plant J.*, 12(5):1179-1188, 1997a.

Yin, Zhu, Dai, Lamb, Beachy, "RF2a, a bZIP transcriptional activator of the phloem-specific rice tungro bacilliform virus promoter, functions in vascular development," *EMBO J.*, 16(17):5247-5259, 1997b.

Zhou *et al.*, "Introduction of exogenous DNA into cotton embryos," *Methods Enzymol.*,

5 101:433-481, 1983.

All of the compositions and methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been described in terms of preferred 10 embodiments, it will be apparent to those of skill in the art that variations may be applied to the composition, methods and in the steps or in the sequence of steps of the method described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents which are both chemically and 15 physiologically related may be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention as defined by the appended claims. Accordingly, the exclusive rights sought to be patented are:

**CLAIMS:**

1. A composition comprising at least a first isolated peptide or polypeptide comprising at least a first isolated coding region that comprises an amino acid sequence that is at least about 90% identical to the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.  
5
- 10 2. The composition according to claim 1, wherein said at least a first isolated coding region comprises an amino acid sequence that is at least about 92% identical to the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.
- 15 3. The composition according to claim 1 or claim 2, wherein said at least a first isolated coding region comprises an amino acid sequence that is at least about 94% identical to the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.
- 20 4. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises an amino acid sequence that is at least about 96% identical to the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.
- 25 5. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises an amino acid sequence that is at least about 98% identical to the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.

6. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises a sequence of at least about 27 contiguous amino acids from SEQ ID NO:2 or SEQ ID NO:4.

5

7. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises a sequence of at least about 30 contiguous amino acids from SEQ ID NO:2 or SEQ ID NO:4.

10

8. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises a sequence of at least about 35 contiguous amino acids from SEQ ID NO:2 or SEQ ID NO:4.

15

9. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises a sequence of at least about 40 contiguous amino acids from SEQ ID NO:2 or SEQ ID NO:4.

20

10. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises a sequence of at least about 50 contiguous amino acids from SEQ ID NO:2 or SEQ ID NO:4.

25

11. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises a sequence of at least about 60 contiguous amino acids from SEQ ID NO:2 or SEQ ID NO:4.

30

12. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises a sequence of at least about 70 contiguous amino acids from SEQ ID NO:2 or SEQ ID NO:4.

5

13. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises a sequence of at least about 80 contiguous amino acids from SEQ ID NO:2 or SEQ ID NO:4.

10

14. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises a sequence of at least about 90 contiguous amino acids from SEQ ID NO:2 or SEQ ID NO:4.

15

15. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.

20

16. The composition according to any preceding claim, wherein said at least a first isolated coding region consists essentially of the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.

25

17. The composition according to any preceding claim, wherein said at least a first isolated coding region consists of the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4.

30

18. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises an amino acid sequence that is encoded by at least a first nucleic acid segment that comprises an at least 26 contiguous nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

5

19. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises an amino acid sequence that is encoded by at least a first nucleic acid segment that comprises an at least about 35 contiguous nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

10

20. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises an amino acid sequence that is encoded by at least a first nucleic acid segment that comprises an at least about 50 contiguous nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

15

21. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises an amino acid sequence that is encoded by at least a first nucleic acid segment that comprises an at least about 65 contiguous nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

20

25 22. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises an amino acid sequence that is encoded by at least a first nucleic acid segment that comprises an at least about 80 contiguous nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

30

23. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises an amino acid sequence that is encoded by at least a first nucleic acid segment that comprises an at least about 95 contiguous nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

5

24. The composition according to any preceding claim, wherein said at least a first isolated coding region comprises an amino acid sequence that is encoded by at least a first nucleic acid segment that comprises an at least about 110 contiguous nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:3.

10

25. A composition comprising at least a first hybridoma cell line that produces a monoclonal antibody having immunospecificity for a peptide or polypeptide as defined in any one of claims 1 to 24.

15

26. A composition comprising at least a first monoclonal antibody, or an antigen-binding fragment thereof, that has immunospecificity for a peptide or polypeptide as defined in any one of claims 1 to 24.

20

27. The composition according to claim 26, comprising a light chain variable region, a heavy-chain variable region, a Fab fragment, a F(ab)<sub>2</sub> fragment, an Fv fragment, an scFv fragment, or an antigen-binding fragment of said antibody.

25

28. A composition comprising: (a) at least a first isolated polynucleotide that comprises at least a first isolated nucleic acid segment that encodes (i) at least a first peptide or polypeptide as defined in any one of claims 1 to 24, or (ii) an antibody or antigen

30

binding fragment as defined in claim 26 or claim 27; or (b) at least a first isolated polynucleotide that comprises at least a first nucleic acid segment that comprises an at least 26 contiguous nucleotide sequence from SEQ ID NO:1 or SEQ ID NO:3.

5

29. The composition according to claim 28, comprising at least a first isolated polynucleotide that comprises at least a first nucleic acid segment that encodes (a) a peptide or polypeptide as defined in any one of claims 1 to 24, or (b) an antibody or antigen binding fragment as defined in claim 26 or claim 27.

10

30. The composition according to claim 28 or claim 29, comprising at least a first isolated polynucleotide comprising at least a first isolated nucleic acid segment that comprises at least 30 contiguous nucleotides from SEQ ID NO:1 or SEQ ID NO:3.

15

31. The composition according to any one of claims 28 to 30, comprising at least a first isolated polynucleotide comprising at least a first isolated nucleic acid segment that comprises at least 45 contiguous nucleotides from SEQ ID NO:1 or SEQ ID NO:3.

20

32. The composition according to any one of claims 28 to 31, comprising at least a first isolated polynucleotide comprising at least a first isolated nucleic acid segment that comprises at least 70 contiguous nucleotides from SEQ ID NO:1 or SEQ ID NO:3.

25

33. The composition according to any one of claims 28 to 32, comprising at least a first isolated polynucleotide comprising at least a first isolated nucleic acid segment that comprises at least 85 contiguous nucleotides from SEQ ID NO:1 or SEQ ID NO:3.

30

34. The composition according to any one of claims 28 to 33, comprising at least a first isolated polynucleotide comprising at least a first isolated nucleic acid segment that comprises at least 100 contiguous nucleotides from SEQ ID NO:1 or SEQ ID NO:3.

5

35. The composition according to any one of claims 28 to 34, comprising at least a first isolated polynucleotide comprising at least a first isolated nucleic acid segment that comprises at least 115 contiguous nucleotides from SEQ ID NO:1 or SEQ ID NO:3.

10

36. The composition according to any preceding claim, wherein said isolated coding region is from 27 to about 5,000 amino acids in length, or said nucleic acid segment is from 27 to about 15,000 nucleotides in length.

15

37. The composition according to any preceding claim, wherein said isolated coding region is from 35 to about 4,500 amino acids in length, or said nucleic acid segment is from 40 to about 13,000 nucleotides in length.

20

38. The composition according to any preceding claim, wherein said isolated coding region is from 55 to about 4,000 amino acids in length, or said nucleic acid segment is from 70 to about 11,000 nucleotides in length.

25

39. The composition according to any preceding claim, wherein said isolated coding region is from 75 to about 3,500 amino acids in length, or said nucleic acid segment is from 100 to about 9,000 nucleotides in length.

30

40. The composition according to any preceding claim, wherein said isolated coding region is from 95 to about 3,000 amino acids in length, or said nucleic acid segment is from 130 to about 7,000 nucleotides in length.

5

41. The composition according to any preceding claim, wherein said isolated coding region is from 115 to about 2,000 amino acids in length, or said nucleic acid segment is from 160 to about 5,000 nucleotides in length.

10

42. The composition according to claim 29 or claim 30, comprising at least a first isolated polynucleotide comprising at least a first isolated nucleic acid segment that consists essentially of an at least 30 contiguous nucleotide sequence from SEQ ID NO:1 or

15

SEQ ID NO:3.

20

43. The composition according to any one of claims 29, 30, or 42, comprising at least a first isolated polynucleotide comprising at least a first isolated nucleic acid segment that consists of an at least 30 contiguous nucleotide sequence from SEQ ID NO:1 or

SEQ ID NO:3.

25

44. The composition according to any one of claims 28 to 43, wherein said nucleic acid segment is operably positioned under the control of at least a first heterologous promoter.

45. The composition according to any one of claims 28 to 44, wherein said nucleic acid segment is operably positioned under the control of at least a first heterologous plant-expressible promoter.

5

46. The composition according to any one of claims 28 to 45, wherein said nucleic acid segment is operably positioned under the control of at least a first heterologous constitutive, inducible, or tissue-specific plant-expressible promoter.

10

47. The composition according to any one of claims 28 to 46, wherein said nucleic acid segment is operably positioned under the control of at least a first heterologous constitutive, inducible, or tissue-specific plant-expressible promoter selected from the group consisting of corn sucrose synthetase 1, corn alcohol dehydrogenase 1, corn light harvesting complex, corn heat shock protein, pea small subunit RuBP carboxylase, Ti plasmid mannopine synthase, Ti plasmid nopaline synthase, petunia chalcone isomerase, bean glycine rich protein 1, Potato patatin, lectin, CaMV 35S, ALS, ubiquitin, globulin 1, cruciferin, napin,  $\beta$ -conglycinin, phaseolin,  $\gamma$  zein, and the S-E9 small subunit RuBP carboxylase promoter.

15

48. The composition according to any one of claims 28 to 47, wherein said nucleic acid segment is comprised within a vector.

20

49. The composition according to any one of claims 28 to 48, wherein said nucleic acid segment is comprised within a plant artificial chromosome, or a cosmid, plasmid, baculoviral, or viral vector.

25

30

50. The composition according to any preceding claim, wherein said polypeptide or said polynucleotide is comprised within a recombinant host cell.

5 51. The composition according to any preceding claim, wherein said polypeptide or said polynucleotide is comprised within a bacterial, fungal, or plant host cell.

10 52. The composition according to any preceding claim, further comprising at least a first detection reagent.

15 53. A virus comprising a polypeptide as defined in any one of claims 1 to 24, or a polynucleotide as defined in any one of claims 28 to 52.

54. A recombinant host cell comprising a polypeptide as defined in any one of claims 1 to 24, or a polynucleotide as defined in any one of claims 28 to 52.

20 55. The recombinant host cell according to claim 54, wherein said host cell is a bacterial cell.

25 56. The recombinant host cell according to claim 54 or claim 55, wherein said host cell is an *Escherichia*, *Salmonella* or *Agrobacterium* cell.

30 57. The recombinant host cell according to claim 54, wherein said host cell is an eukaryotic cell.

58. The recombinant host cell according to claim 57, wherein said cell is a plant cell.

5

59. The recombinant host cell according to claim 57 or claim 58, wherein said cell is a monocotyledonous or a dicotyledonous plant cell.

10 60. A transgenic plant comprising:

(a) a polypeptide as defined in any one of claims 1 to 24;

(b) an antibody or an antigen binding fragment as defined in claim 26 or claim 27;

15

(c) a polynucleotide as defined in any one of claims 28 to 52;

(d) a virus as defined in claim 53; or

20 (e) a recombinant host cell as defined in any one of claims 54 to 59.

61. The transgenic plant according to claim 60, characterized as a grain, tree, legume, fiber, vegetable, fruit, berry, nut, citrus, grass, cactus, succulent, floral, or ornamental plant.

25

62. The transgenic plant according to claim 60 or claim 61, characterized as a corn, rice, millet, tobacco, alfalfa, soybean, bean, sorghum, pea, *Brassica*, safflower, potato, coconut, palm, pumpkin, squash, poppy, sesame, peanut, cocoa, coffee, tomato, flax,

canola, sunflower, cotton, kapok, wheat, oat, barley, walnut, pecan, almond, or rye plant.

5 63. A progeny or seed of any generation of the transgenic plant according to any one of claims 60 to 62.

10 64. A progeny of any generation of the seed according to claim 63.

65. A seed of any generation of the progeny according to claim 64.

15 66. A plant grown from the seed according to claim 65. \*

67. A kit comprising a composition according to any one of claims 1 to 52, and instructions for using said kit.

20

68. The kit according to claim 67, wherein said kit further comprises at least a first detection reagent.

25

69. A method for detecting a PEAMT- or a  $\Delta$ PEAMT-encoding polynucleotide in a sample, comprising the steps of:

30 (a) contacting a population of polynucleotides suspected of encoding a PEAMT or a  $\Delta$ PEAMT polypeptide with at least a first labeled polynucleotide as defined in any

one of claims 28 to 52, under conditions effective to allow hybridization of substantially complementary nucleic acids; and

(b) detecting the hybridized complementary nucleic acids so formed.

5

70. A method for detecting a PEAMT- or a  $\Delta$ PEAMT peptide or polypeptide in a sample, comprising the steps of:

10 (a) contacting a sample suspected of containing a PEAMT or a  $\Delta$ PEAMT peptide or polypeptide with at least a first labeled antibody or antigen binding fragment as defined in any one of claims 25 to 27, under conditions effective to allow the formation of immunocomplexes; and

15 (b) detecting the immunocomplexes so formed.

71. A method of increasing the amount of PEAMT or  $\Delta$ PEAMT polypeptide in a plant cell comprising, (a) providing to said plant cell a biologically effective amount of a polypeptide as defined in any one of claims 1 to 24; or (b) expressing in said plant cell a biologically effective amount of a polynucleotide as defined in any one of claims 28 to 52.

25 72. A method of altering the level of phosphodimethylethanolamine, phosphomono-methylethanolamine, phosphatidylcholine, phosphocholine, choline, glycine betaine, or choline-*O*-sulfate in a plant cell comprising, expressing in said plant cell a biologically effective amount of a polynucleotide as defined in any one of claims 28 to 52.

30

73. A method of increasing the lipid content of a plant seed comprising, expressing in a transgenic plant an amount of a polynucleotide as defined in any one of claims 28 to 52, effective to increase the lipid content in the seed of said plant relative to the lipid content of a seed from an untransformed plant.

5

74. A method of increasing the choline or phosphatidylcholine content of a plant seed comprising, (a) providing to said plant cell a biologically effective amount of a polypeptide as defined in any one of claims 1 to 24; or (b) expressing in a transgenic plant an amount of a polynucleotide as defined in any one of claims 28 to 52, effective to increase the choline or phosphatidylcholine content in the seed of said plant relative to the choline or phosphatidylcholine content of a seed from an untransformed plant.

10

15

75. A method of decreasing the ethanolamine or phosphoethanolamine content of a plant seed comprising, (a) providing to said plant cell a biologically effective amount of a polypeptide as defined in any one of claims 1 to 24; or (b) expressing in a transgenic plant an amount of a polynucleotide as defined in any one of claims 28 to 52, effective to decrease the ethanolamine or phosphoethanolamine content in the seed of said plant relative to the ethanolamine or phosphoethanolamine content of a seed from an untransformed plant.

20

25

76. A method for altering the biosynthesis of a compound selected from the group consisting of phosphodimethylethanolamine, phosphomonomethylethanolamine, phosphatidyl-choline, phosphocholine, choline, glycine betaine, and choline-*O*-sulfate in a plant, said method comprising the steps of expressing in said plant a selected nucleic acid segment that comprises a polynucleotide as defined in any one of claims 28 to 52, wherein said nucleic acid segment is operably linked to a promoter

30

that expresses said segment in said plant; and (b) growing said plant under conditions effective to modulate the biosynthesis of said compound in said plant.

5 77. The method of claim 76, wherein the biosynthesis of said compound in said transformed plant is elevated relative to that in an untransformed plant of the same species.

10 78. The method of claim 76, said method further comprising the steps of (c) growing said plant under conditions effective for obtaining seeds from said plant, and (d) collecting the seeds so produced by said plant.

15 79. The method of claim 78, said method further comprising the step of (e) transforming said plant cell with at least at least a second polynucleotide that encodes at least a second polypeptide involved in phosphatidylcholine biosynthesis, wherein said second polynucleotide is operably linked to at least a first promoter capable of expressing said second polynucleotide in said plant cell to produce said second polypeptide in said cell.

20

80. A method for increasing the lipid content of a plant seed, said method comprising the steps of (a) growing a transgenic plant according to any one of claims 60 to 62, under conditions effective to produce seed in said plant, and (b) obtaining the seed produced from said plant.

25

81. The method of claim 80, wherein said seed are corn, rice, millet, tobacco, alfalfa, soybean, bean, sorghum, pea, *Brassica*, sugar beets, safflower, potato, tomato, flax, canola, sunflower, cotton, kapok, wheat, oat, barley, or rye seed.

5

82. A method for improving the osmotic stress tolerance of a plant, said method comprising the steps of (a) transforming said plant with a first polynucleotide as defined in any one of claims 28 to 52, and a second polynucleotide comprising a second nucleic acid segment that encodes a choline monooxygenase, choline oxidase, or choline dehydrogenase polypeptide, each of said nucleic acid segments operably linked to at least a first promoter that expresses said nucleic acid segments in said plant to produce the encoded polypeptides; and (b) growing the plant so transformed under conditions effective to improve the osmotic stress tolerance of said plant.

10

83. The method of claim 82, wherein the level of glycine betaine or choline-*O*-sulfate in said transformed plant is elevated relative to the level of glycine betaine or choline-*O*-sulfate in an untransformed plant of the same species.

15

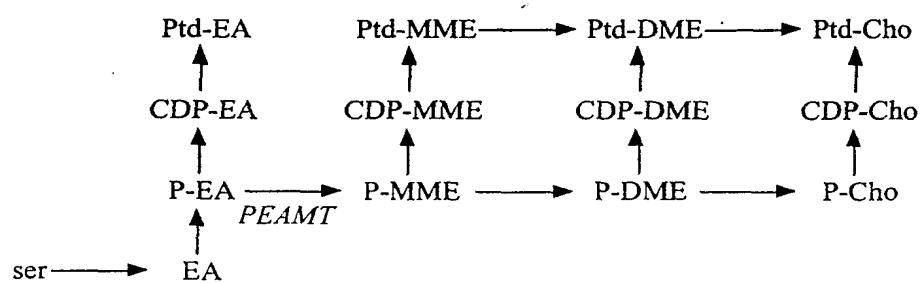
84. The method of claim 82, further comprising the step of providing to the cells of said transformed plant ethanolamine or phosphoethanolamine in an amount effective to increase the level of phosphocholine in the cells of said plant.

20

85. A method for increasing the cryoprotectant properties of a plant, said method comprising the steps of (a) transforming said plant with a first polynucleotide as defined in any one of claims 28 to 52, and a second polynucleotide comprising a nucleic acid segment that encodes a choline monooxygenase, choline oxidase, or choline dehydrogenase polypeptide, each of said nucleic acid segments operably

30

linked to a promoter that expresses said segments in said plant; and (b) growing the plant so transformed under conditions effective to increase the cryoprotectant properties of said plant.

**FIG. 1A**

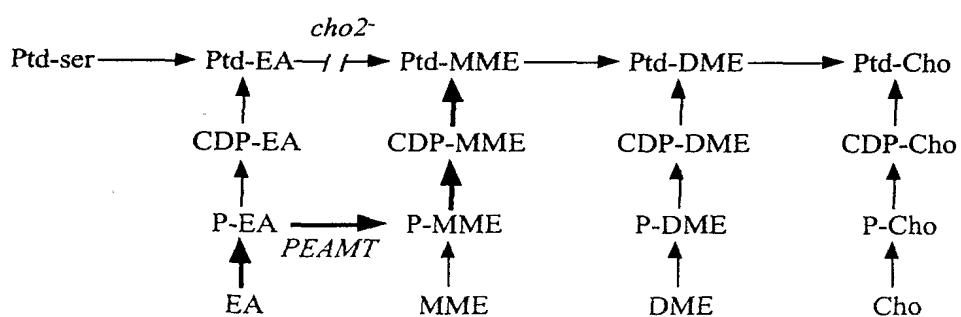
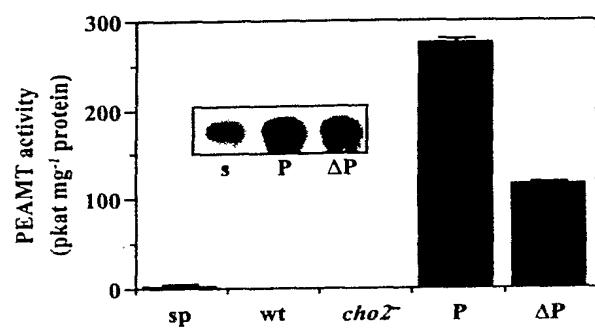
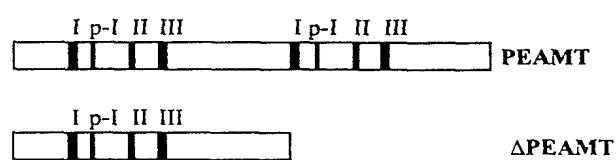


FIG. 1B

3/10

**FIG. 2A**



**FIG. 2B**

So 1 M A A S A M G V L Q E R E K V F K K Y W I E H S V D L T V E A M M L D S Q A S D L D K V E R P E V L S Y L P P Y E G K S V L E L G A G I G R F T G  
 At 1 M A A S F M E R E K T R D Y Q K N Y W I E H S A D L T V E A M M L D S R A S D L D K V E R P E V L S Y L P P Y E G K S V L E L G A G I G R F T G

I

So 73 E L A Q K A S Q V I A L D F I E S V I K K N E S I N G H Y K N V K F M C A D V T S P S L N I S P N S V E I F S N W L L M Y L S D K E V E R E L V  
 At 73 E L A Q K A G E L I A L D F I D N V I K K N E S I N G H Y K N V K F M C A D V T S P D L K I T D G S Y L E I F S N W L L M Y L S D K E V E L L A

II

So 145 E R M I L K W L K P G G Y I F F R E S C F H Q S G D H K R K S N P T H Y R E P R F Y E K I F K E C H M Q D D S G N S I E L S V I G C K C I G A Y V  
 At 145 E R M V G N I K V G G Y I F F R E S C F H Q S G D S K R K S N P T H Y R E P R F Y E K V F Q E C Q T R E A X G N S I E L S V I G C K C I G A Y V

III

So 217 K S K K N Q N Q I S W L W Q K V D S E D D R G F O R F L D S S O Y K I F N S I L R Y E R V F G P G Y V S T G G L E T T K E F V S K L D L K P G Q K  
 At 217 K N K K N Q N Q I C W M W O K V U S E N D R G F O R F L D N V Q Y K S S G I L R Y E R V F G Q G Y V S T G G L E T T K E F V E K M N L K P G Q K

I

post-I

So 289 V L D V G C G I G G G D F Y M A E N Y D V E V V G I D L S I N M I S F A L E R S I G L K C A V E F E V A D C T K K D Y F I N S F D V I Y S R D T  
 At 289 V L D V G C G I G G G D F Y M A E K F D V H V V G I D L S V N M I S F A L E R A I G L S C S V E F E V A D C T T K H Y F D N S F D V I Y S R D T

II

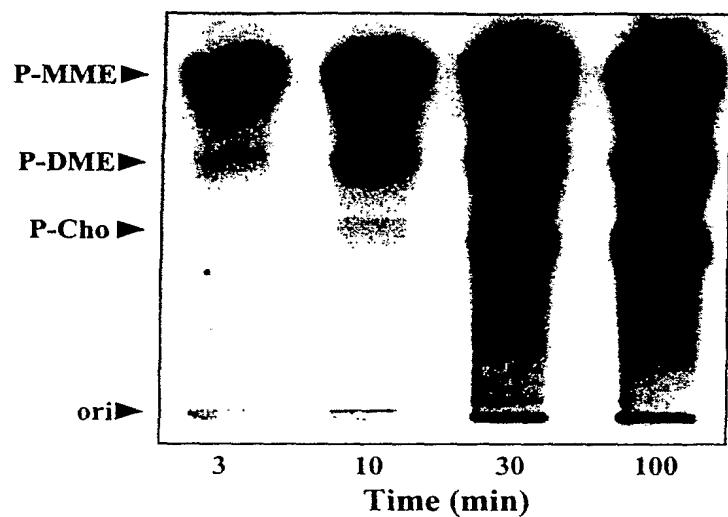
So 361 I L H I Q D K P A L F R S F H K W L K P G G K V L I S D Y C K S A G T P S A E F A A Y I R Q R G Y D L H D V K A Y G K M L K D A G F V E V I A E  
 At 361 I L H I Q D K P A L F R T F F K W L K P G G K V L I S D Y C R S P K T P S A E F S E Y I K Q R G Y D L H D V Q A Y G Q M L K D A G F T E V I A E

III

So 433 N R T D Q F I Q V L Q N E L D A L E Q E K D D F T D D F C E E D W N D I V D G W K A Y V R T T E G E C O N G L F I A K N M  
 At 433 D R T D Q F M Q V L K R E L D R V E K E K F I S D F E K E D Y D I V G G W K S P E R C A S D E J K M S L F I A K N N

FIG. 3

6/10

**FIG. 4A**

7/10

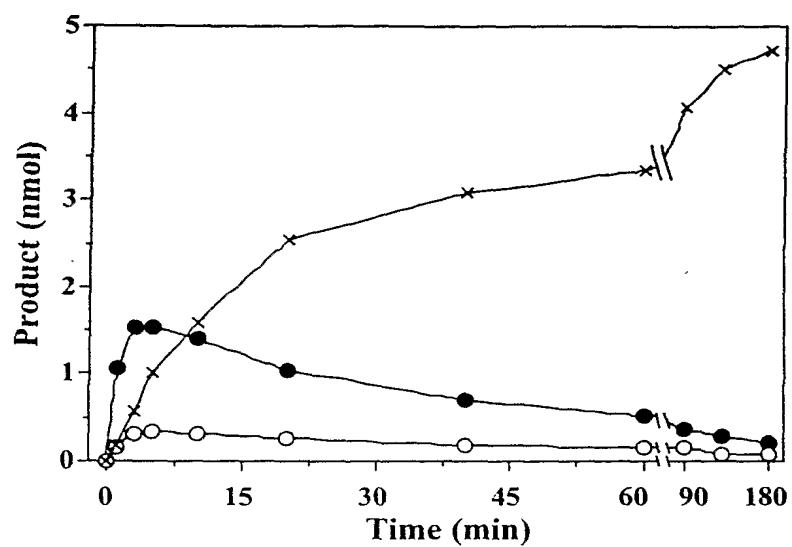
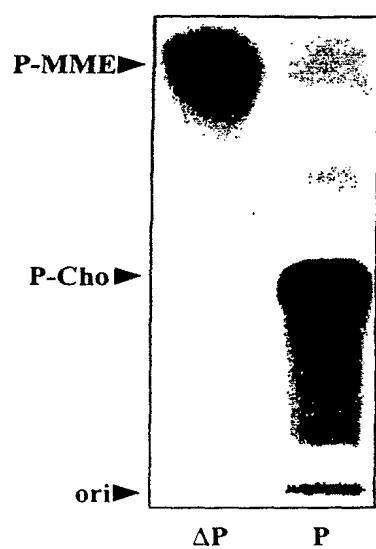
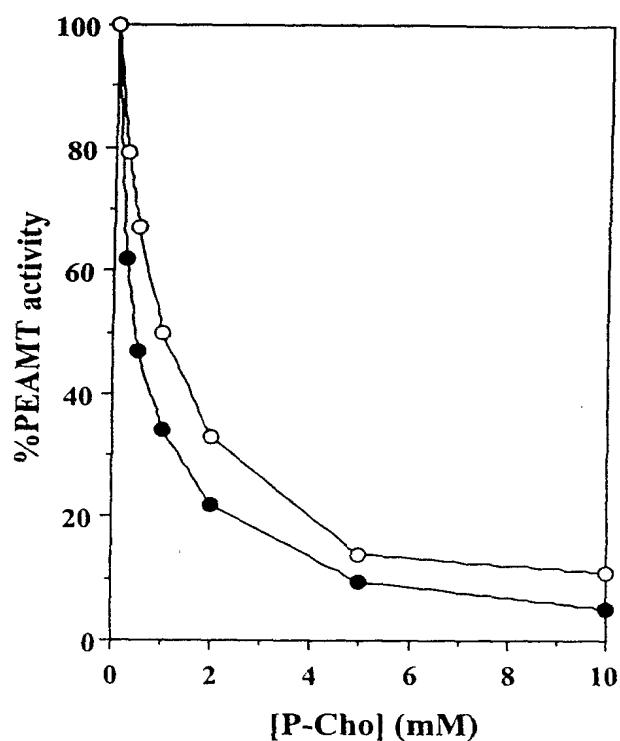


FIG. 4B



**FIG. 5A**

9/10

**FIG. 5B**

10/10

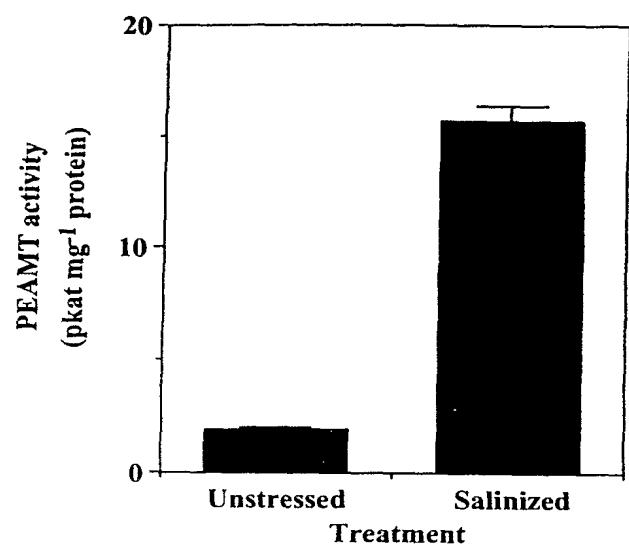


FIG. 6

## SEQUENCE LISTING

<110> Hanson, Andrew D.  
Nuccio, Michael L.  
Henry, Susan A.

<120> S-ADENOSYL-L-METHIONINE: PHOSPHOETHANOLAMINE N-METHYLTRANSFERASE COMPOSITIONS AND METHODS FOR MODULATING LIPID BIOSYNTHESIS IN PLANTS

<130> 4300.012710

<140> Unknown

<141> 2001-03-15

<150> 09/525, 885

<151> 2000-03-15

<160> 8

<170> PatentIn Ver. 2.1

<210> 1

<211> 2235

<212> DNA

<213> *Spinacia oleracea*

<400> 1

cattcatttg	aagcgtggaa	gtagtagttt	tgtggtagag	tgaatttgc	actcctactg	60
ctcatgcggc	agagaggcag	ggcttcgaac	cgtagatcca	ggacttttc	tcgttctcgc	120
attgccattg	aggtcacta	atactttaa	ctatctcc	cttttctt	cccacaaatt	180
ctcggtttc	acgcacatta	atctcaccta	ttttctagct	tcttcatttt	ctcaatcaat	240
ctctcggtt	attatggccg	cttcagctat	gggagtgtg	caagagagag	aggtgttcaa	300
gaaatactgg	atgaacact	ctgttattt	gactgttg	gctatgatgc	ttgattcaca	360
agcttcagat	cttgacaaag	tggaggcacc	tggaggtactt	tccatgcttc	cacccatata	420
aggaaagtct	gtcttagaac	tcgggtctgg	tattggctgt	tttactggtg	aattggccga	480
gaaagctagc	caggtcatcg	ctctggattt	cattgagagt	gttataaaga	agaatgaaag	540
cataaaatggg	cattacaaaa	atgttgaagtt	tatgtgtgt	gatgtgacat	ciccaagtct	600
caacatttca	ccaaattccg	tggatatcat	attctccaaat	tggctactca	tgtatcttc	660
tgtatgaagag	gttgagcgtc	tgttggaaag	gatgttgaaa	tggttgaagc	caggaggata	720
cattttcttc	agagaatctt	gttttcatca	atcaggagat	cacaagcgc	aaagcaatcc	780
aacccactac	cgtgaacacta	ggttctacac	caagatctc	aaagaatgcc	atatgcaaga	840
tgattctggg	aactcttatg	agctctccct	aattggctgc	aaatgtattt	gagcttatgt	900
caaaagcaag	aagaatcaga	accagataag	ctgggtatgg	cagaaaagtt	attcagagga	960
tgacaagggg	ttccagcgat	tcttggattt	tagtcaatac	aagtttaaca	gcatactgcg	1020
ttatgagcgt	gtatgggtc	ctgggttatgt	tagtaccgga	ggactcgaaa	caaccaagga	1080
gtttgtatca	aagcttgact	tgaaggctgg	ccagaaggc	ctagatgtgg	gttgtggcat	1140
aggtggaggt	gatttttaca	tggcagagaa	ctatgtatgtt	gaggttgttg	gaattgtatct	1200
ctccatttaat	atgatttctt	ttgcccattt	ggcgtcaattt	ggcctcaaat	gtgctgttga	1260
gtttgaggtg	gcagattgca	ccaagaaaga	ttaccctgaa	aactctttt	atgtcatctt	1320
cagccgttat	accattctgc	atattcaggaa	caaaccctgt	ttatttagat	ccttccacaa	1380
atggttgaaa	cctggaggca	aagtttttat	tagtactac	tgtaaagagt	ctggtacacc	1440
ttcagctgaa	tttgcgtcat	acatcaggca	gaggggatat	gatctccacg	atgtgaaggc	1500
atatggcaag	atgcttaaaag	atgctggatt	cgttgaggtt	attgttgaga	ataggactga	1560
ccagttcatt	caagttctgc	agaaggaaact	agatgtctt	gaacaggaga	aggatgactt	1620
cattgtatgt	ttctctgagg	aggattataa	cgacatagtt	gatgggttgg	aggccaagtt	1680
ggtgaggact	acagagggtt	agcaacaatg	gggtttgttc	attgccaaga	aatgtgaag	1740
aatgagctgg	tggaaagcagc	acggtgcott	ttttctagtat	tagtttatca	atgtattttc	1800
agttcatgga	ctgtatata	aaaatctacc	ataaagctgt	gagttgcaaa	ctgaaagatg	1860
atttcttata	gtcacttctg	aattagcaca	agcagtgaag	ttcgcataag	aaactgaagg	1920
gaactcatgg	agttgcagac	gaaatcatca	aaacggcaga	accactctc	tatataagaga	1980
tctagtggtt	aagttatgtt	ttttgtacat	tttccgttcc	aagttcactc	aatcttacca	2040

tcataatata accgctttta cttcttata tggtgattt aagtgcac tctttgttag 2100  
 taatgtgtat tagttgttg aaagtggac ttgcaacaca cttattcaca agtgtgtagg 2160  
 gaaatatgga ttttgttata gtatgtactg cacttagttg ttaaaaggat acttcctacg 2220  
 ttttcttctg ttgca 2235

<210> 2  
 <211> 494  
 <212> PRT  
 <213> Spinacia oleracea

<400> 2  
 Met Ala Ala Ser Ala Met Gly Val Leu Gln Glu Arg Glu Val Phe Lys  
 1 5 10 15  
 Lys Tyr Trp Ile Glu His Ser Val Asp Leu Thr Val Glu Ala Met Met  
 20 25 30  
 Leu Asp Ser Gln Ala Ser Asp Leu Asp Lys Val Glu Arg Pro Glu Val  
 35 40 45  
 Leu Ser Met Leu Pro Pro Tyr Glu Gly Lys Ser Val Leu Glu Leu Gly  
 50 55 60  
 Ala Gly Ile Gly Arg Phe Thr Gly Glu Leu Ala Glu Lys Ala Ser Gln  
 65 70 75 80  
 Val Ile Ala Leu Asp Phe Ile Glu Ser Val Ile Lys Lys Asn Glu Ser  
 85 90 95  
 Ile Asn Gly His Tyr Lys Asn Val Lys Phe Met Cys Ala Asp Val Thr  
 100 105 110  
 Ser Pro Ser Leu Asn Ile Ser Pro Asn Ser Val Asp Ile Ile Phe Ser  
 115 120 125  
 Asn Trp Leu Leu Met Tyr Leu Ser Asp Glu Val Glu Arg Leu Val  
 130 135 140  
 Glu Arg Met Leu Lys Trp Leu Lys Pro Gly Gly Tyr Ile Phe Phe Arg  
 145 150 155 160  
 Glu Ser Cys Phe His Gln Ser Gly Asp His Lys Arg Lys Ser Asn Pro  
 165 170 175  
 Thr His Tyr Arg Glu Pro Arg Phe Tyr Thr Lys Ile Phe Lys Glu Cys  
 180 185 190  
 His Met Gln Asp Asp Ser Gly Asn Ser Tyr Glu Leu Ser Leu Ile Gly  
 195 200 205  
 Cys Lys Cys Ile Gly Ala Tyr Val Lys Ser Lys Lys Asn Gln Asn Gln  
 210 215 220  
 Ile Ser Trp Leu Trp Gln Lys Val Asp Ser Glu Asp Asp Lys Gly Phe  
 225 230 235 240  
 Gln Arg Phe Leu Asp Ser Ser Gln Tyr Lys Phe Asn Ser Ile Leu Arg  
 245 250 255  
 Tyr Glu Arg Val Phe Gly Pro Gly Tyr Val Ser Thr Gly Gly Leu Glu  
 260 265 270

Thr Thr Lys Glu Phe Val Ser Lys Leu Asp Leu Lys Pro Gly Gln Lys  
 275 280 285  
 Val Leu Asp Val Gly Cys Gly Ile Gly Gly Asp Phe Tyr Met Ala  
 290 295 300  
 Glu Asn Tyr Asp Val Glu Val Val Gly Ile Asp Leu Ser Ile Asn Met  
 305 310 315 320  
 Ile Ser Phe Ala Leu Glu Arg Ser Ile Gly Leu Lys Cys Ala Val Glu  
 325 330 335  
 Phe Glu Val Ala Asp Cys Thr Lys Lys Asp Tyr Pro Glu Asn Ser Phe  
 340 345 350  
 Asp Val Ile Tyr Ser Arg Asp Thr Ile Leu His Ile Gln Asp Lys Pro  
 355 360 365  
 Ala Leu Phe Arg Ser Phe His Lys Trp Leu Lys Pro Gly Gly Lys Val  
 370 375 380  
 Leu Ile Ser Asp Tyr Cys Lys Ser Ala Gly Thr Pro Ser Ala Glu Phe  
 385 390 395 400  
 Ala Ala Tyr Ile Arg Gln Arg Gly Tyr Asp Leu His Asp Val Lys Ala  
 405 410 415  
 Tyr Gly Lys Met Leu Lys Asp Ala Gly Phe Val Glu Val Ile Ala Glu  
 420 425 430  
 Asn Arg Thr Asp Gln Phe Ile Gln Val Leu Gln Lys Glu Leu Asp Ala  
 435 440 445  
 Leu Glu Gln Glu Lys Asp Asp Phe Ile Asp Asp Phe Ser Glu Glu Asp  
 450 455 460  
 Tyr Asn Asp Ile Val Asp Gly Trp Lys Ala Lys Leu Val Arg Thr Thr  
 465 470 475 480  
 Glu Gly Glu Gln Gln Trp Gly Leu Phe Ile Ala Lys Lys Met  
 485 490

<210> 3  
 <211> 1120  
 <212> DNA  
 <213> Spinacia oleracea

<400> 3  
 cattcatttg aagcgtggaa gtagtagttt tgggttagag tgaatttgat actcctactg 60  
 ctcatgcggc agagaggcag ggcttcgaac cgtagatcca ggacttttc tcgttctcgc 120  
 attgccattt agggtcacta atactttaa ctatctcctt ctttttctt cccacaattt 180  
 ctgcgttttc acgcacatta atctcaccta ttttctagct tcttcatttt ctcaatcaat 240  
 ctctcgtgtt attatggccg cttcagctat gggagtgttg caagagagag aggtgttcaa 300  
 gaaaatactgg attgaacact ctgttgattt gactgtttag gctatgtatgc ttgattcaca 360  
 agtttcagat cttgacaaag tggagcgacc tgaggtactt tccatgcttc caccttatga 420  
 agggaaagtct gtcttagaac tcggtgctgg tattggtcgt tttactggtg aattggccga 480  
 gaaaagctagc caggtcatcg ctctggattt cattgagagt gttataaaga agaatgaaag 540  
 cattaaatggg cattacaaaa atgtgaagtt tatgtgtgct gatgtgacat ctccaagtct 600  
 caacatttca ccaaattccg tggatatcat attctccaat tggctactca tgtatcttc 660  
 tggatgaagag gttgagcgtc tggttgaaag gatgttgaaa tgggtgaagc caggaggata 720

cattttcttc agagaatctt gttttcatca atcaggagat cacaagcgca aaagcaatcc 780  
 aacccactac cgtgaaccta ggttctacac caagatctc aaagaatgcc atatgcaaga 840  
 tgattctggg aactcctatg agctctccct aattggctgc aaatgtattg gagcttatgt 900  
 caaaagcaag aagaatcaga accagataag ctggttatgg cagaaagttg attcagagga 960  
 tgacaagggg ttccagcgt tcttggattc tagtcaatac aagtttaaca gcatactg 1020  
 ttatgagcgt gtatttggtc ctggttatgt tagtaccgaa ggactcgaaa caaccaagga 1080  
 gtttgtatca aagcttgact tgaaggctgg gatccccggg 1120

<210> 4  
 <211> 289  
 <212> PRT  
 <213> Spinacia oleracea

<400> 4  
 Met Ala Ala Ser Ala Met Gly Val Leu Gln Glu Arg Glu Val Phe Lys  
 1 5 10 15  
 Lys Tyr Trp Ile Glu His Ser Val Asp Leu Thr Val Glu Ala Met Met  
 20 25 30  
 Leu Asp Ser Gln Ala Ser Asp Leu Asp Lys Val Glu Arg Pro Glu Val  
 35 40 45  
 Leu Ser Met Leu Pro Pro Tyr Glu Gly Lys Ser Val Leu Glu Leu Gly  
 50 55 60  
 Ala Gly Ile Gly Arg Phe Thr Gly Glu Leu Ala Glu Lys Ala Ser Gln  
 65 70 75 80  
 Val Ile Ala Leu Asp Phe Ile Glu Ser Val Ile Lys Lys Asn Glu Ser  
 85 90 95  
 Ile Asn Gly His Tyr Lys Asn Val Lys Phe Met Cys Ala Asp Val Thr  
 100 105 110  
 Ser Pro Ser Leu Asn Ile Ser Pro Asn Ser Val Asp Ile Ile Phe Ser  
 115 120 125  
 Asn Trp Leu Leu Met Tyr Leu Ser Asp Glu Val Glu Arg Leu Val  
 130 135 140  
 Glu Arg Met Leu Lys Trp Leu Lys Pro Gly Gly Tyr Ile Phe Phe Arg  
 145 150 155 160  
 Glu Ser Cys Phe His Gln Ser Gly Asp His Lys Arg Lys Ser Asn Pro  
 165 170 175  
 Thr His Tyr Arg Glu Pro Arg Phe Tyr Thr Lys Ile Phe Lys Glu Cys  
 180 185 190  
 His Met Gln Asp Asp Ser Gly Asn Ser Tyr Glu Leu Ser Leu Ile Gly  
 195 200 205  
 Cys Lys Cys Ile Gly Ala Tyr Val Lys Ser Lys Lys Asn Gln Asn Gln  
 210 215 220  
 Ile Ser Trp Leu Trp Gln Lys Val Asp Ser Glu Asp Asp Lys Gly Phe  
 225 230 235 240  
 Gln Arg Phe Leu Asp Ser Ser Gln Tyr Lys Phe Asn Ser Ile Leu Arg  
 245 250 255

Tyr Glu Arg Val Phe Gly Pro Gly Tyr Val Ser Thr Gly Gly Leu Glu  
260 265 270

Thr Thr Lys Glu Phe Val Ser Lys Leu Asp Leu Lys Pro Gly Ile Pro  
275 280 285

Gly

<210> 5  
<211> 11  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: SYNTHETIC  
OLIGONUCLEOTIDE

<400> 5  
ctcgagatct g 11

<210> 6  
<211> 15  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: SYNTHETIC  
OLIGONUCLEOTIDE

<400> 6  
tcgacagatc tcgag 15

<210> 7  
<211> 9  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: SYNTHETIC  
OLIGONUCLEOTIDE

<400> 7  
ctcgtgcac 9

<210> 8  
<211> 13  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: SYNTHETIC  
OLIGONUCLEOTIDE

<400> 8  
gatctggcac gag 13